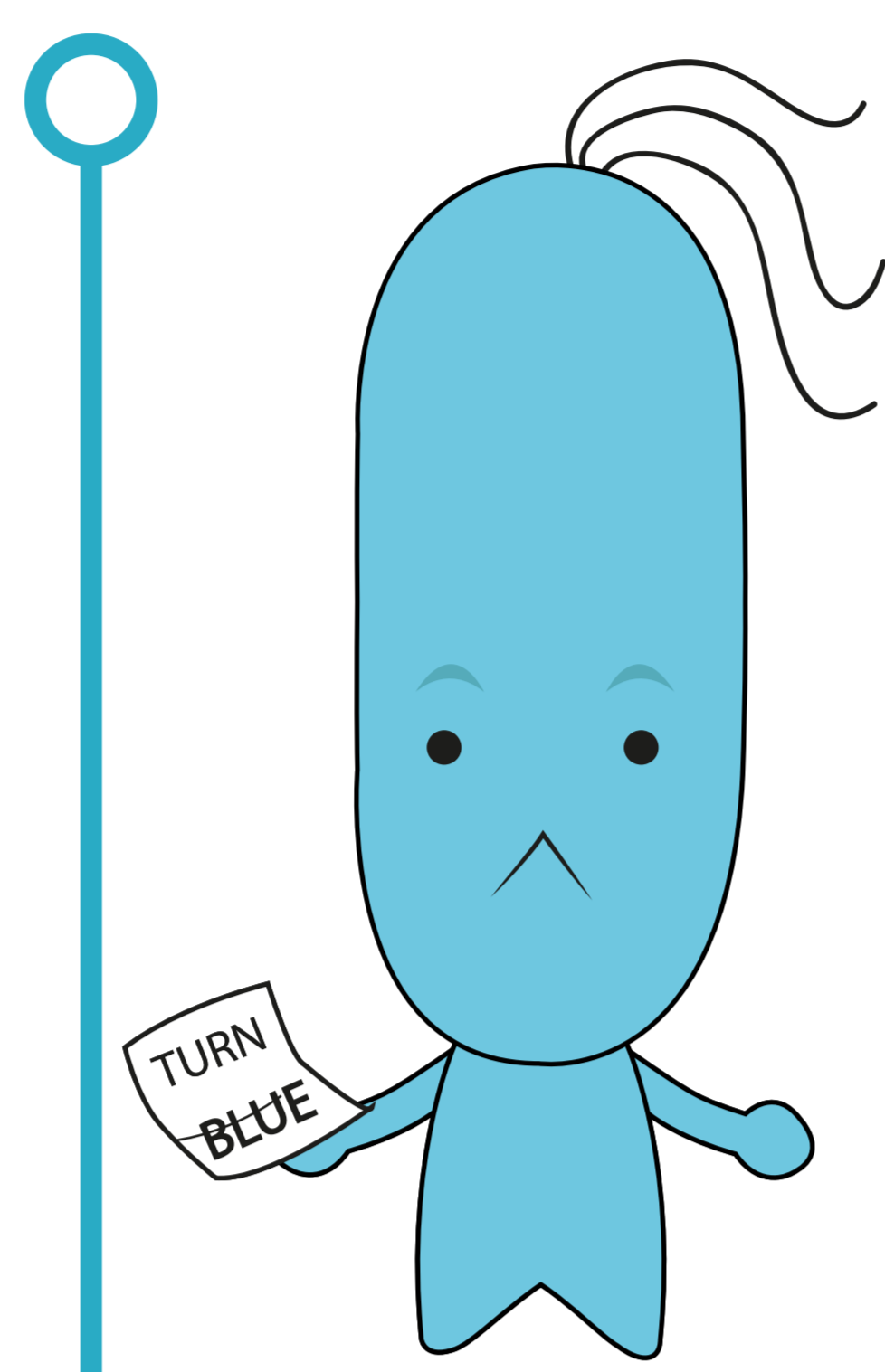


Team Sydney\_Australia

**Almost half of all fruit and and vegetables produced are damaged, lost, or wasted!**

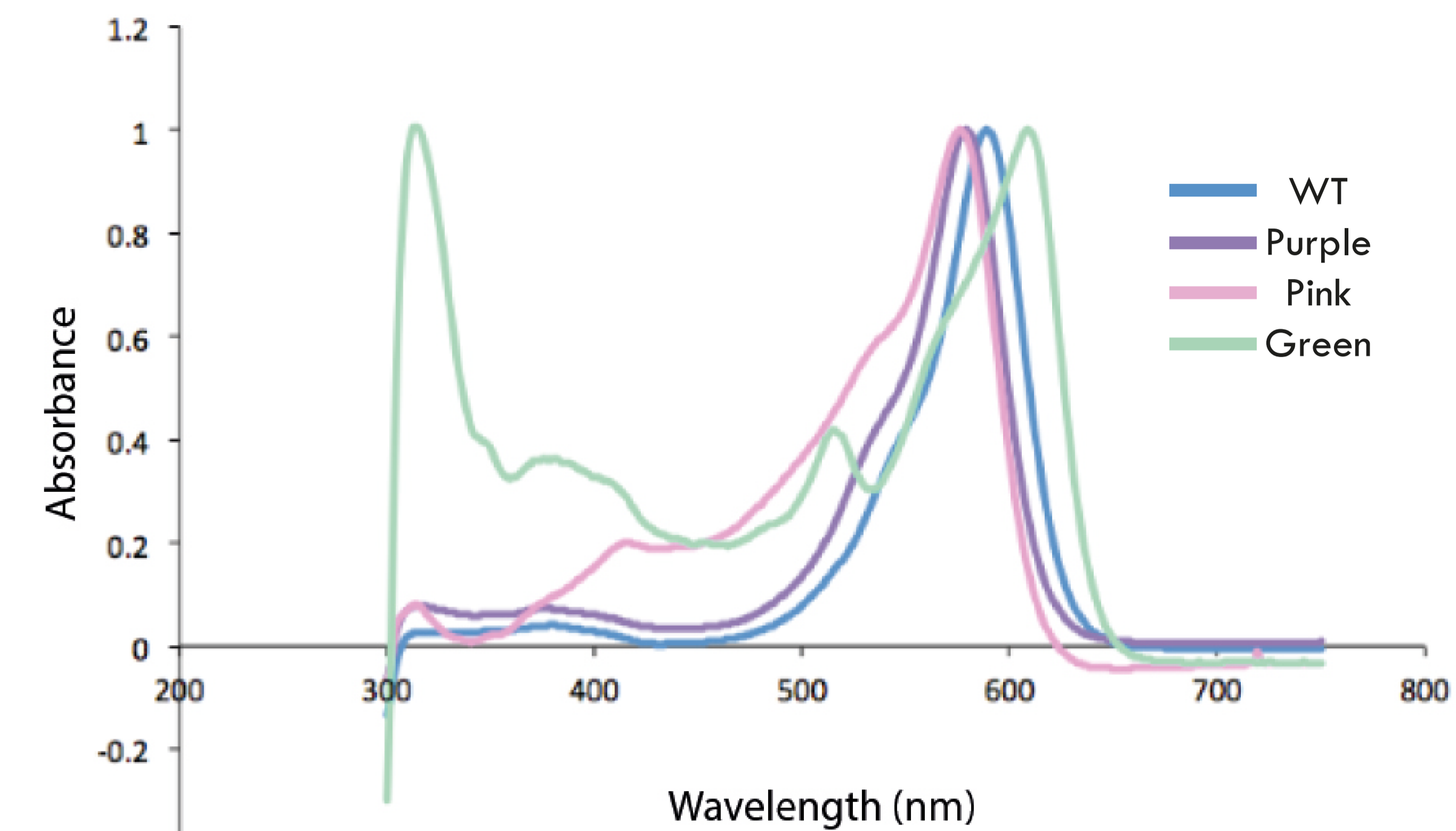
**What can synthetic biology do about it?**

Ethylene is a major hormone involved in the ripening of fruits. High demand for seasonal fruits represents a challenge in terms of long storage and transportation times. Small amounts of ethylene can cause unwanted ripening and spoilage, leading to wastage. We need a way of determining ethylene levels, but current methods are expensive, labour intensive, and frequently not portable. Our solution was to create an ethylene biosensor by engineering bacteria



We hijacked the ethylene regulatory system so that *E. coli* could express a chromoprotein (amilCP) upon detection of ethylene.

AmilCP is a blue chromoprotein and an iGEM part that we improved through error prone PCR. We generated 3 colour mutants with shifted absorbance peaks



