

# Single-temperature Double Digest

## Introduction

This is the Double Digest Protocol with Standard Restriction Enzymes, using a common reaction and same incubation temperature for both enzymes.

More information from NEB can be found [here](#).

Double Digests can be designed using [NEB's Double Digest Finder](#).

See the [NEBuffer Activity/Performance Chart with Restriction Enzymes](#) for the incubation temperatures.

[NEBcloner](#) will help guide your reaction buffer selection when setting up double digests.

## Materials

- › DNA **1 µg**
- › [NEBuffer](#)
  - › 1X
- › [NEB Restriction Enzymes](#)
- › Deionized Water

## Procedure

### Single Temperature DD Reaction

- ✓ 1. Set up the following reaction (total reaction volume 50 µl).

Table2		
	A	B
1		Reagent Volumes (µl)
2	Buffer (10x)	5
3	DNA *	Input Volume for ng
4	Restriction Enzyme #1 **	1
5	Restriction Enzyme #2 **	1
6	Deionized Water (µl)	48
7	Total Volume (µl)	50

\* Recommended maximum of 1 µg of substrate per 10 units of enzyme.

\*\* Restriction Enzymes should be added to the mixture last.

- ✓ 2. Mix components by pipetting the reaction mixture up and down, or by "flicking" the reaction tube.
- ✓ 3. Quick ("touch") spin-down in a microcentrifuge. Do not vortex the reaction.
- ✓ 4. Incubate for 1 hour at the enzyme-specific appropriate temperature.

01:00:00



Can be decreased to 5-15 minutes by using a [Time-Saver™ Qualified Restriction Enzyme](#)

See the [NEBuffer Activity/Performance Chart with Restriction Enzymes](#) for the incubation temperatures.