iGEM Thessaloniki - UParis **Collaboration Report**

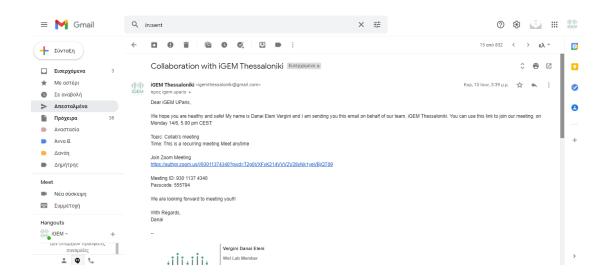


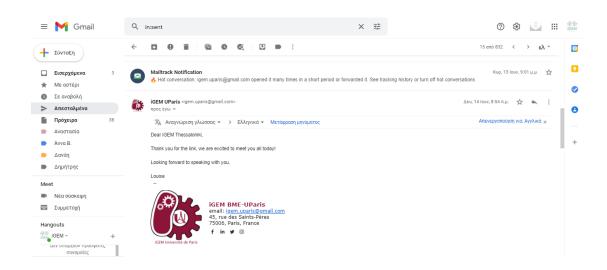
iGEM Thessaloniki and UParis 2021



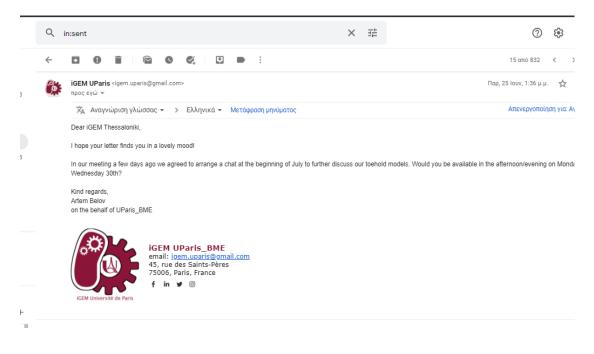
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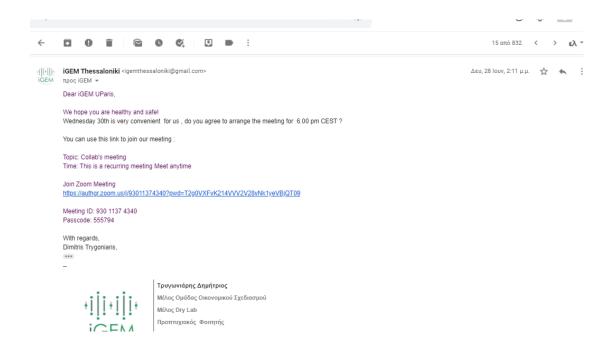
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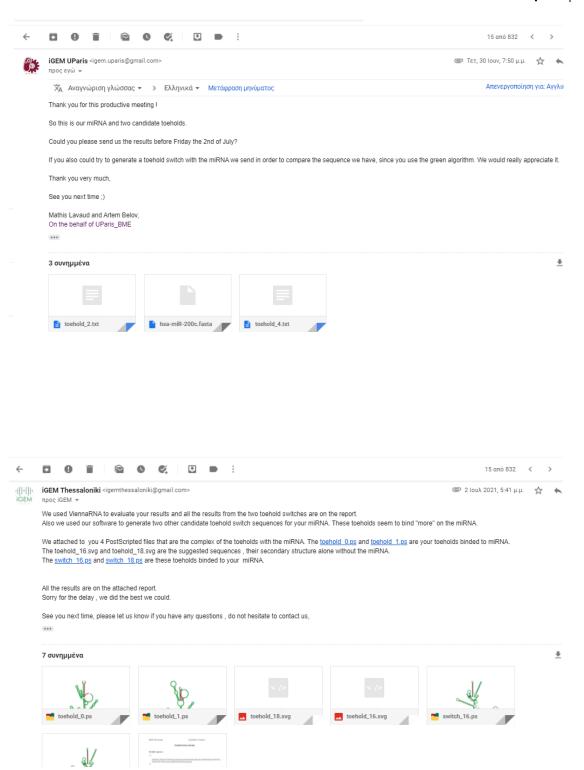


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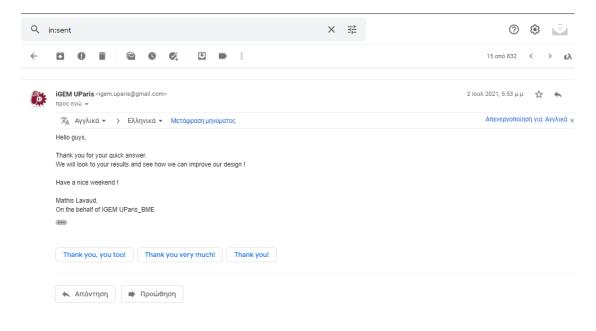