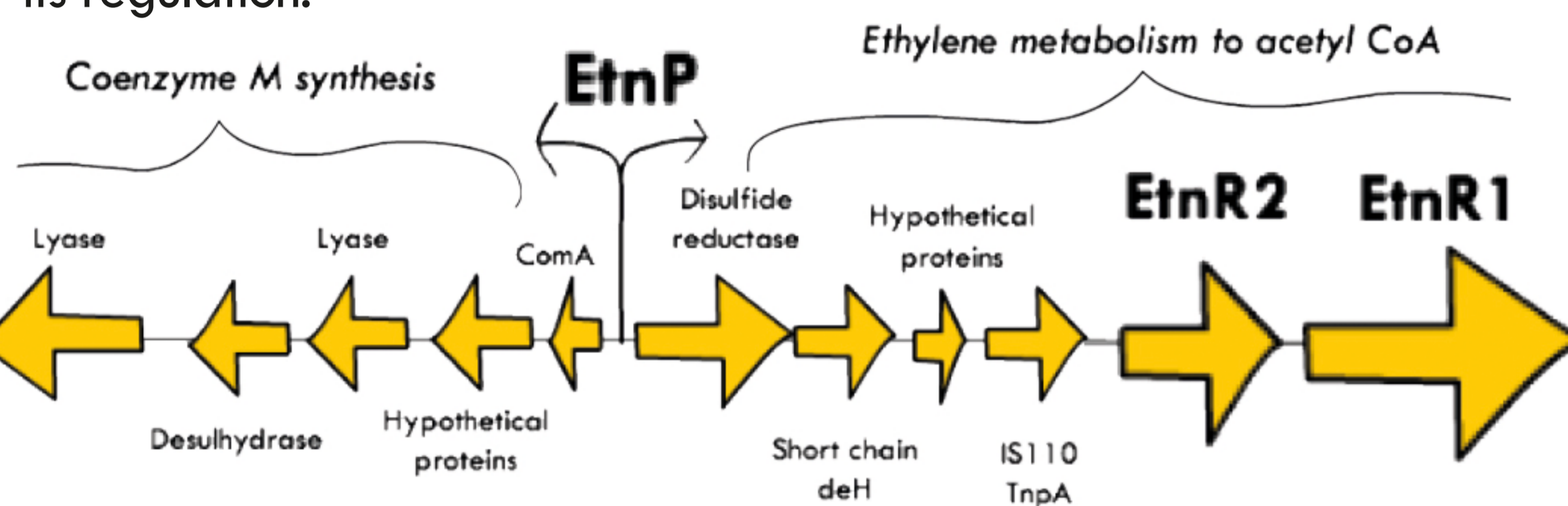
to **SENSE** ethylene

to **EXPRESS**

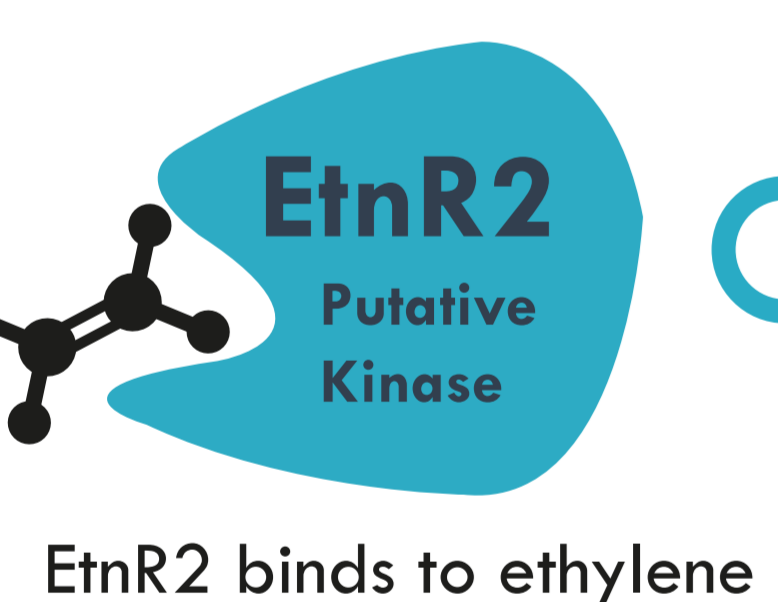
a visible output

to **KEEP** our produce **FRESH**

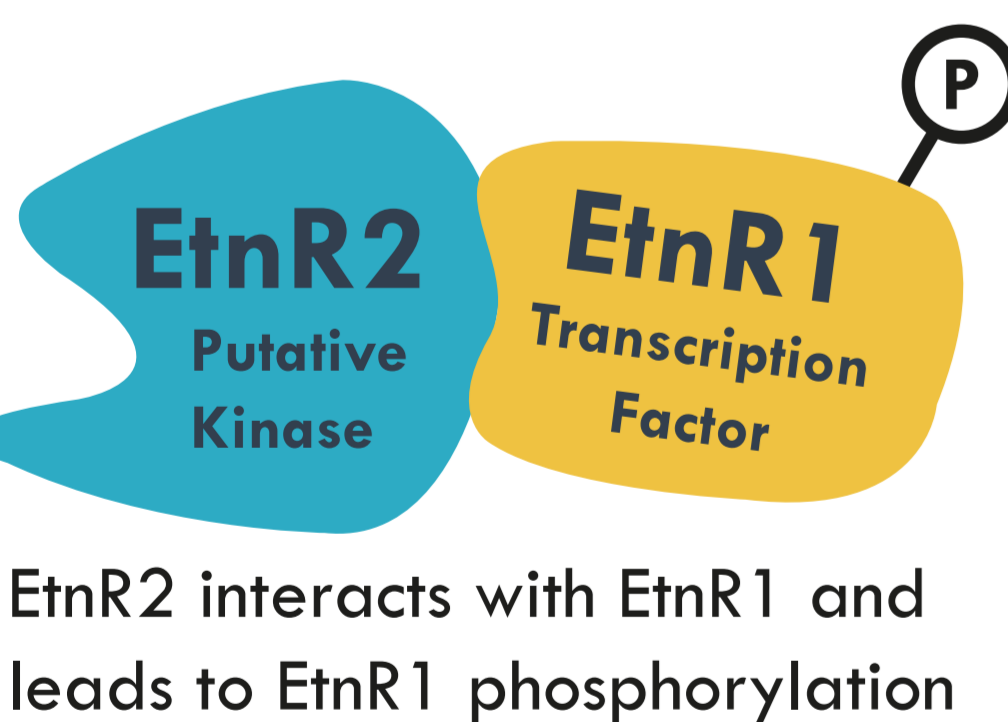
For our biosensor to sense ethylene, we turned to the ethylene-oxidising *Mycobacterium* NBB4. We investigated two genes from its ethylene metabolism operon (below), which we suspected were responsible for its regulation.



