

Protocol for Sysbio plates and media

LB plates:

	1 L	4 L
Peptone from casein (Tryptone)	10 g	40 g
NaCl	10 g	40 g
Yeast Extract	5 g	20 g
Set pH to 7.0 with NaOH		
Agar agar	16 g	64 g
Autoclave		

LB + Amp 100 mg/L plates:

Follow protocol for LB plates, add ampicillin after autoclavation.

Ampicillin sodium salt: 100 mg/ml (1000X stock) dissolved in 50% EtOH/H₂O and sterile-filtered.

OBS! Handle ampicillin with care and avoid breathing in dust/fumes.

LB + Chl 25 mg/L plates:

Chloramphenicol: 25 mg/mL (1000x stock) dissolved in 100% EtOH and sterile filtered (store at -20°C). Final concentration to use 25 mg/L (=1mL/L) for both media and plates. Add after autoclavation.

Make stock:

Weigh 0.25 g of Chloramphenicol. Add 10 ml of 95% EtOH (70% may also be used), dissolve completely. Stock may be kept at -20°C for 1 year.

YPD plates:

	1 L	4 L
Peptone from meat	20 g	80 g
Yeast Extract	10 g	40 g
Agar agar	20 g	80 g
H ₂ O	0,8 L	3,5 L
Autoclave		
D(+)-Glucose	20 g	80 g
H ₂ O	0,2 L	0,5 L
Autoclave glucose separately		

YPD + G418 200 mg/L plates:

Follow protocol for YPD plates, add G418 after autoclavation.

G418 disulfate salt: 50mg/ml dissolved in H₂O and sterile-filtered (or Formedium G418-S solution 50 mg/ml sterile-filtered). Final concentration: 200 mg/L (=4ml/L).

OBS! Handle G418 with care and avoid breathing in dust/fumes.

YPD + G418 200 mg/L + nourseothricin 100 mg/L plates:

Follow protocol for YPD plates, add G418 and NAT after autoclavation, when temperature of agar mix is 50°C.

Add 4mL G418 50mg/ml. Final concentration: 200 mg/L (=4ml/L).

Add 1 mL nourseothricin 100 mg/uL.

SD dropout plates:

Solution 1:

	1 L	4 L
Agar agar (final conc. 2%)	20 g	80 g
H ₂ O	750 ml	3 L
Autoclave and keep at a room temperature of 60°C.		

Solution 2:

Complete supplement mix Drop out (select one of the below dropout aa mix)

-URA	0.77 g	3,08 g
-HIS	0.77 g	3,08 g
-URA -HIS	0.75 g	3 g
Yeast Nitrogen Base without amino acids	6.9 g	27,6 g
D(+)-Glucose	20 g	80 g
H ₂ O	250 ml	1 L

Set pH to 5.5 – 6.0

Let the solution dissolve and set pH. Gently heat Solution 2 in the microwave until the temperature is about 60°C. Stir until completely dissolved and filter-sterilize. Add solution 2 to the autoclaved agar-solution (solution 1) and mix thoroughly.

5-FOA plates:**600ml****4 L***Solution 1:*

Agar agar (final conc. 2%)

12 g

80 g

H₂O

255 ml

3 litre

Autoclave and keep at a temperature of 60°C.

Solution 2:

Yeast Nitrogen Base without amino acids

4.14 g

27,6 g

Complete supplement mix Drop out: -URA

462 mg

3,08 g

Uracil 99+%

30 mg

0,2 g

5-Fluoroorotic acid

600 mg

4 g

D(+)-Glucose

12 g

80 g

MQ H₂O

345 ml

1 litre

Gently heat Solution 2 in the microwave until the temperature is about 60°C. Stir until dissolved completely. Filter-sterilize. Add the autoclaved agar-solution (solution 1). Mix thoroughly, pH is not set.

OBS! Handle 5-FOA with care and avoid breathing in dust/fumes.

Chemicals used:**Brand****Product number:**

Peptone from casein

Merck

art nr 1.07213.1000

NaCl

Merck

art nr 1.06404.1000

Yeast Extract

Merck

art nr 1.11926.1000

Agar agar

Merck

art nr 1.01614.1000

Ampicillin sodium salt

AppliChem

art nr A0839.0025

Peptone from meat

Merck

art nr 1.07224.1000

D(+)-glucose

Merck

art nr 1.08342.9025

G-418 disulfate salt

Sigma Aldrich

art nr A1720

Complete supplement mix Drop out: -URA

Formedium

art nr DCS0169

Complete supplement mix Drop out: -HIS

Formedium

art nr DCS0079

Complete supplement mix Drop out: -HIS,-URA

Formedium

art nr DCS0529

Yeast Nitrogen Base without amino acids

Formedium

art nr CYN0402

Uracil 99+%

Alfa Aesar

art nr A15570

5-Fluoroorotic acid

Formedium

art nr 5FOA10

Document updates:

Name	Date	Update
Åsa Rensfeldt	2016-04-04	Addition of amounts for 4 L batches and safety reminders regarding antibiotics and 5-FOA.
Angelica Ardehed	2016-11-18	New article no and company for 5-FOA