

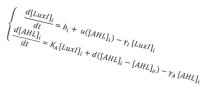


Synthetic Biology and Biosystems Control Lab Valencia UPV

Modeling: ODEs and Hill Functions Section 1: ODEs, Law of mass action and the central dogma by Alejandro Vignoni (alvig2@upv.es)

An iGEM Measurement Committee Webinar Week 2, June 23rd, 2020

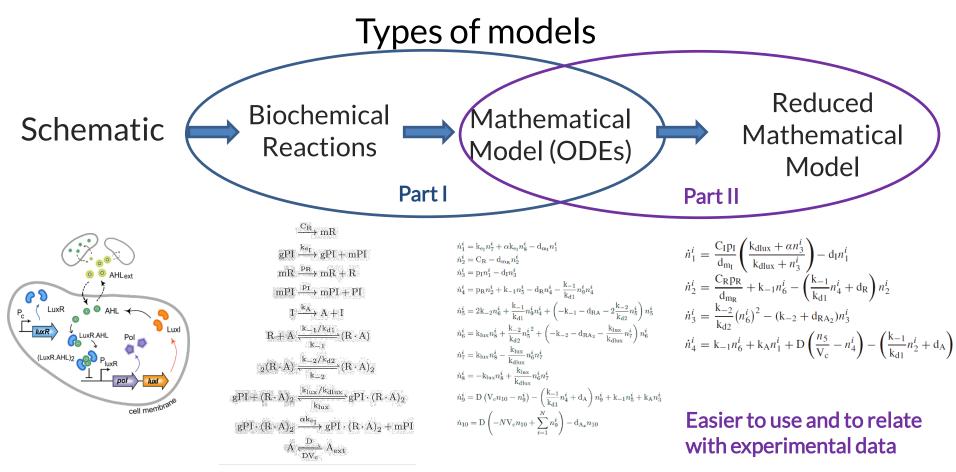




Today Webinar's Topics



- A Section 1: ODEs, the law of mass action, and the central dogma (15 min)
- Section 2: Derivation of a Hill function from the law of mass action (15 min)
- A Section 3: Hill function examples and intuitions: effects of parameters on activators, repressors, hybrid promoters, using a Matlab exploration package. (15min)
- ▲ Q&A (at the end of each 15 minutes block, total 15 min)



From Y. Boada (2018)

But what is an Ordinary Differential Equation (ODE)?

These are equations with variables and their derivatives

If we have any function (the typical one):

y = f(t) (y only depends on the variable t, but we could have $y = f(t, x_1, x_2, ..., x_n)$)

Do you remember the definition of the derivative of a function?

$$\dot{y} = \frac{df(t)}{dt} = \lim_{h \to 0} \frac{f(t+h) - f(t)}{h}$$

(we can have higher order derivatives $y'', y''', y^{(n)}$)

But they can be very challenging and difficult to solve!!

- D. Velarabel+ D. farabil - darab. (arab DVc (arabe) - D(crebi)-2ks (DNA) + 2 Ka (DNBancarobi) = - Kb. [ONA arec]. [aubi] + Ku [DNA anCarabi R6 JOHA ALGA

But if they're so complicated... how do we solve them?

We can solve differential equations in two ways:

A Analytically: solving for the unknown...

3a(y+G)2+(3y+G+4)4 a2C3 (y+A)32222414 39 b)2 (X+-

A **Numerically:** in an approximate way.

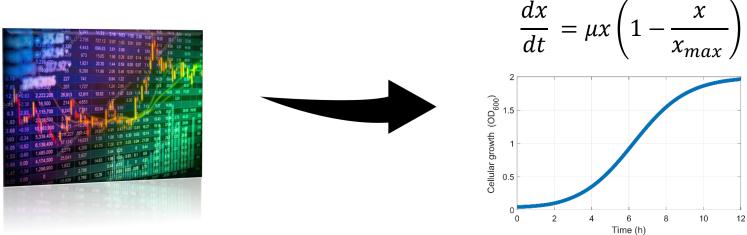


$$\dot{y} \simeq \frac{f(h+h) - f(x)}{h}$$

(with an h very small)

Why do we use them?

Differential equations describe biological behaviour, physical laws, human activities, and much more....



And the set of equations that describe a system, a phenomenon... is known as <u>ODE model</u>

Software for Ordinary Differential Equations (ODEs) solving

•<u>MATLAB</u>, a technical computing application (MATrix LABoratory)

FREE LICENSE WITH iGEM and the Measurement Committee has some software programed in MATLAB for flow cytometry and plate reader data analysis and calibration.

- •<u>Maxima</u>, an open-source <u>computer algebra system</u>.
- •<u>COPASI</u>, a free software package for the integration and analysis of ODEs.
- •<u>GNU Octave</u>, a high-level language, basically a open-source versión of MATLAB.
- •<u>Scilab</u>, an open source application for numerical computation.
- •<u>Maple</u>, a proprietary application for symbolic calculations.
- •<u>Mathematica</u>, a proprietary application primarily intended for symbolic calculations.
- •Julia (programming language), a high-level language primarily intended for numerical computations.
- •<u>SageMath</u>, an open-source application that uses a Python-like syntax with a wide range of capabilities spanning several branches of mathematics.
- •<u>SciPy</u>, a Python package that includes an ODE integration module.

•<u>GNU R</u>, an open source computational environment primarily intended for statistics, which includes packages for ODE solving.



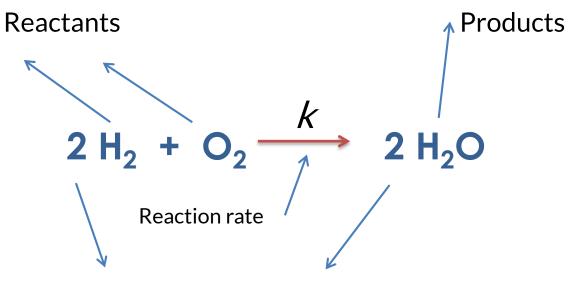


Let us begin this journey **Part I** from:

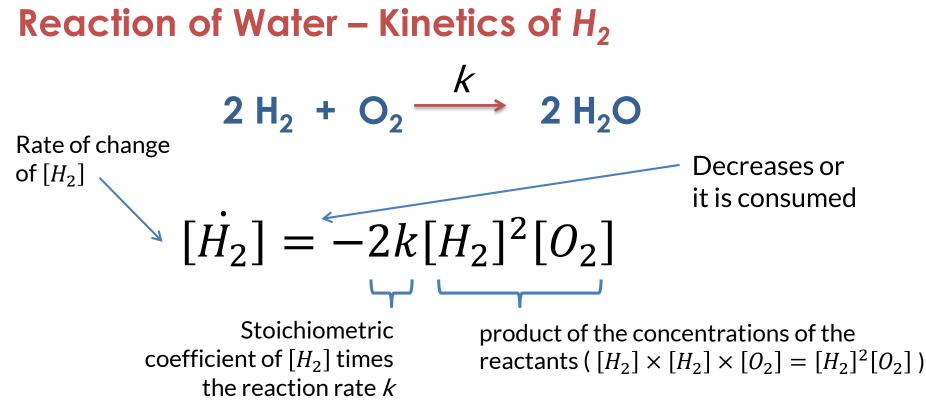


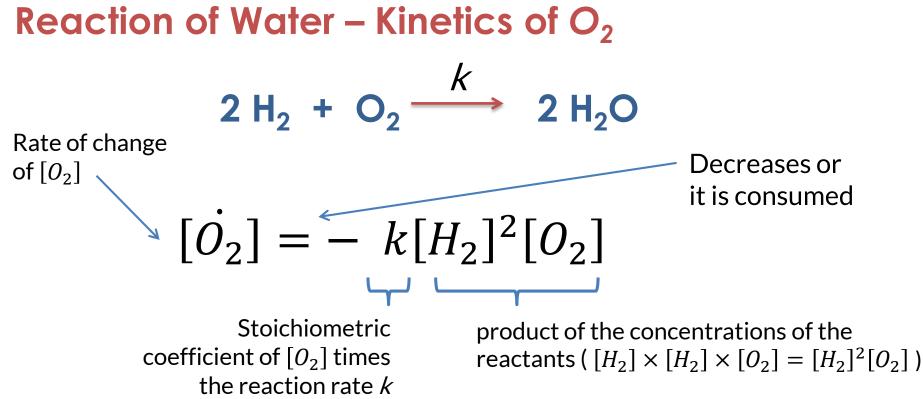
Reminder: Law of mass action and kinetic equations

Example: Reaction of Water



Stoichiometric coefficients





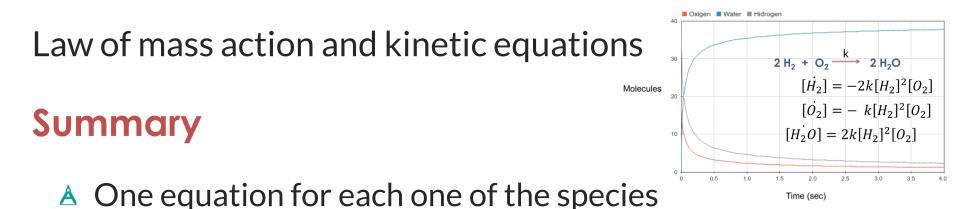
times the reaction rate k

Reaction of Water – Kinetics of
$$H_2O$$

 $2 H_2 + O_2 \xrightarrow{k} 2 H_2O$
Rate of change
of $[H_2O]$
 $[H_2O] = +2k[H_2]^2[O_2]$
Stoichiometric
coefficient of $[H_2O]$
product of the concentrations of the
reactants ($[H_2] \times [H_2] \times [O_2] = [H_2]^2[O_2]$)

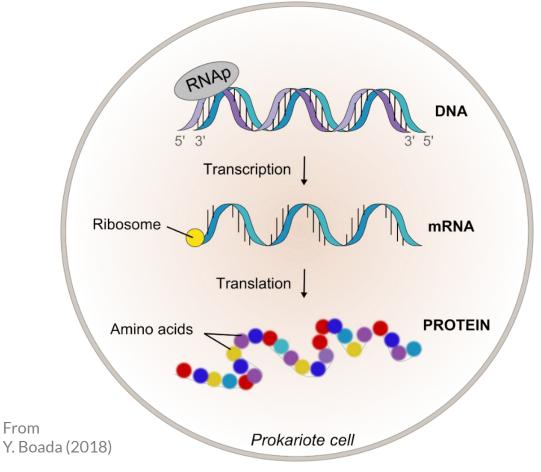
Reaction of Water – Kinetics $2 H_2 + O_2 \xrightarrow{k} 2 H_2O$ $[\dot{H}_2] = -2k[H_2]^2[O_2]$ $[\dot{O}_2] = -k[H_2]^2[O_2]$ $[H_2O] = 2k[H_2]^2[O_2]$





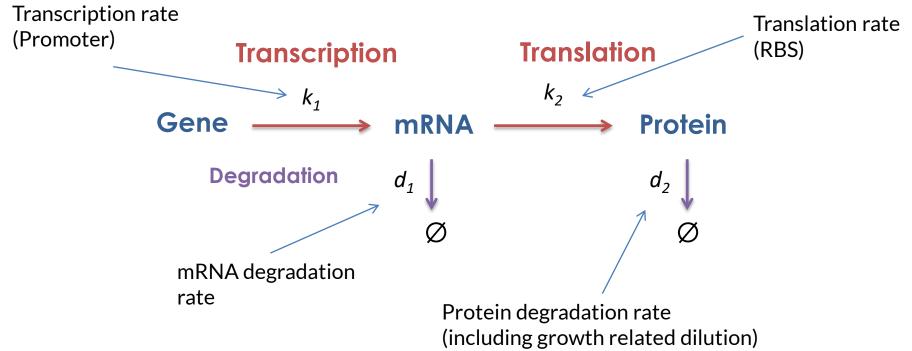
- A Rate of change of [A] (concentration of species A) is proportional to:
 - \land Stoichiometric coefficient of *A* time reaction rate *k*
 - A The product of the concentrations of the reactants