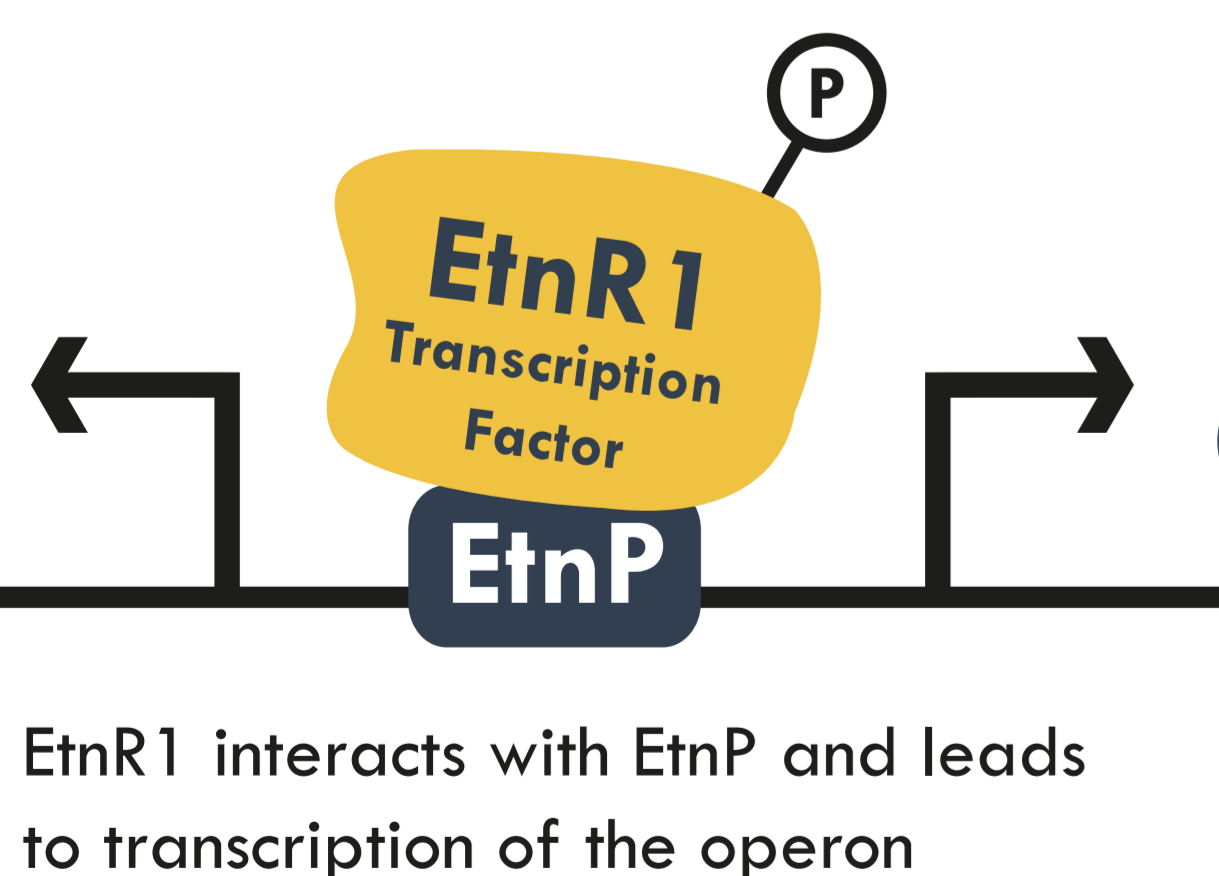
Suspected EtnR1 and EtnR2 interactions and the assays to characterise them.



