







Human Practices

Our project meets a need!

We turned to experts in various fields (doctors, scientists, ethics professors) to assess our project. They gave us insight which we integrated to improve the points, highlighted in green.

Teaching the public



Through fairs, youth educational meet-ups and university open-days, we: - raised awareness about antibiotics use

- explained synthetic biology and working
- principles of home-diagnostics devices - let people experiment with printed paperstrips

Sample Processing

Our detection limit

from *in vitro*

transcribed and in

vivo extracted RNA

is in the nM range.

We need an ampli-

fiction scheme to

reach a more rele-

lowest detected

10 nM

100 nM

35 aM

1 fM

vant detection.

Is the public comfortable with our product?

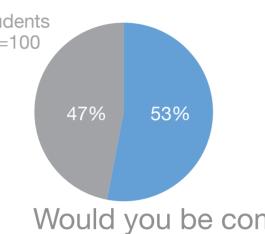
☑, Target choice

My Portability

☑, Less than \$1 per test

Stability on paper

Do you know about synthetic biology?



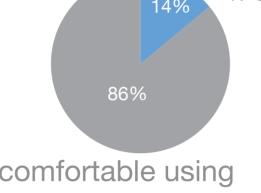
Sensitive: 35 aM

Model

M, Functional on paper

Modular and integrated

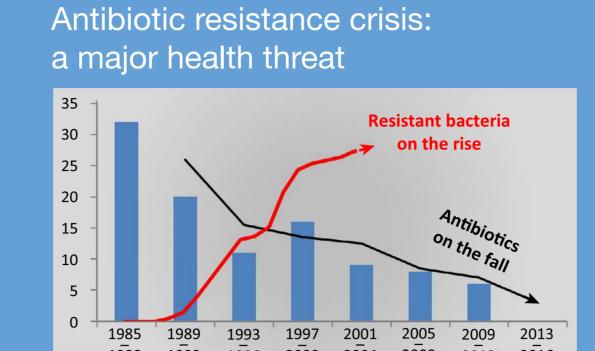
Cas13a



Would you be comfortable using a home-diagnostic device for viral and bacterial infections?

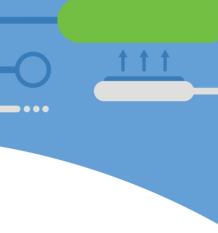
Overview

To prevent further spread of antibiotic resistance, we designed an affordable, rapid point-of-care test for infectious diseases to distinguish viral and bacterial pathogens: CascAID. The modules of this device enable extraction, amplification and detection of target RNA sequences to fulfil the A.S.S.U.R.E.D. criteria.

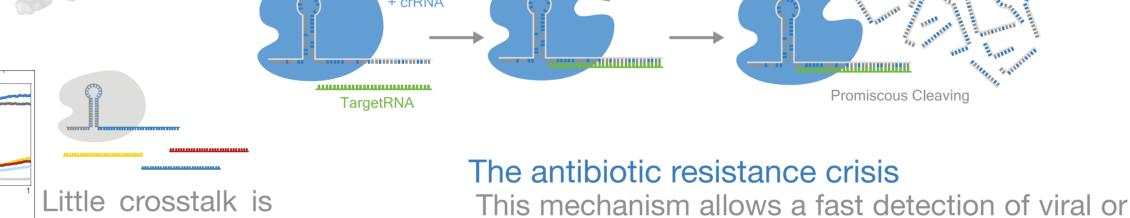


Because antibiotics are prescribed unnecessarily, bacteria grow resistant and simple infections are on the rise.