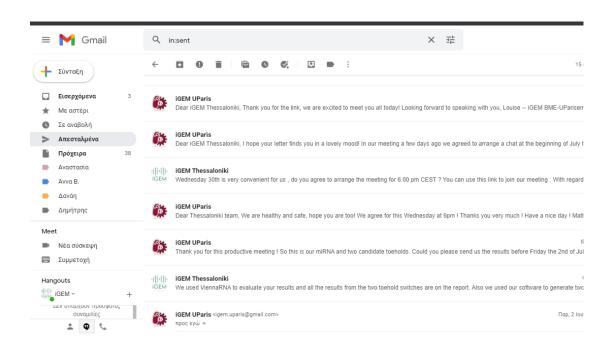


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iGEM Thessaloniki's first report

Toehold Switch evaluation:

Toehold sequences:

- 1. GGGAUCAUUACCCGGCAAAAAAAAAAAAAAAAAUACAGAAAC AGAGGAGAUAUUUUAUGUUUUUUTGCCACCUGGCGGCAG CGCAAAAG
- 2. GGGCCAUCAUUACCCGGAAAAAAAAAAAAAAUACAGAAA CAGAGGAGAUAUUUUAUGUUUUUUCCGGACCUGGCGCA GCGCAAAAG

miRNA sequence:

UAAUACUGCCGGGUAAUGAUGGA

GFP sequence:

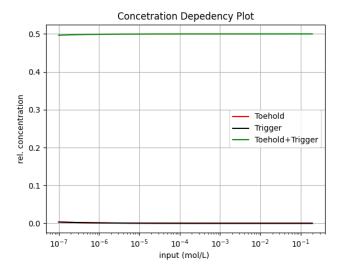
AUGCGUAAAGGAGAAGAACUUUUCACUGGAGUUGUCCCAAU UCUUGUUGAAUUAGAUGGUGAUGUUAAUGGGCACAAAUUUUC UGUCAGUGGAGAGGUGAAGGUGAUGCAACAUACGGAAAA CUUACCCUAAAUUUAUUUGCACUACUGGAAAACUACCUGUUC CGUGGCCAACACUUGUCACUACUUUCGGUUAUGGUGUUCAA UGCUUUGCGAGAUACCCAGAUCACAUGAAACAGCAUGACUU UUUCAAGAGUGCCAUGCCCGAAGGUUACGUACAGGAAAGA ACUAUAUUUUUCAAAGAUGACGGGAACUACAAGACACGUGC UGAAGUCAAGUUUGAAGGUGAUACCCUUGUUAAUAGAAUCG AGUUAAAAGGUAUUGAUUUUAAAGAAGAUGGAAACAUUCUUG GACACAAAUUGGAAUACAACUAUAACUCACACAAUGUAUAC AUCAUGGCAGACAAACAAAGAAUGGAAUCAAAGUUAACUU CAAAAUUAGACACAACAUUGAAGAUGGAAGCGUUCAACUAG CAGACCAUUAUCAACAAAAUACUCCGAUUGGCGAUGGCCC UGUCCUUUUACCAGACAACCAUUACCUGUCCACACAAUCUG CCCUUUCGAAAGAUCCCAACGAAAAGAGAGACCACAUGGU CCUUCUUGAGUUUGUAACCGCUGCUGGGAUUACACAUGGCA UGGAUGAACUAUACAAA

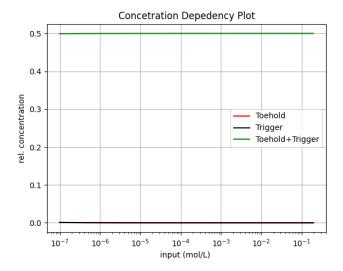
Energy of the binding:

1. Delta G binding: -16.71 kcal/mol, frequency of mfe structure in ensemble 7.95786e-14

2. Delta G binding: -14.63 kcal/mol, frequency of mfe structure in ensemble 2.57708e-14

Concentration Dependency Plots:





Same graphs for both of the toehold switches. Concentration Dependency Plots generated with the use of RNAcofold.exe of ViennaRNA software. The binding of the toehold and the miRNA seem to be independent from the input concentration of the two molecules. So the complex that will lead to GFP expression, only depends on initial concentration of toehold and miRNA.

Duplex output:

The dot-bracket notation refers to the complex that will be formed after the interaction of the two molecules. You can see that not all the miRNA binds upon toehold switch but instead the paired bases that will be formed are 14. The position of the molecules is denoted with the numbers of 3,18 and 6,21 in first case. It means that the paired base of miRNA will be in the position of 4,17 ->14 and from toehold switch 5,20->14.

Because the binding is continuous without other pairs, the perfect match of the two molecules should be considered upon design.