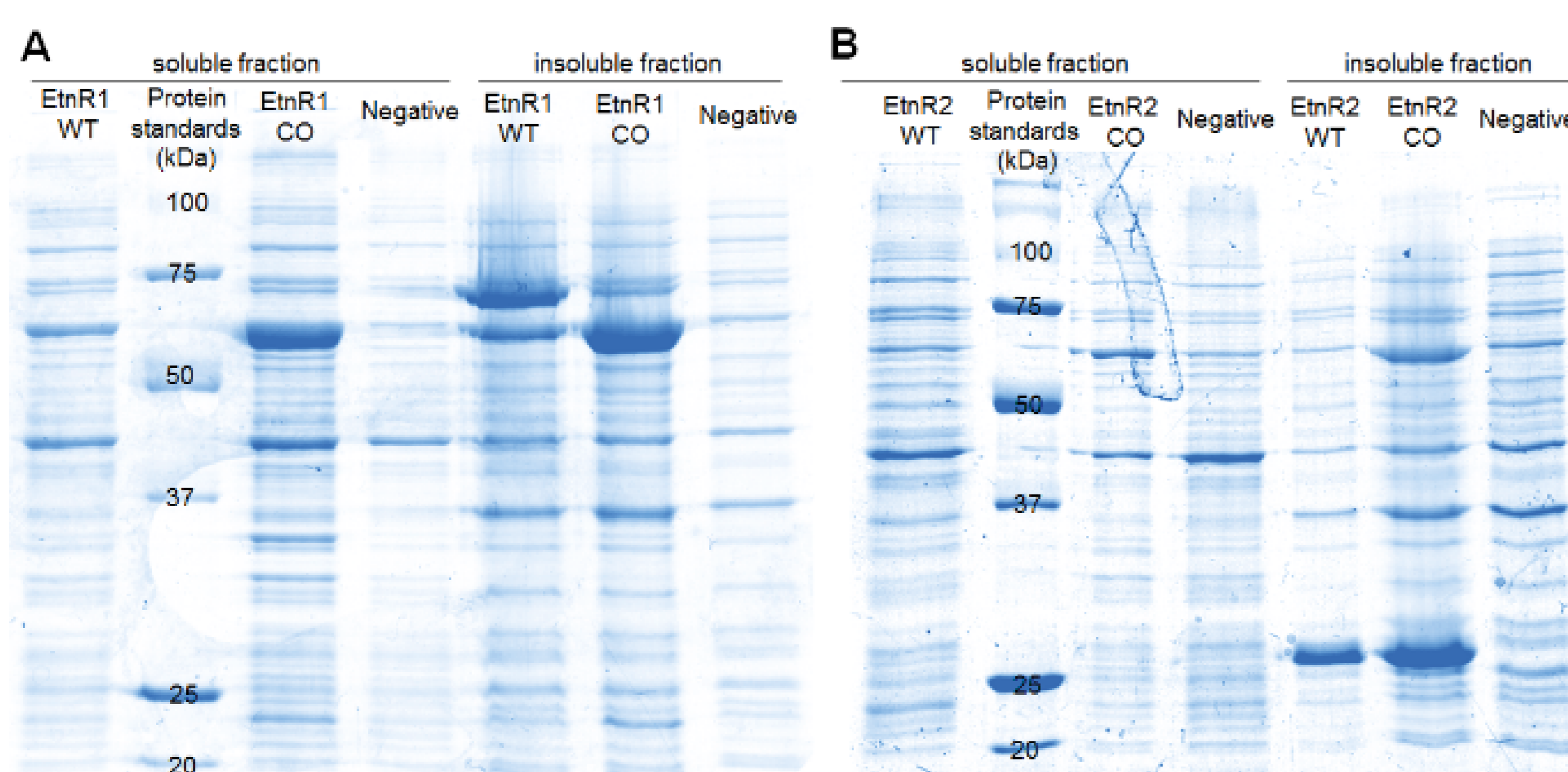
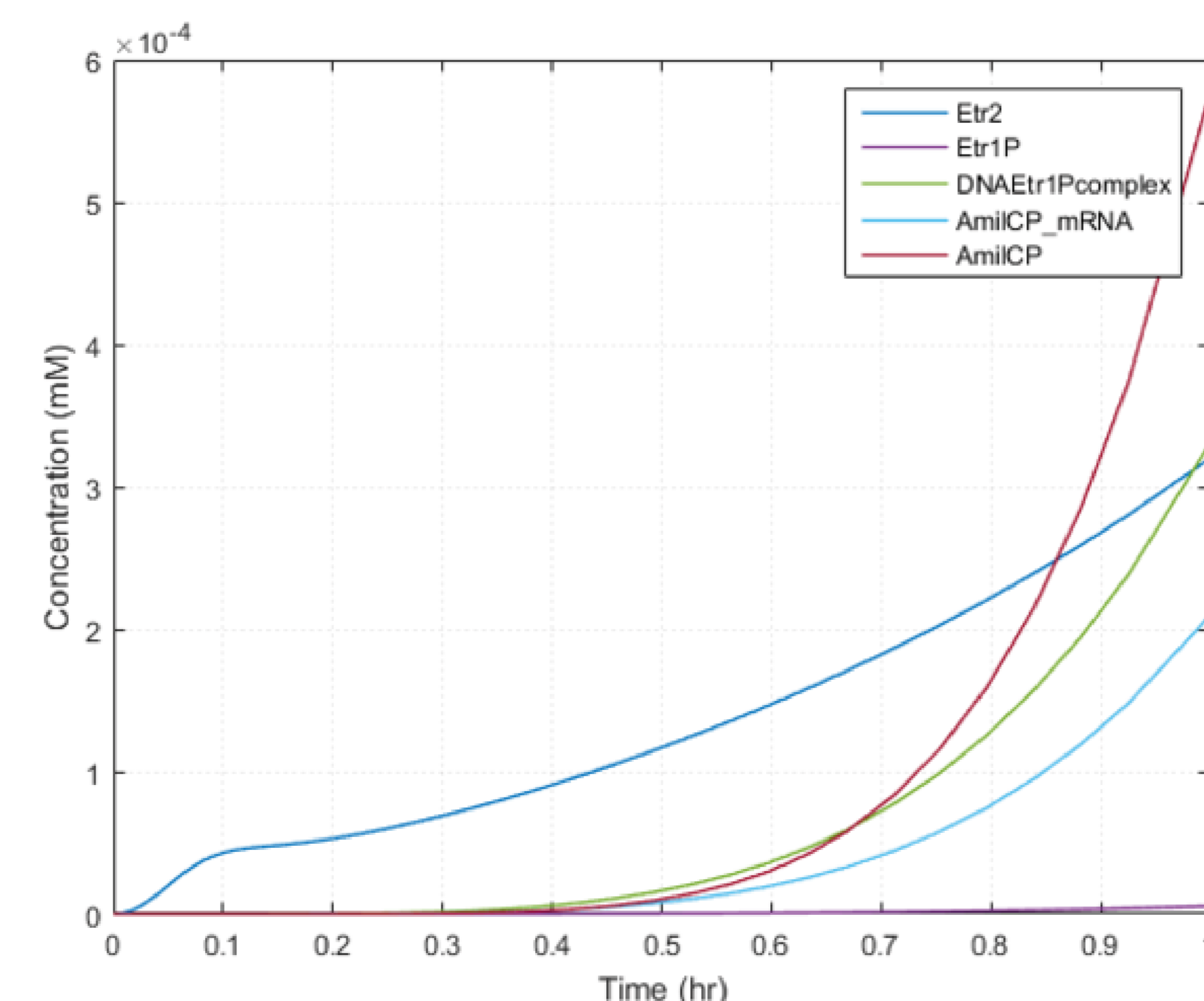