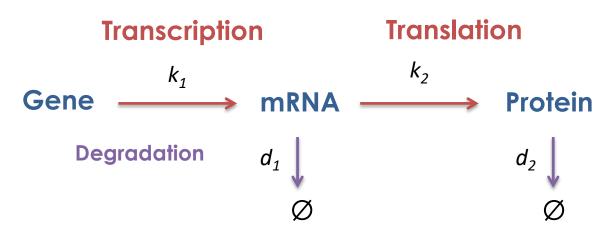
The central dogma of molecular biology

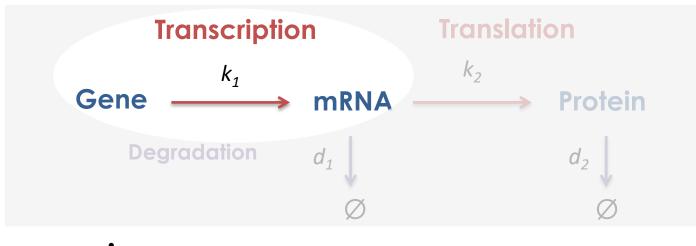


Transcription of DNA by RNA polymerase

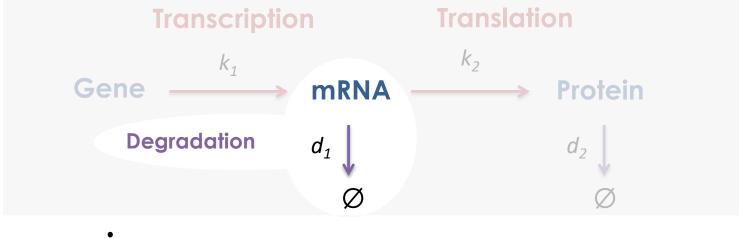
Translation of mRNA by Ribosomes



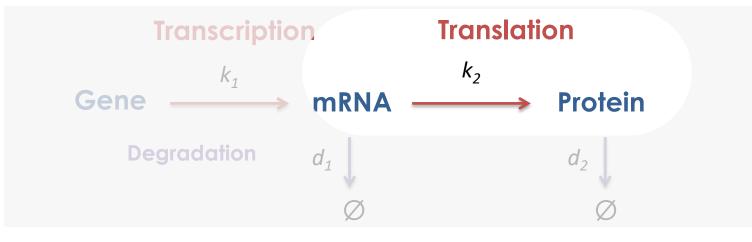




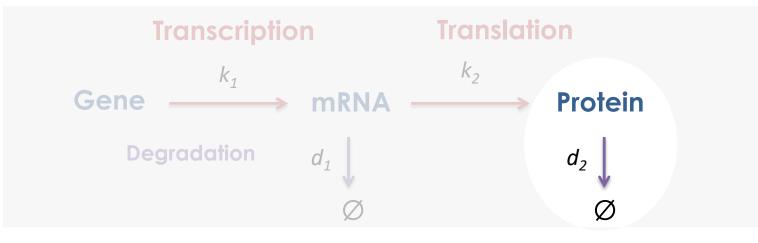
$[mRNA] = k_1[Gene]$



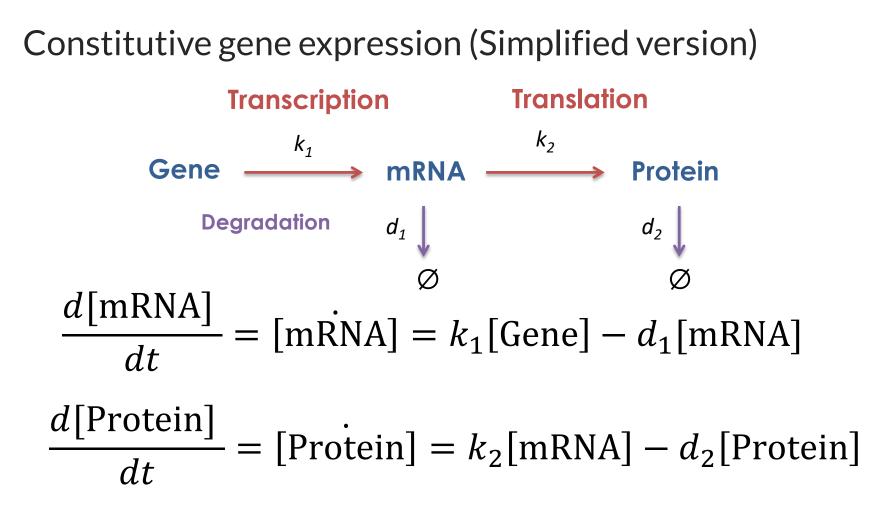
$[mRNA] = k_1[Gene] - d_1[mRNA]$



$[mRNA] = k_1[Gene] - d_1[mRNA]$. $[Protein] = k_2[mRNA]$



$[mRNA] = k_1[Gene] - d_1[mRNA]$ $\dot{Protein} = k_2[mRNA] - d_2[Protein]$



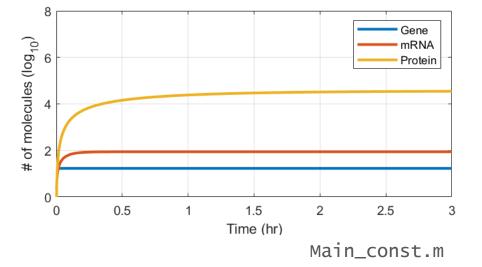
Constitutive gene expression - Simulation

ODE model

 $[mRNA] = k_1[Gene] - d_1[mRNA]$ $[Protein] = k_2[mRNA] - d_2[Protein]$

Function defines the ODE model

```
% Constitutive gene expression model.
% Updated 17/06/2020 Alejandro Vignoni
function [dxdt] = model_const(t,x,p)
%x1 = mRNA
dxdt(1,1) =p.CN*p.k1-p.d1*x(1);
%x2 = Protein
dxdt(2,1)=p.k2*x(1)-p.d2*x(2);
end
```







Constitutive gene expression - Simulation

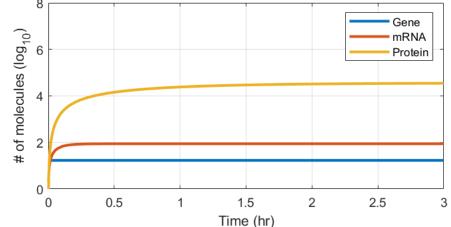
Parameters definition

%Parameters

p.CN = 17;	<pre>% plasmid number pACYC184 (17 copies/cell)</pre>
p.d1 = log(2)/3;	<pre>% mRNA degradation rate [1/min]</pre>
p.d2 = 0.02;	<pre>% degradation rate [1/min]</pre>
p.k2 = 8.23;	<pre>% translation rate [1/min]</pre>
p.k1 = 1.19;	<pre>% transcription rate [1/min]</pre>

Simulation configuration and execution

```
tfin = 60*3; %simulation final time
step = 0.1; %simulation step
tspan = 0:step:tfin-step;
% options for ode function
opti = odeset('AbsTol',1e-8,'RelTol',1e-6);
Init = [0 0]; %initial conditions
```



[t0,x0] = ode23t(@(t,x) model_const(t,x,p),tspan, Init, opti);

https://blogs.mathworks.com/cleve/2014/06/09/ordinary-differential-equations-stiffness/

Constitutive gene expression - Simulation



%Parameters

p.CN = 17;	<pre>% plasmid number pACYC184 (17 copies/ce</pre>	:11)
p.d1 = log(2)/3;	<pre>% mRNA degradation rate [1/min]</pre>	
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	8 —	

Simulation configuration and execution

tfin	=	60*3;	%simulation	final	time
sten	=	01.	*simulation	sten	

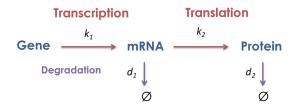


- Sponsor Webinar MathWorks: Modeling and Analysis of Synthetic Biology Systems with SimBiology and MATLAB
- **† Team:** MathWorks
- **Oate:** July 23, 10:00AM EDT
- **Contigention:** Online

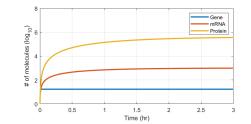


July

Constitutive gene expression - Remarks



 $[m\dot{R}NA] = k_1[Gene] - d_1[mRNA]$ $[Protein] = k_2[mRNA] - d_2[Protein]$



[Gene] is considered a constant value and depends on: the Origin of Replication and the Plasmid Copy Number where the Gene is cloned. We are considering:

- RNA polymerase and Ribosomes are sufficient enough so that they are not limiting the kinetics.
- Binding/Unbinding processes are much faster than transcription and translation.
- Protein degradation includes growth asociated dilution.