Cas13a is a CRISPR-associated protein which recognizes RNA sequences, with the help of a complementary CRISPR RNA (crRNA). It then becomes an unspecific RNase. Its singlenucleotide specificity makes it ideal for differentiating bacteria and viruses, by recognizing a unique 28-nucleotides sequence.



fluorescence

no fluorescence

E.coli Norovirus HCV no target target

We clearly differentiated

bacteria and viruses

suggested by our

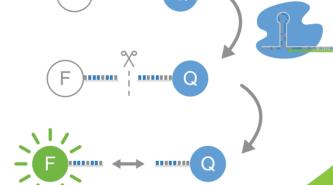
E.coli

Norovirus

HCV

bacterial pathogens and clear differentiation bet-

Fluorescence of RNase

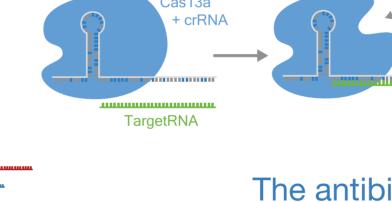


Paper support optimization

and toxic to Cas13a

Cas13a

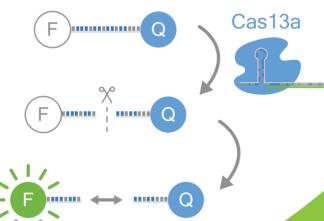
fighting the antibiotic crisis



The antibiotic resistance crisis

Readout reaction

Alert rises strongly after cleavage of the quencher.



Detection after amplification: in vitro

Sensitive and Specific Rapid: readout in < 30 minutes M. Robust: across experimenters Universal: any target sequence

Functionality of Cas13a

Cas13a Characterization

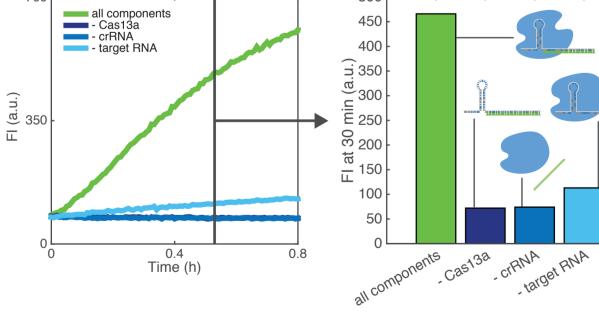
Target concentration (nM)

target source

in vivo

in vitro

in vivo

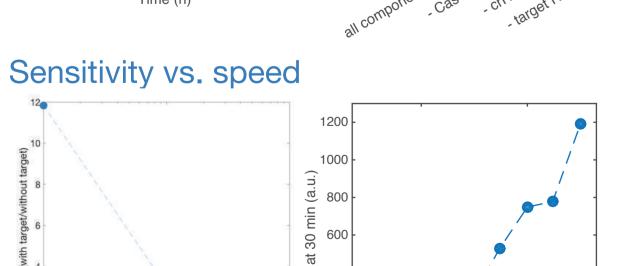


Gel for in vivo lysis + gel for RPA-TX

amplification

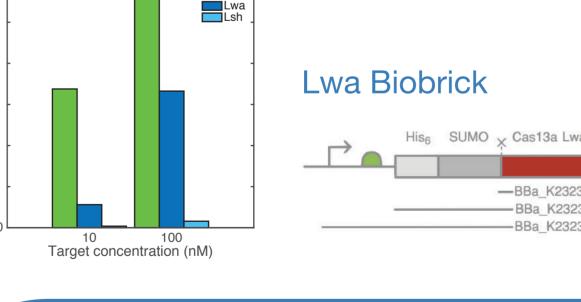
RPA-TX

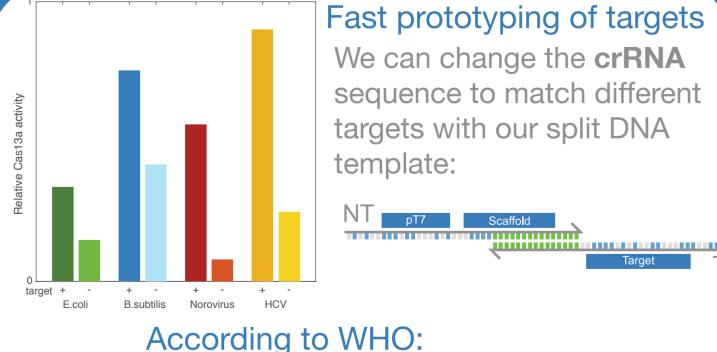
DNA



Cas13a sensitivity goes down with increasing concentration, while the kinetics of cleavge go up. We chose a compromise concentration of 10 nM Cas13a. The detection limit was then 10 nM target RNA.