For the two sequences:

GGAUCAUUACCCGGCAAAAAAAAAAAAAAAAAUACAGAA GGAGAUAUUUUAUGUUUUUUTGCCACCUGGCGGCAGCGCAAAAG (1.0, 14)

AGGAGAUAUUUUAUGUUUUUUCCGGACCUGGCGCAGCGCAAAAG (1.0, 14)

You have 14 paired bases after interaction. The first argument of the tuple (1.0) refers that all the bases of the miRNA that was paired with toehold switch, are continuous and that favors the binding.

The last evaluation is on free energy of the RBS-linker. We need to minimize this energy and the best is to have 0 kcal/mol (linear structure of the RBS-Linker so the ribosome can initiate translation).

CAGAAACAGAGGAGAUAUUUUAUGUUUUUUUUGCCACCUGGC GGCAGCGCAAAAG

```
(((....(((((((((...)))))))))))....)))(((....))).....(-10.30)
.,((((((...((...)),.)))))) | {(((...{{|,...}}}.)),.... [-11.68]
```

frequency of mfe structure in ensemble 0.106941; ensemble diversity 16.88

CAGAAACAGAGGAGAUAUUUUAUGUUUUUUCCGGACCUGGC GGCAGCGCAAAAG

```
(((...(.(((((((((((...))))))))))...)))(((....)))..... [-12.47]
```

frequency of mfe structure in ensemble 0.549266; ensemble diversity 1.79

The first number is the mfe structure and the second number is the centroid structure. The energies are -10.30 kcal/mol and -11.68 kcal/mol for the first case. That means that after binding, the structure does not favor the linearity of RBS-linker.

The sequences that we predict to behave best are:

- GGGUCCAUCAUUACCCGGCAGUAUUGUAGUGUCAGAA ACAGAGGAGAACACACAUGAAUACUGCCAACCUGGCGG CAGCGCAAAAG
 - 1. GGGCCAUCAUUACCCGGCAGUAUUGUAGUGUGUCAGA AACAGAGGAGAACACACAUGAAUACUGCCAACCUGGCG GCAGCGCAAAAG

miRNA sequence:

UAAUACUGCCGGGUAAUGAUGGA

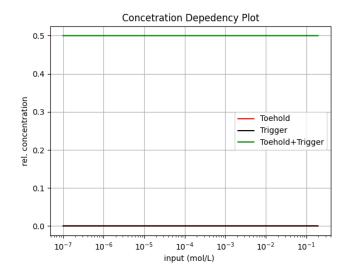
GFP sequence:

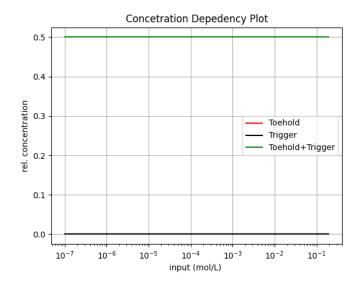
AUGCGUAAAGGAGAAGAACUUUUCACUGGAGUUGUCCCAAU UCUUGUUGAAUUAGAUGGUGAUGUUAAUGGGCACAAAUUUUC UGUCAGUGGAGAGGUGAAGGUGAUGCAACAUACGGAAAA CUUACCCUAAAUUUAUUUGCACUACUGGAAAACUACCUGUUC CGUGGCCAACACUUGUCACUACUUUCGGUUAUGGUGUUCAA UGCUUUGCGAGAUACCCAGAUCACAUGAAACAGCAUGACUU UUUCAAGAGUGCCAUGCCCGAAGGUUACGUACAGGAAAGA ACUAUAUUUUUCAAAGAUGACGGGAACUACAAGACACGUGC UGAAGUCAAGUUUGAAGGUGAUACCCUUGUUAAUAGAAUCG AGUUAAAAGGUAUUGAUUUUAAAGAAGAUGGAAACAUUCUUG GACACAAAUUGGAAUACAACUAUAACUCACACAAUGUAUAC AUCAUGGCAGACAAACAAAGAAUGGAAUCAAAGUUAACUU CAAAAUUAGACACAACAUUGAAGAUGGAAGCGUUCAACUAG CAGACCAUUAUCAACAAAAUACUCCGAUUGGCGAUGGCCC UGUCCUUUUACCAGACAACCAUUACCUGUCCACACAAUCUG CCCUUUCGAAAGAUCCCAACGAAAAGAGAGACCACAUGGU CCUUCUUGAGUUUGUAACCGCUGCUGGGAUUACACAUGGCA UGGAUGAACUAUACAAA

Energy of the binding:

- 1. Delta G binding -22.35 kcal/mol, frequency of mfe structure in ensemble 2.12499e-10
- 2. Delta G binding =-21.48kcal/mol, frequency of mfe structure in ensemble 1.55045e-10

Concentration Dependency Plots:





Duplex output:

The main difference here is that all the miRNA bind to the toehold switch since the 23 bases are paired upon interaction.