Questions? Ask writing in the chat or contact me by email (alvig2 [at] upv [dot] es)

Stay tuned, next Section 2:

Derivation of a Hill function from

the law of mass action (15 min)









Synthetic Biology and Biosystems Control Lab Valencia UPV

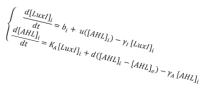
Modeling: ODEs and Hill Functions Section 2: Derivation of the Hill Function

by Alejandro Vignoni (alvig2@upv.es)



An iGEM Measurement Committee Webinar

Week 2, June 23rd, 2020

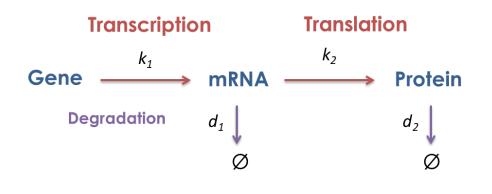


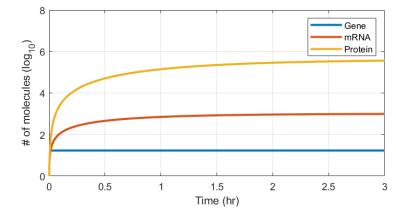
Today Webinar's Topics



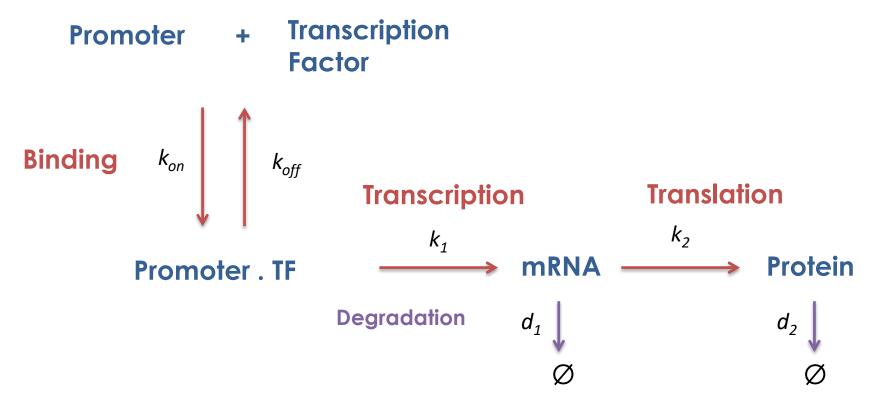
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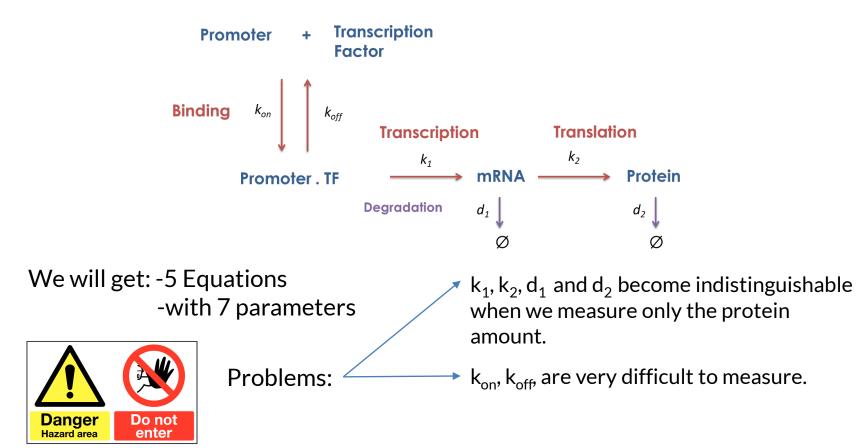
Remember: Constitutive gene expression

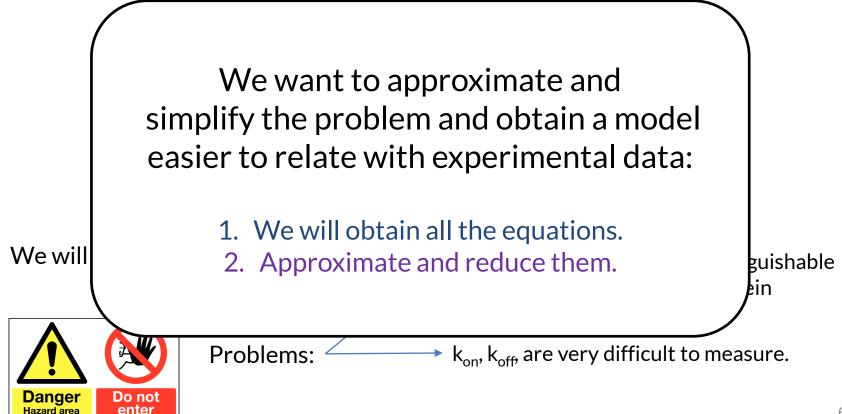




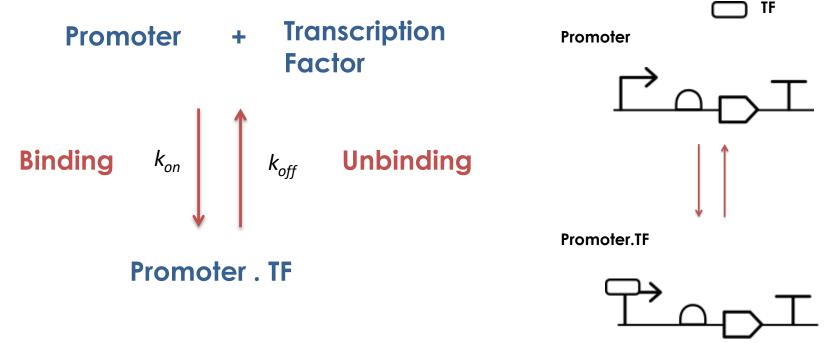
 $[mRNA] = k_1[Gene] - d_1[mRNA]$ $[Protein] = k_2[mRNA] - d_2[Protein]$

