

# RNA Isolation Protocol

# RNA isolation

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## Introduction

In this protocol we expose how to isolate the IVT-RNA. We always used the measures for a 24W-plate. In this case, instead of TriZol we use Quiazol.

## Materials

### › Quiazol

- › Aspect: pink
- › Localization: Under the air flow cabin, right drawer, inside a carton box
- › Use in air flow cabin
- › Maintain at RT

### › Chloroform

- › **Always** use in an air flow cabin!

### › Isopropanol

- › Maintain at RT
- › We have a 50mL falcon (iGEM bench)

### › Ethanol 85%

- › Dilution is in our 50mL falcon (iGEM bench)
- › If not diluted--> Ethanol 100% is under the air flow cabin, right drawer, the biggest bottle

### › Glicoblue

- › For visualization of the RNA pellet!!!

## Procedure

### Obtaining of two-phases

1. **Lyse** cells by resuspending directly in **Quiazol**: pipet up and down until homogeneized
2. **Lyse** cells by resuspending directly in **Trizol**, pipet up and down until homogenized
  - Well of a 6W plate – 1ml Trizol
  - Well of a 24W plate – 0.5ml Trizol
3. For a well of a **24W plate**:
  - Add 0.1 ml chloroform (or chloroform-isoamyl alcohol) per 0.5 ml TRIzol Reagent initially used. Cap tubes and shake vigorously for 15 seconds. Do not vortex.
4. **Centrifuge** samples at no more than 12,000 x g for 10 minutes at 2–8°C.

5. **Examine phasing.** Clear, the aqueous phase should be the entire atop Phase Lock Gel. The phenol-chloroform phase and cloudy interphase should be below the Phase Lock Gel layer. If this is not the case, add another 0.1 ml chloroform (or chloroform-isoamyl alcohol) per 0.5 ml TRIzol Reagent used initially and shake vigorously. Repeat centrifugation and re-examine phasing.
6. **Transfer** aqueous phase containing RNA to a fresh tube
7. **Precipitate** RNA with isopropanol: use 400 µl isopropanol per ml Trizol initially used (1ul of Glicoblue will help to visualize the pellet)
8. **Incubate** > 20 min at -20 °C
9. **Centrifuge** > 20 min full speed at 4°C
10. **Discard** supernatant
11. **Wash** pellet by gently pipetting with 0.5 ml 85% ethanol for every 0.5 ml of TRIzol used. Mix by flicking and inverting tube or vortexing (flying pellet)
12. **Centrifuge** 5 min full speed at 4°C
13. Carefully **aspirant** supernatant.
14. **Repeat** ethanol 85% wash 2x
15. Air-dry RNA **pellet** for 5-10 minutes (be careful not to overdry)
16. **Dissolve** pellet in 40 ul of H<sub>2</sub>O at 30°C for 5 min with shaking (65°C if overdried)
17. **Pipette** up and down and quantify in nanodrop.

\*\* Note: the protocol expose in this pdf is not exactly the same as those set out in the bibliography part.  
We did it followinf a lab protocol. \*\*

## Bibliography

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