

Preparing a 96-well plate for HHL assay

1. Set-up

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

Column1: 180µl LB-medium + 20µl H₂O
Column2: 180µl cells + 20µl H₂O
Column3: 200µl cells in 0.01nM HHL
Column4: 200µl cells in 0.1nM HHL
Column5: 200µl cells in 1nM HHL
Column6: 200µl cells in 10nM HHL
Column7: 200µl cells in 50nM HHL
Column8: 200µl cells in 100nM HHL
Column9: 200µl cells in 200nM HHL
Column10: 200µl cells in 500nM HHL
Column11: 200µl cells in 1000nM HHL
Column12: 200µl cells in 2000nM HHL

2. Pipetting scheme

2.1. Stock solutions

Final con. / µM	Stock used / µM	Vol. of stock / µl	Vol. of water / µl
150	15000	12	1188
15	150	120	1080
1.5	15	120	1080
0.15	1.5	120	1080
0.015	0.15	120	1080

2.2. 94-well plate

The volumes below are sufficient for pipetting four well plates at a time.

Final con. / nM	Stock used / µM	Vol. of stock / µl	Vol. of water / µl
0.01	0.015	4.8	715.2
0.1	0.015	48	672
1	0.15	48	672
10	0.15	480	240
50	1.5	240	480
100	1.5	480	240
200	15	96	624
500	15	240	480
1000	15	480	240
2000	150	96	624

Pipet 20µl of the above concentrations into the corresponding wells and add 180µl of cell suspension right before the measurement of fluorescence.

3. Tecan plate reader

Start the machine 20 minutes in advance and heat the chamber to 37°C. Put in the plate and start the measurement with the following setting:

- Measure OD at 612 nm
- Measure fluorescence with an excitation wavelength of 485nm and an emission wavelength of 485nm for both GFP and YFP
- Shake
- Repeat measurements every 5min for 3 hours

4. Data processing

- Normalize the data by dividing all fluorescence data by the corresponding optical density
- Subtract the obtained value of the reference column (column 2) from the calculated relative fluorescence of the wells containing HHL
- Plot the fluorescence data over the time