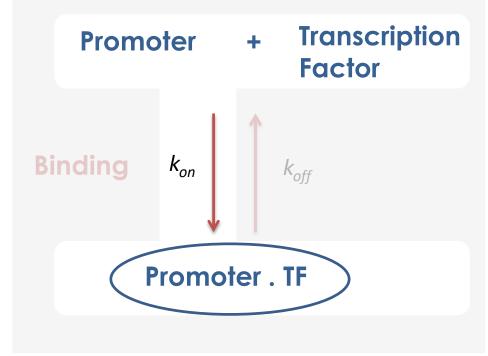


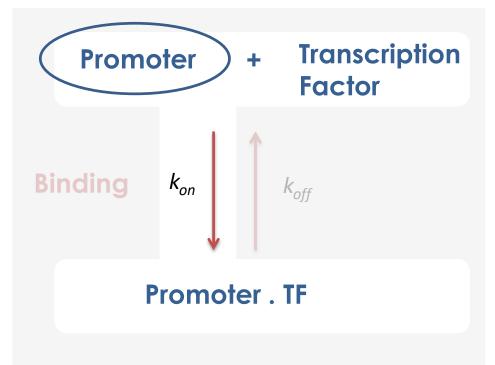




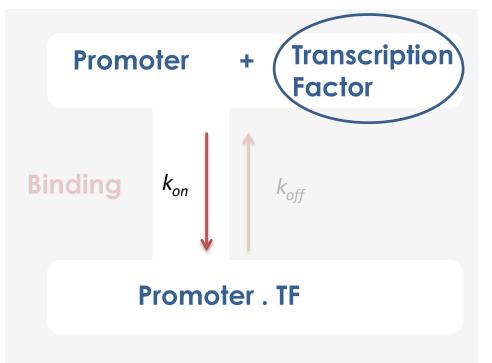
Part I: Getting the model





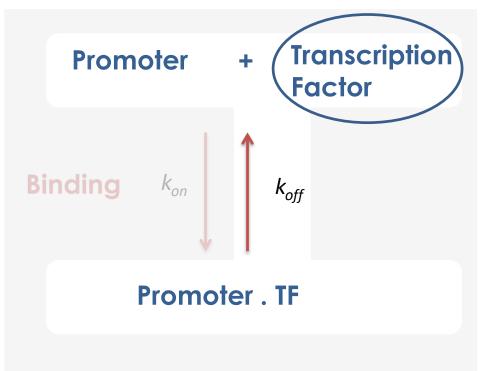


$$[Prom] = -k_{on} [Prom][TF]$$



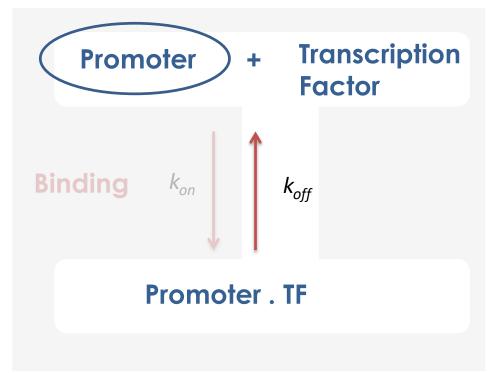
$$[Prom] = -k_{on} [Prom][TF]$$

$$[TF] = -k_{on} [Prom][TF]$$



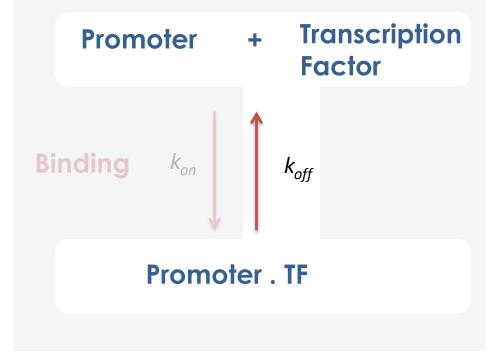
$$[Prom] = -k_{on} [Prom][TF]$$

$$[TF] = -k_{on} [Prom][TF] +k_{off} [Prom. TF]$$



$$[Prom] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$$

$$[TF] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$$



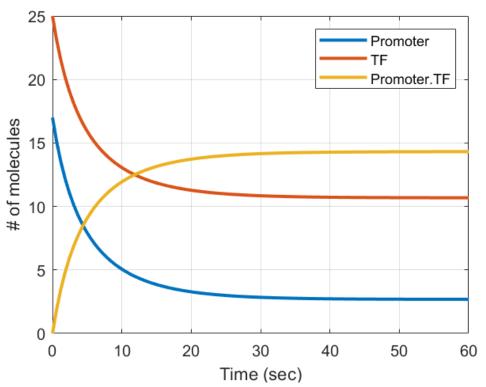
 $[Prom.TF] = k_{on} [Prom][TF]$ $-k_{off}[Prom.TF]$

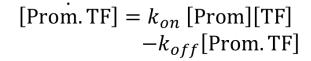
$$[Prom] = -k_{on} [Prom][TF] +k_{off} [Prom. TF]$$

$$[TF] = -k_{on} [Prom][TF] +k_{off} [Prom. TF]$$

Simulation

Main_TF.m





 $[Prom] = -k_{on} [Prom][TF]$ $+k_{off}[Prom.TF]$

$$[TF] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$$

Starting with: 17 Promoters (Plasmid copy number) 25 molecules of Transcription Factor (TF) $k_{on} = 0.5$ molecules⁻¹ min⁻¹ $k_{off} = 1$ min⁻¹

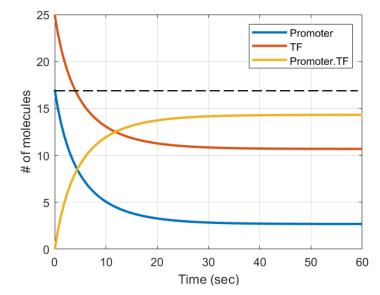


Part II: Model Reduction

 $[Prom] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$ $[TF] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$ $[Prom. TF] = k_{on} [Prom][TF] - k_{off} [Prom. TF]$

Remarks

- A First two equations are equal (Blue and red)!
- A The sum of the first one and the third one is identically zero (Blue and yellow)!
- A We can use this fact (promoter invariance) to simplify the equations and reduce the model.





Promoter invariance (constant Plasmid Copy Number)

 $[Prom] = -k_{on} [Prom][TF] + k_{off} [Prom. TF]$ $[Prom. TF] = k_{on} [Prom][TF] - k_{off} [Prom. TF]$

