

# Developing the Fluoride Riboswitch as a Technology to Combat Excessive Water Fluoridation



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## Introduction

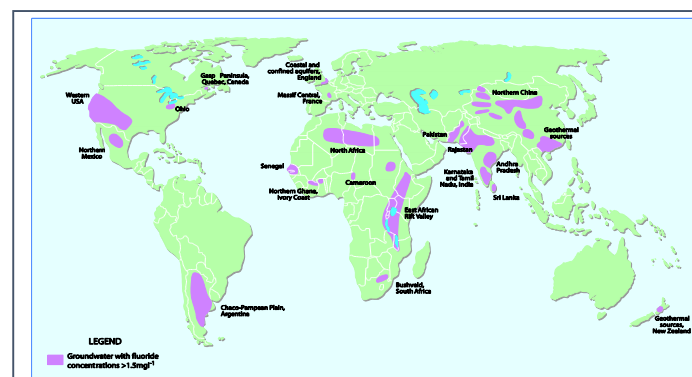
Fluoride is present in all bodies of water. Within the oceans, rivers, lakes, and groundwater, the levels of fluoride depend on the erosion of the sediments in the area. When fluoride concentrations are at toxic level, major health concerns arise. According to WHO, concentrations at or above 1 mg/kg of body weight are deemed poisonous and recommends concentrations of fluoride do not exceed **1.5mg/mL** in water sources. Constant exposure to 10 mg/L to 6 mg of fluoride can lead to fluorosis and developmental and reproductive concerns (**Fig 1**).



**Fig. 1: A child with skeletal fluorosis**

<http://www.inrem.in/fluorosis/health.html>

In certain areas in countries like China, India, and Sri Lanka, water sources are decentralized and residents can experience concentrations of fluoride as high as 30 mg/L (**Fig.2**).

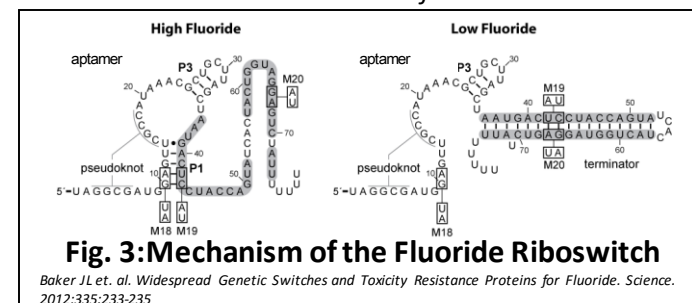


**Fig. 2: Map of documented occurrences of high-fluoride groundwater**

Edmunds W.M., Smedley P.L. (2013) Fluoride in Natural Waters. In: Selinus O. (eds) Essentials of Medical Geology. Springer, Dordrecht

## Our Solution

We envision developing technologies utilizing fluoride riboswitches that can sequester, bioremediate, or detect fluoride in water ... *but what is a fluoride riboswitch?*



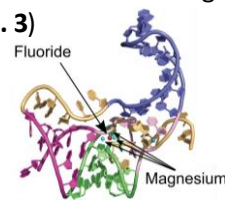
**Fig. 3: Mechanism of the Fluoride Riboswitch**

Baker J.L et. al. Widespread Genetic Switches and Toxicity Resistance Proteins for Fluoride. Science. 2012;335:233-235

The **Fluoride Riboswitch** is an RNA transcriptional riboswitch that contains two functional elements: the aptamer, which binds to fluoride, and the terminator which regulates transcription of the downstream gene. When fluoride is present the terminator is released allowing transcription of the downstream gene (**Fig. 3**)

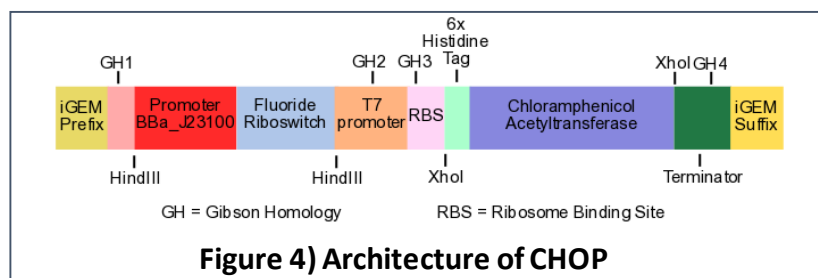
### Crystal structure of the fluoride riboswitch aptamer domain

Ren A et. al. "Fluoride ion encapsulation by Mg2+ ions and phosphates in a fluoride riboswitch" 2012 Nature 486, 85-89



## Our Design: CHOP

In order to develop technologies using the fluoride riboswitch we first need a platform to characterize the responsiveness of fluoride riboswitches that was amenable to high-throughput selection methodologies. To achieve this we developed the **fluoride riboswitch regulated chloramphenicol acetyltransferase operon (CHOP)** (**Figure 4**). To determine if CHOP would work we used the fluoride riboswitch from *B. cereus* which has been characterized in the literature