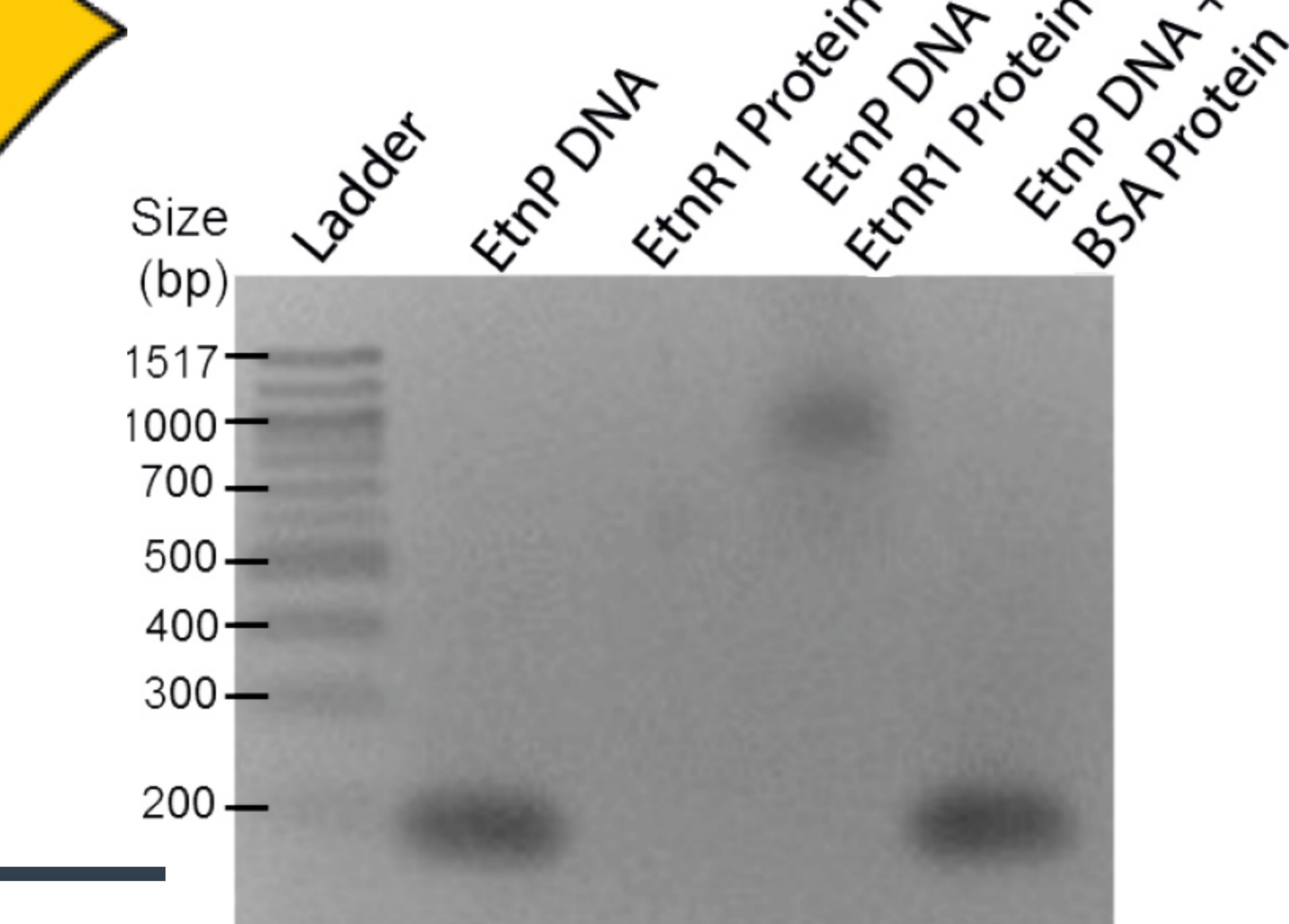
Mass spectroscopy for phosphorylation of EtnR2