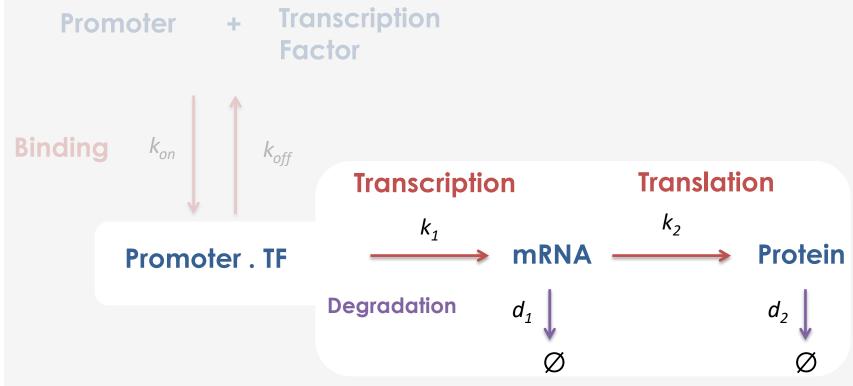
[Prom. TF] + [Prom] = 0

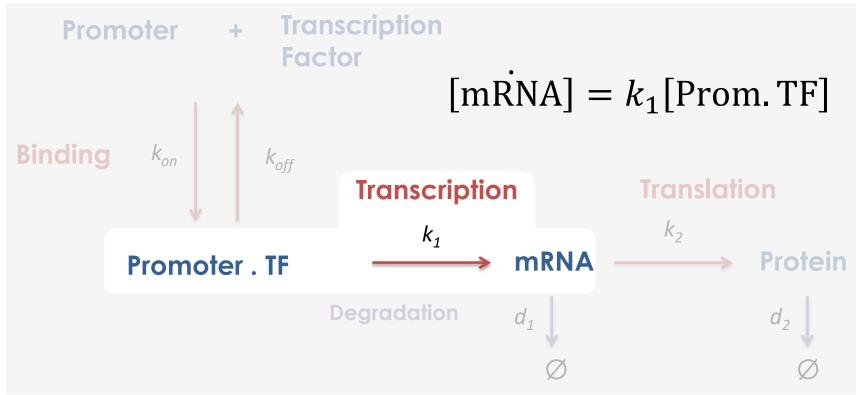
+

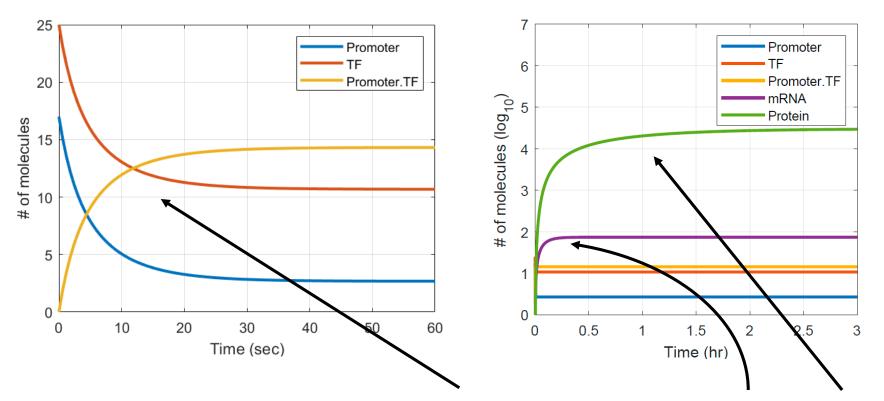
 $[Prom, TF] + [Prom] = C_N \leftarrow Plasmid Copy Number$

 $[Prom] = C_N - [Prom. TF]$ Save this one, we will use it later.

Part I: Getting the model







Note the difference in time scales: Binding in the seconds, transcription/translation from minutes to hours. 19



Fast Transcription Factor – Promoter binding

Because of the difference in time scales: Binding in the seconds, transcription/translation from minutes to hours; we can say that TF rapidly binds to the promoter and this reaction reaches equilibrium very fast.

This is called Quasy Steady State Approximation (QSSA).

$$[Prom. TF] = k_{on} [Prom][TF] - k_{off}[Prom. TF]$$

$$0 = k_{on} [Prom][TF] - k_{off}[Prom. TF]$$

From invariance (previous slide)

[Prom TF] ≈ 0

$$[Prom] = C_N - [Prom. TF]$$

Fast Transcription Factor – Promoter binding

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This is called Quasy Steady State Approximation (QSSA).

$$[Prom. TF] = k_{on} [Prom][TF] - k_{off}[Prom. TF]$$

$$0 = k_{on} \text{ [Prom][TF]} - k_{off} \text{[Prom. TF]}$$

From invariance (previous slide)

[Prom. TF] ≈ 0

 $[Prom] = C_N - [Prom. TF]$

Using these two, we will derive the Hill function

Replacing the free promoter equation into the TF bound Promoter one:

$$[Prom] = C_N - [Prom. TF]$$

$$0 = k_{on} [Prom][TF] - k_{off}[Prom. TF]$$

$$0 = k_{on} (C_N - [Prom. TF])[TF] - k_{off}[Prom. TF]$$

Solving for the TF bound Promoter:

 $k_{on} (C_N - [Prom. TF])[TF] = k_{off}[Prom. TF]$

 k_{on} [TF] $C_N - k_{on}$ [TF][Prom. TF] = k_{off} [Prom. TF]

 k_{on} [TF] $C_N = k_{on}$ [TF][Prom. TF] + k_{off} [Prom. TF] A bit of algebra...

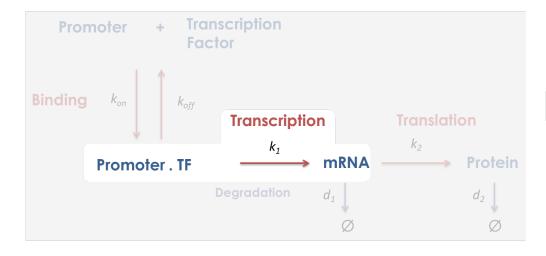
 $k_{on} [TF]C_N = (k_{on} [TF] + k_{off}) [Prom. TF]$ k [TF] [TF] [TF]

$$[\text{Prom.TF}] = C_N \frac{k_{on}[\text{TF}]}{k_{on}[\text{TF}] + k_{off}} = C_N \frac{[\text{TF}]}{\frac{k_{off}}{k_{on}} + [\text{TF}]} = C_N \frac{[\text{TF}]}{K_d + [\text{TF}]}$$

Gene expression regulation by Transcription Factors (TF) We get the Hill function (with Hill coefficient n=1) Hill Function (Activator) $[Prom. TF] = C_N \frac{[TF]}{K_d + [TF]}$ 0.8 [Prom. TF] **Activator** TF $C_N = 1$ molecule Promoter 0.2 $K_d = 500$ molecules 10^{2} 10^{0} 10¹ 10^{3} 10^{4} 10⁵

TF



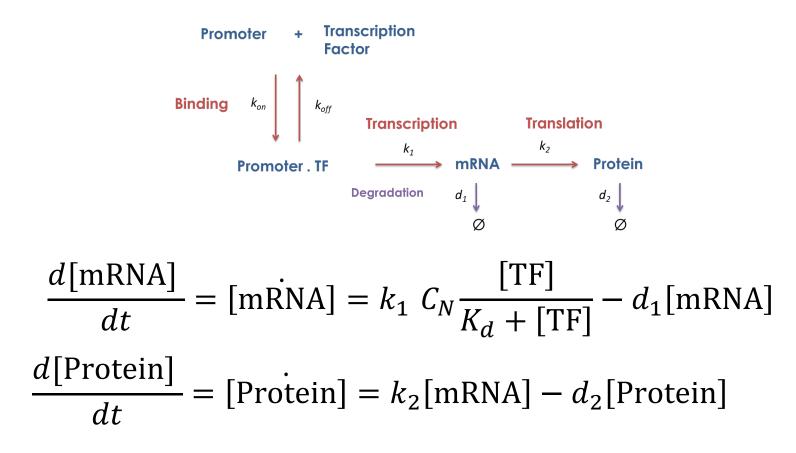


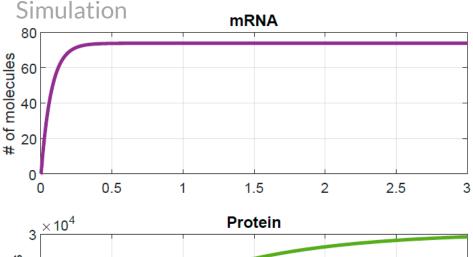
[Prom. TF] =
$$C_N \frac{[TF]}{K_d + [TF]}$$