Comparing different Cas13a

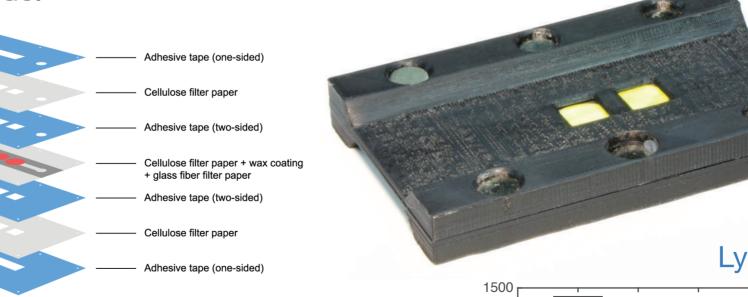




E. coli - 3rd most critical multiresistant strain B. subtilis - test for gram positive bacteria Norovirus - main cause for gastroenteritis HCV - 399 000 deaths per year

Paperstrip

Paperstrips make our test independent from expensive lab equipment, easy to use and storable. It also allows for multiplexing using paperfluidics. This allows for densely-packed, detectorindependent readout, similar to a QR-code.



We built a sample holder where the paperstrip is held. It has two windows to analyze a blank and the sample. It can be clipped into our fluorescence detector,

Lyophilization of Cas13a After lyophilization, the Cas13a

, Affordable

We used the design-test cycle and the "keep it

- nitrocellulose; most available but autofluorescent

- glass fiber paper; functional when treated with BSA

simple" principle to choose our paper support:

, Distributable

Scalable: paperfluidics

reaction mix is hardly functional. This step would need optimization, but cryoprotectants can help with stability.

Affordable \$0.85 per test 10 aM ensitive

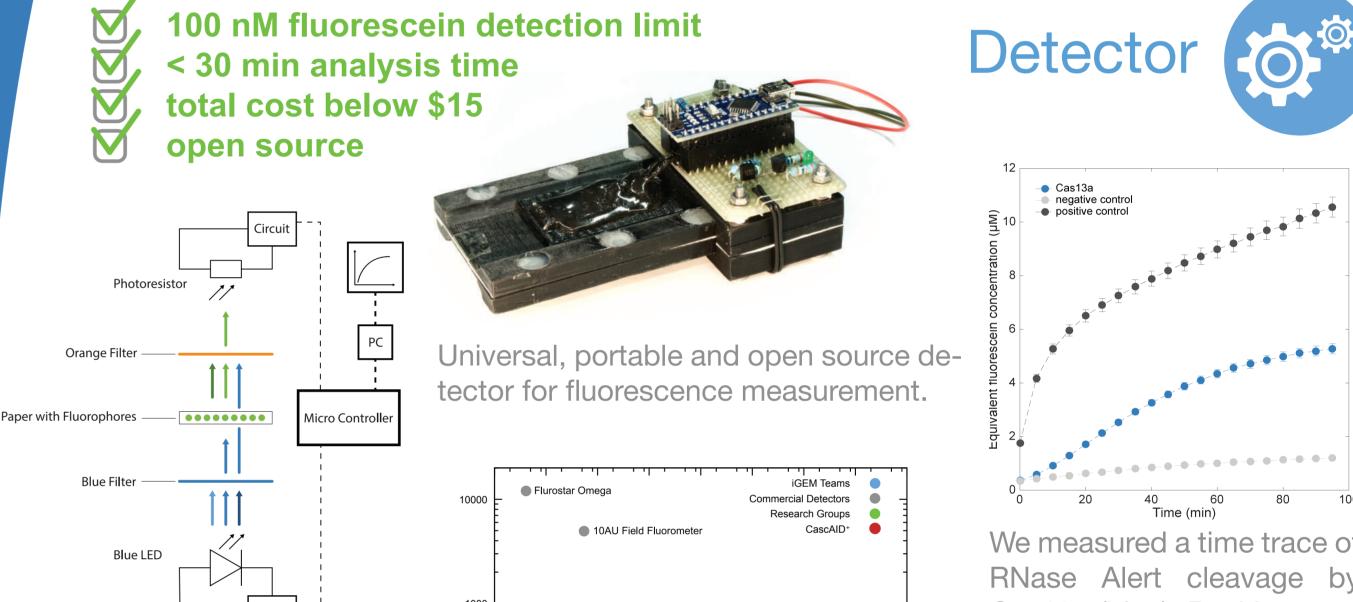
pecific strong target orthogonality **User-friendly** easy setup Rapid & Robust 30 min detection time no power outlet quipment-free fits in your pocket Deliverable