The binding in the second toehold is different. Here the binding involves 22/23 bases of miRNA (small difference)

Perfect matches:

The perfect matches depend upon the RNAduplex.exe output.

Delta G RBS-Linker:

1. CAGAAACAGAGGAGAACACACAUGAAUACUGCCAAC CUGGCGCAGCGCAAAAG

((((((()))))) { -8.00 d=6.09}	
frequency of mfe structure in ensemble 0.282723; e diversity 7.58	nsemble
Free energy = -8.00 kcal/mol	
CAGAAACAGAGGAGAACACACAUGAAUACUGC CUGGCGGCAGCGCAAAAG	CCAAC
(((((()))))) (-8.00)	
[-8.78]	
((((((()))))) { -8.00 d=6.09}	
frequency of mfe structure in ensemble 0.282723; e diversity 7.58	nsemble

iGEM Thessaloniki's second report

Toehold Switch evaluation:

Toehold Switch sequence generation second GGGCAUCAUUACCCGGCAGUAUAAAAAAAAAAAAAAACAGAAACAGAGG AGAUAUUUUAUGUAUACUGCCGACCUGGCGGCAGCGCAAAAG

miRNA-200 c :

UAAUACUGCCGGGUAAUGAUGGA

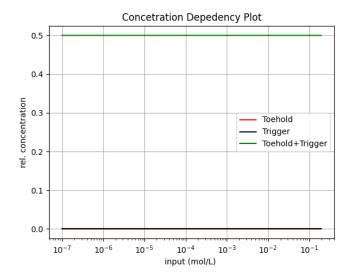
GFP:

AUGCGUAAAGGAGAACUUUUCACUGGAGUUGUCCCAAUUCUU GUUGAAUUAGAUGGUGAUGUUAAUGGGCACAAAUUUUCUGUCAGUG GAGAGGGUGAAGGUGAUGCAACAUACGGAAAACUUACCCUAAAU UUAUUUGCACUACUGGAAAACUACCUGUUCCGUGGCCAACACUUG UCACUACUUUCGGUUAUGGUGUUCAAUGCUUUGCGAGAUACCCAG AUCACAUGAAACAGCAUGACUUUUUCAAGAGUGCCAUGCCCGAAG GUUACGUACAGGAAAGAACUAUAUUUUUCAAAGAUGACGGGAACU ACAAGACACGUGCUGAAGUCAAGUUUGAAGGUGAUACCCUUGUUA AUAGAAUCGAGUUAAAAGGUAUUGAUUUUAAAGAAGAUGGAAACAU UCUUGGACACAAUUGGAAUACAACUAUAACUCACACAAUGUAUAC AUCAUGGCAGACAAACAAAGAAUGGAAUCAAAGUUAACUUCAAA AUUAGACACAUUGAAGAUGGAAGCGUUCAACUAGCAGACCAU UAUCAACAAAUACUCCGAUUGGCGAUGGCCCUGUCCUUUUACCA GACAACCAUUACCUGUCCACACAAUCUGCCCUUUCGAAAGAUCCC AACGAAAAGAGACCACAUGGUCCUUCUUGAGUUUGUAACCGCU GCUGGGAUUACACAUGGCAUGGAUGAACUAUACAAA

Energy of the binding:

delta G binding=-20.74 (better than before -16.71kcal/mol), frequency of mfe structure in ensemble 4.26997e-13

Concentration Dependency Plots:



Immediate binding upon interaction and the concentration of the complex is independent from the initial amount of miRNA and toehold switches molecules. The results are the same as the first generation of sequences.

Duplex output:

The part of the miRNA that binds to toehold switch starts from 4 position to 22 position. The binding includes the formation of 19 hydrogen bonds. This result is better than the previous one since the interaction of the first generation of toehold switches had only 14 base pairs upon interaction.

The perfect matches are 19 and the domain on interaction is predicted to be continuous. So the perfect matches for 19 pairs are 100% for this toehold switch sequence.

Energy of RBS-Linker:

CAGAAACAGAGGAGAUAUUUUAUGUAUACUGCCGACCUGGCGGC **AGCGCAAAAG**

•••••	(((((())))).	{-10.30 d	=5.43}		
frequency	of mfe	structure	in ensemble	0.148513;	ensemble	diversity
7.39						

The Gibbs free energy of the domain of RBS-Linker Structure after binding, is -11.20 kcal/mol and this deviate from linear behavior that we would like. The linear behavior is achieved through the 0 kcal/mol free energy value for this domain of the molecule.

To conclude the results of this molecule are predicted to be better with the software of ViennaRNA compared to the first generation of toehold switch sequences.