**Figure 4) Architecture of CHOP**

### How CHOP works:

- Chloramphenicol acetyltransferase (CAT) provides resistance to the antibiotic chloramphenicol

- When sufficient fluoride enters the cell to activate the riboswitch CAT is transcribed allowing the bacteria to grow

- CHOP needs to be used in *E. coli* that have the fluoride exporter gene *crcB* deleted ( $\Delta$ *crcB*) so that fluoride can accumulate inside the cell. **In the literature  $\Delta$ *crcB* is reported to be sensitive to fluoride concentrations above 500 $\mu$ M.**

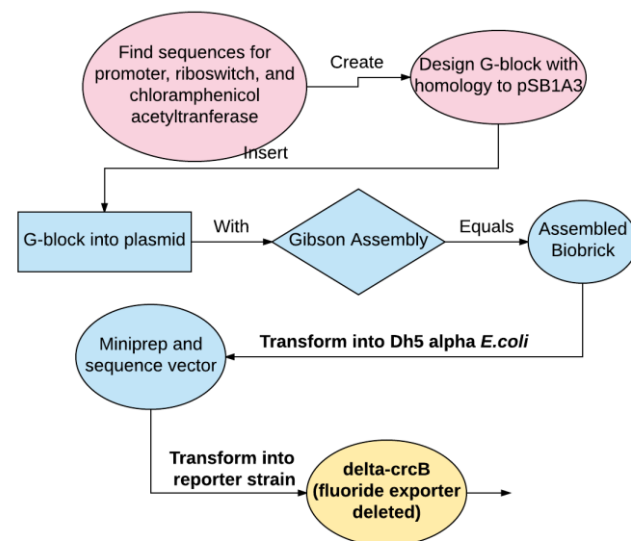
### How can YOU use CHOP:

- Characterize Transcriptional Riboswitches: New promoter riboswitch pairs can easily be cloned in using Gibson: order at G-block with appropriate overhangs (see the sequence for part BBa\_K2290000 to find the appropriate overhangs) and linearize the vector with HindIII. This system should be amenable to screen any transcription riboswitch.

- Regulate New Genes with Transcriptional Riboswitches: New genes of interest can easily be cloned in using Gibson: order at G-block with appropriate overhangs and linearize the vector with XhoI.