Pulldown assay for binding of EtnR1 and EtnR2

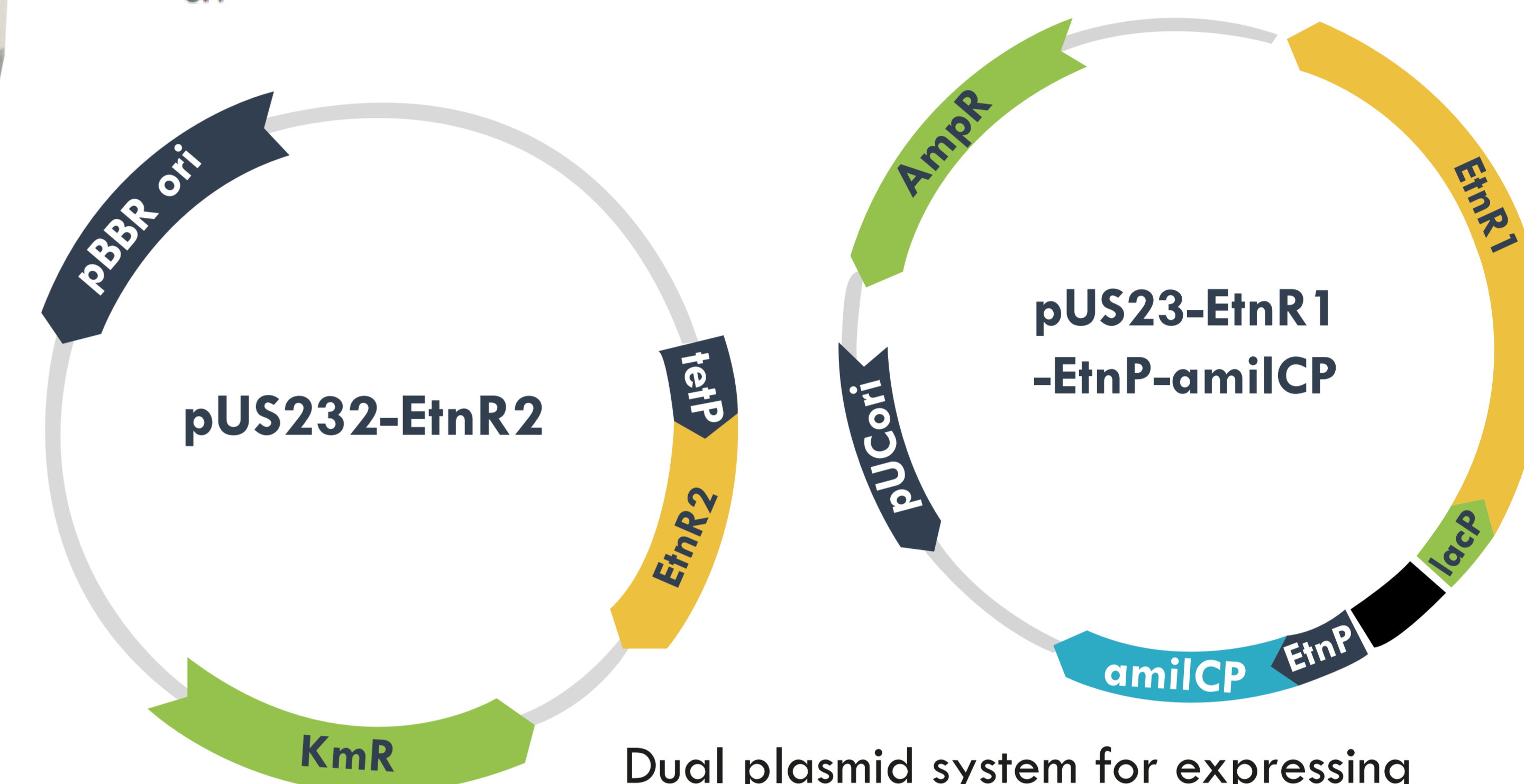


Electrophoretic mobility shift assay for binding of EtnR1 to EtnP



Above: Computational model of protein expression upon stimulation by 20mM of ethylene using MATLAB.

Left: Confirmation of expression of EtnR1 and EtnR2 in *E. coli* using SDS-PAGE.



Dual plasmid system for expressing EtnR1 and EtnR2, allowing ethylene-inducible expression of amilCP.

As an intermediate product whilst cell-free technology is being developed, a latex nanoporous biocoating will be used to immobilised GM *E. coli* on a paper base.