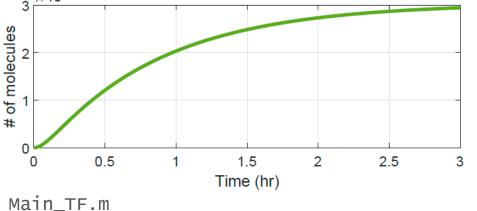
 $[mRNA] = k_1[Prom. TF]$

The complete equation for the mRNA

$$[\dot{mRNA}] = k_1 C_N \frac{[TF]}{K_d + [TF]} - d_1[mRNA]$$







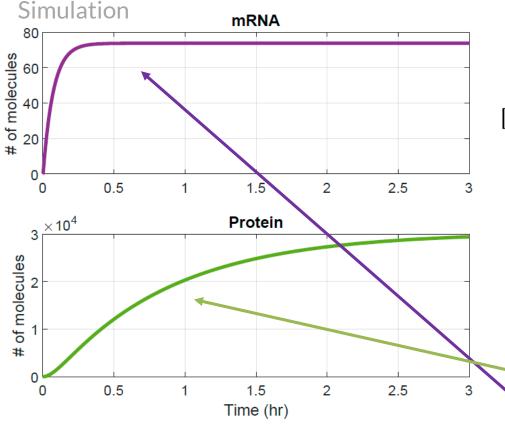
$$[m\dot{R}NA] = k_1 C_N \frac{[TF]}{K_d + [TF]} - d_1[mRNA]$$
$$[Protein] = k_2[mRNA] - d_2[Protein]$$

Parameters:

CN = 17 molecules (Plasmid copy number) Kd = 2 molecules

TF = 25 molecules (Transcription Factor) The other parameters same than constitutive

MATLAB



$$[m\dot{R}NA] = k_1 C_N \frac{[TF]}{K_d + [TF]} - d_1[mRNA]$$
$$[Protein] = k_2[mRNA] - d_2[Protein]$$

Parameters:

CN = 17 molecules (Plasmid copy number)

Kd = 2 molecules

TF = 25 molecules (Transcription Factor)

The other parameters same than constitutive

Note the difference in time scales: transcription (mRNA) in minutes, translation (Protein) hours.

MATLAB

Now, as mRNA is much faster than Protein production... we use the same trick than before (QSSA):

$$\begin{bmatrix} mRNA \end{bmatrix} \approx 0 \\ 0 = k_1 C_N \frac{[TF]}{K_d + [TF]} - d_1 [mRNA] \Longrightarrow [mRNA] = \frac{k_1}{d_1} C_N \frac{[TF]}{K_d + [TF]}$$

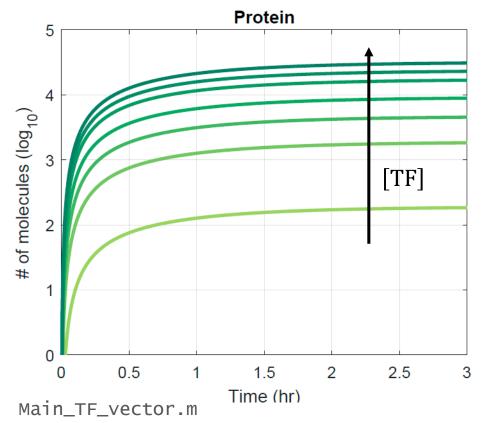
$$\frac{d[\text{Protein}]}{dt} = [\text{Protein}] = \alpha \frac{[\text{TF}]}{K_d + [\text{TF}]} - d_2[\text{Protein}]$$

$$\alpha = k_2 \frac{k_1}{d_1} C_N$$

29



Simulation



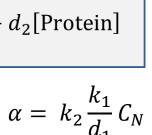
$$[Protein] = \alpha \frac{[TF]}{K_d + [TF]} - d_2[Protein]$$

With:

 α = 720 molecules min⁻¹ K_d = 2 molecules d_2 = 0.02 min⁻¹

(this means 34 min of doubling time)

[TF]: from 0.1 molecule to 25 molecules of Transcription Factor

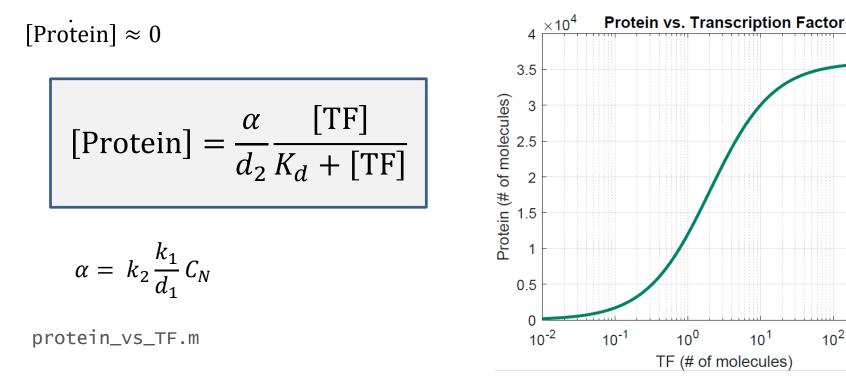




Now, if we want the steady state we can use the same trick (QSSA) that we used before (equilibrium expression of protein, data at the end of the experiment)

 10^{2}

 10^{3} 31



Questions? Ask writing in the chat or contact me by email (alvig2 [at] upv [dot] es)

Stay tuned, next Section 3:

Hill function examples and intuitions: effects of parameters on activators,

repressors, hybrid promoters, using a Matlab exploration package.







ECCEPT 2020 Measurement

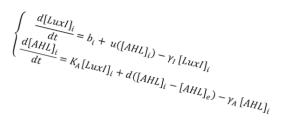
Synthetic Biology and Biosystems Control Lab Valencia UPV

Modeling: ODEs and Hill Functions Section 3: Hill function examples and intuitions by Alejandro Vignoni (alvig2@upv.es)