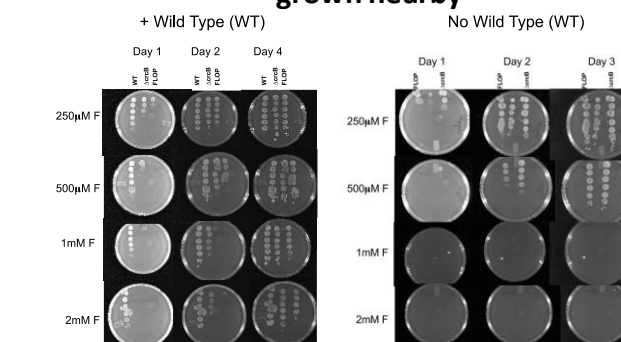
## Cloning Methodology

### Design and Cloning

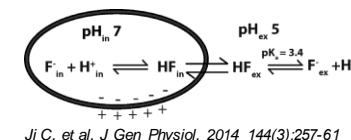


## Results

### WT *E. coli* protect $\Delta$ *crcB* from fluoride toxicity when grown nearby

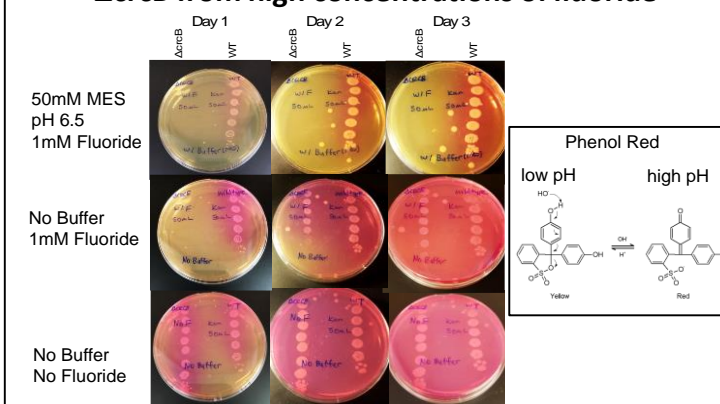


### Intracellular fluoride accumulation is pH-dependent

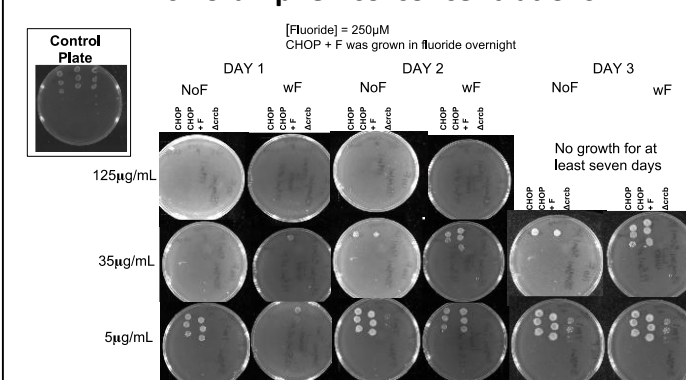


Ji C. et al. J Gen Physiol. 2014 144(3):257-61

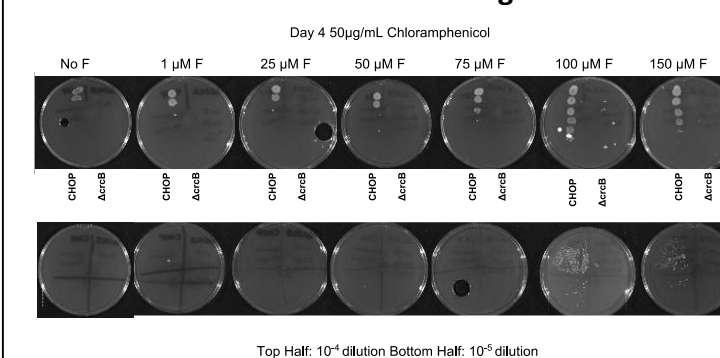
### WT *E. coli* alter the pH of the agar plates to protect $\Delta$ *crcB* from high concentrations of fluoride



### Fluoride dependence of CHOP growth on different chloramphenicol concentrations



### Characterizing the responsiveness of the *B. cereus* fluoride riboswitch using CHOP



## Acknowledgements

We would like to thank the Kuhlman Lab for providing lab equipment and reagents, Dr. Joseph Harrison from UNC-Chapel Hill for instruction and guidance, UNC-Chapel Hill for lab space, Bo Zhao and Minnie Langlois for helping with experiments, Dr. Randy Stockbridge for troubleshooting, and Siyu Wang for guidance in Wiki coding. All of the donors who gave us funding, including Qorvo and the countless donors who made personal contributions. We would also like to thank Mr. William Vincent, Dr. Stephen Snyder, and Ms. Patricia Berge for their time and effort in supporting the team.

## Conclusion

- CHOP allows for characterization of the responsiveness of fluoride riboswitches
- WT *E. coli* can protect the  $\Delta$ *crcB* strain from fluoride toxicity by altering the pH of the plates
- The *B. cereus* fluoride riboswitch works best at 100 $\mu$ M fluoride concentration (1.8mg/mL). Therefore riboswitches with improved responsiveness to fluoride would be desirable for future applications.

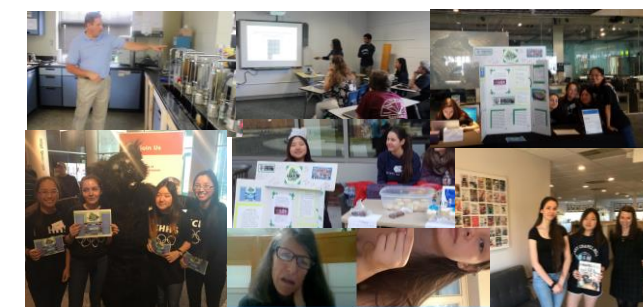
## Future Directions

First, we will use CHOP to screen for native riboswitches or evolve the *B. cereus* riboswitch to find riboswitches with varied responsiveness to fluoride. With these tools we hope to:

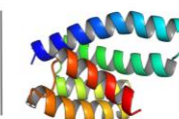
- **Sequester** fluoride from drinking water by attaching many copies of the RNA to nanoparticles.
- **Bioremediate** fluoride-contamination water by controlling the expression of enzymes that can metabolize fluoride (i.e. fluorinase)
- **Detect** fluoride-contaminated water by controlling the expression of reporter genes (i.e. fluorescent proteins) with a suite of riboswitches with differential responsiveness to fluoride.

## Human Practices

- Darwin Day at the Raleigh Museum of Natural Sciences
- OWASA facility tour + documentary style video
- Collaboration in creating a collection of Tasty-Lab Videos (<https://www.youtube.com/watch?v=TraYbTe7Lh8>)
- Hosted Southeastern Mini iGEM Convention
- Reddit forum collaboration with Duke regarding the ethics of synthetic biology
- Rewriting the Building with Bio Malaria Kit → pertinent issue at this current point in time (the Zika Virus) with Duke and Gaston Day
- Interviews with Laura Craig (<https://www.youtube.com/watch?v=qr19hiCimz8>) and Anitha Pius



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