After consulting with potential consumers (Zespri, Avocados Australia, and Fresh Produce Group) we selected three designs for our final biosensor chassis: a fruit sticker, an industrial sticker for use on shipping containers, and a plate and strip system for warehouses.

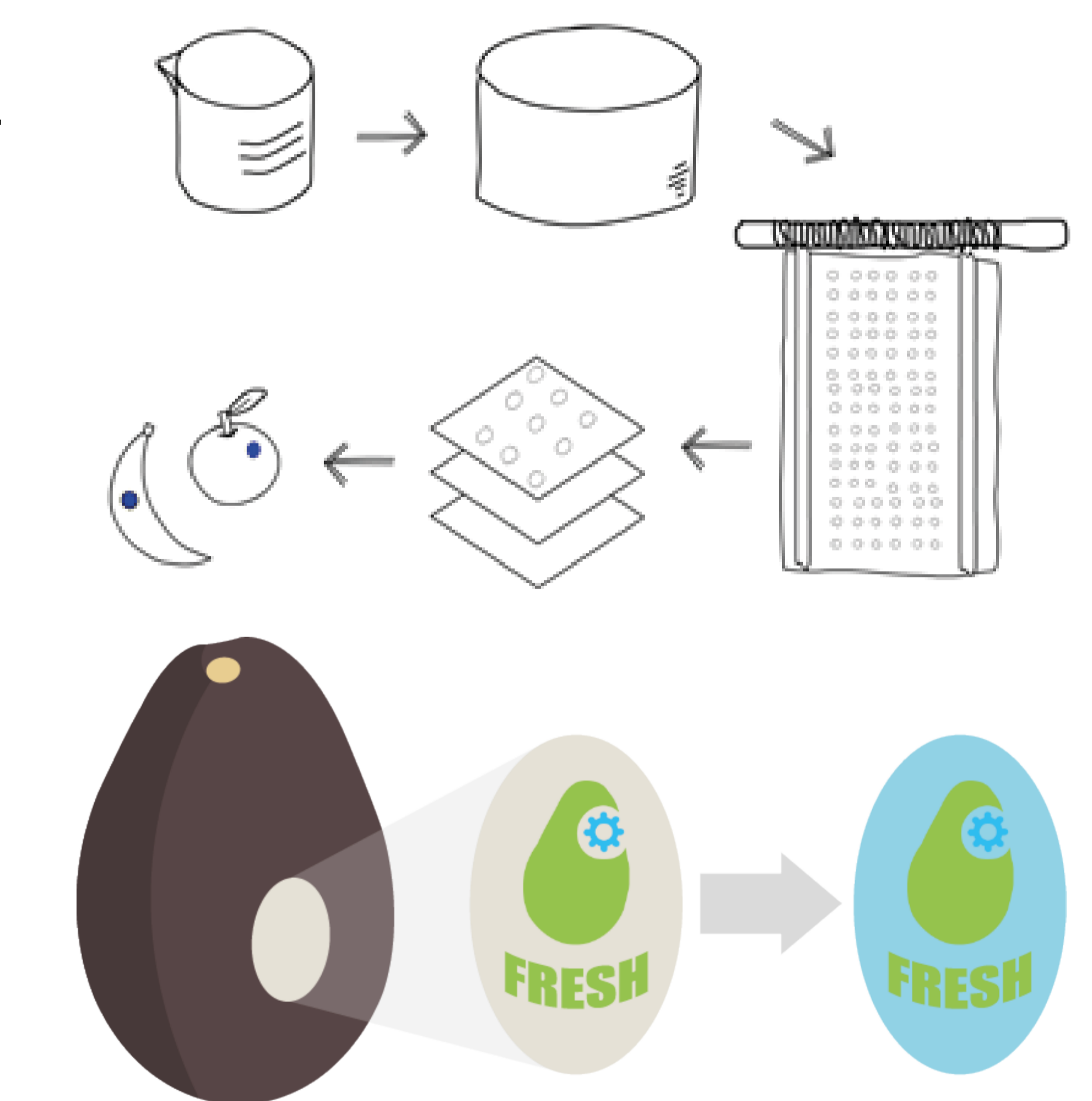


### Attributions

Flickinger, M., Schottel, J., Bond, D., Aksan, A. and Scriven, L. (2007). Painting and Printing Living Bacteria: Engineering Nanoporous Biocatalytic Coatings to Preserve Microbial Viability and Intensify Reactivity. *Biotechnology Progress*, 23(1), pp.2-17.

All experiments were performed by the iGEM USyd team. Thanks should be given to the following for their invaluable help: Dr Nicholas Coleman (supervisor) Edward Hancock (cosupervisor) Brian Jones (cosupervisor) Various lab companions: Frances, Mark, Deb Expert advice from: Antony Allen (Avocados Australia) Joseph Eckman (The Fresh Produce Group) Frank Bollen (Zespri Kiwifruits)

Contact Nicholas Coleman: nicholas.coleman@sydney.edu.au



0.01C vs. 0.04C  
Our fruit sticker vs. Current fruit labelling

0.01C vs. \$15,000+  
Our fruit sticker vs. Gas chromatography