An iGEM Measurement Committee Webinar

Week 2, June 23rd, 2020

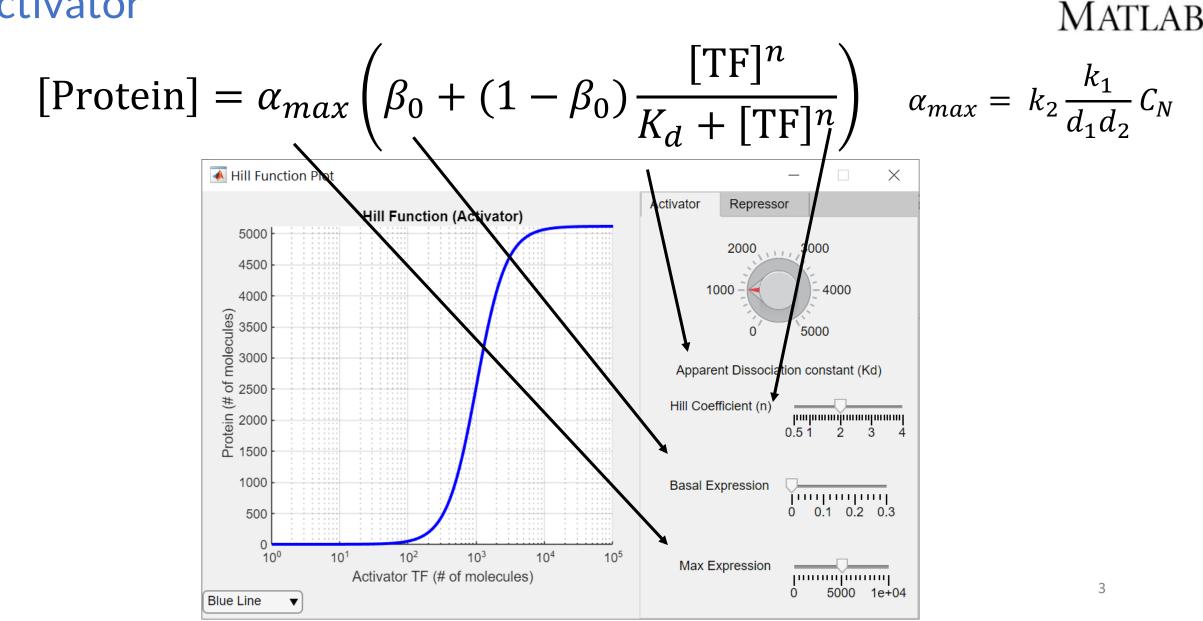


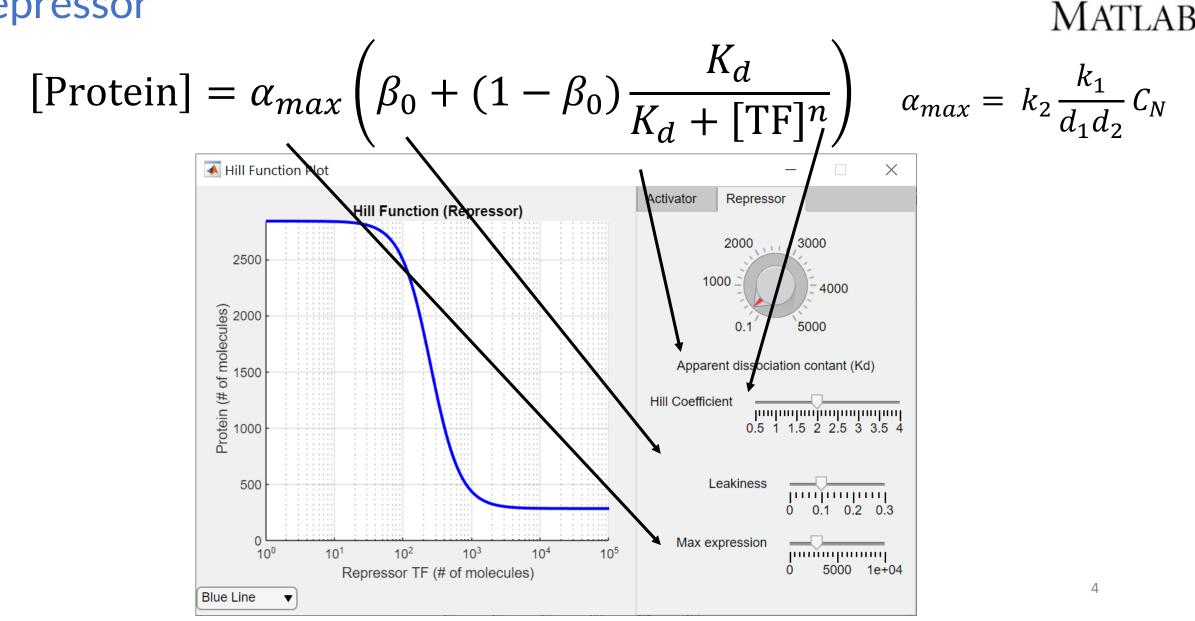


Today Webinar's Topics

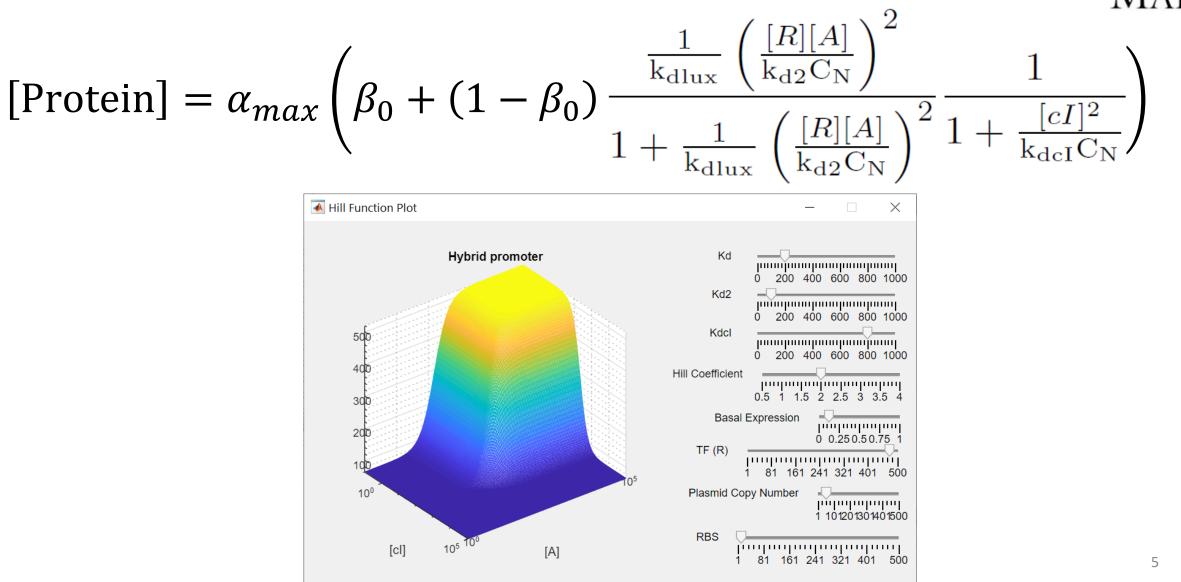


- A Section 1: ODEs, the law of mass action, and the central dogma (15 min)
- A Section 2: Derivation of a Hill function from the law of mass action (15 min)
- A Section 3: Hill function examples and intuitions: effects of parameters on activators, repressors, hybrid promoters, using a Matlab exploration package. (15min)
- AQ&A (at the end of each 15 minutes block, total 15 min)





Gene expression regulation by Transcription Factors (TF) Hybrid Promoter



Questions? Contact me by email (alvig2 [at] upv [dot] es)

Thank You & Have an Exceptional Year of iGEM!

Next Modeling seminar Week 3a Modeling circuits with ODEs and experimental data, stay tuned!



Go check out the Measurement Hub! <u>https://2020.igem.org/Measurement</u>