Results We successfully used Cas13a in a fluorescencebased assay on paper to distinguish viral and bacterial RNAs with a detection limit of 10 aM. We built the most affordable and sensitive fluorescence detector in iGEM, to our knowledge. Cost calculation

disposable components cost per test 0.310 \$ processing chip paper strip (incl. enymes & chemicals) 0.460 \$ cost per test¹

reusable components 0.014\$ flourescence detector 0.026\$ processing unit 0.007 \$ pressure supply 0.006\$ energy supply

0.030 \$ Rasberry Pi 0.853\$



We measured a time trace of RNase Alert cleavage by Cas13a (blue). Positive control is cleavage by RNase A, negative control is only RNase Alert.

Not only is our detector the most sensitive and least expensive one ever built by an iGEM team, it convinces with a price of less than 15 \$ at a detection threshold of 100 nM fluorescein.

Lukas Novak et al., 2006

Rafal Walczak, 2015

Feng-Bo Yang et al., 2009

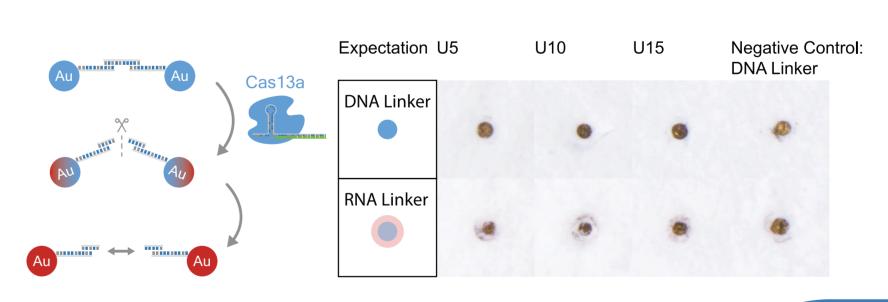
4 alternative readouts M, Functional AuNPs, Spinach-Aptamer Easy to interpret

Scheme of our detec-

tor. Light changes the

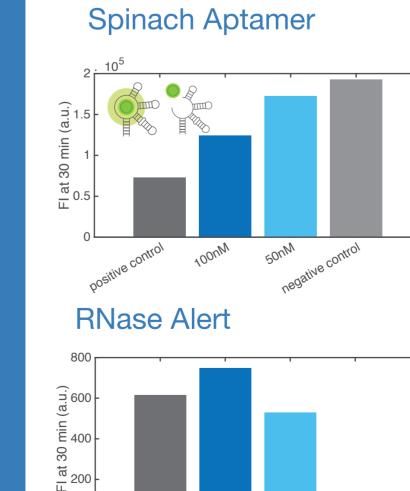
resistence of the pho-

toresistor.

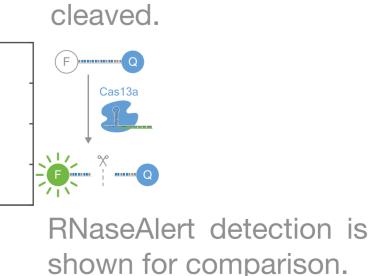


Gold nanoparticles (AuNP) allow for a colorimetric readout on paper. RNAcrosslinked AuNPs are dispersed by RNase activity, leading to a shift in absorbance, and a visible signal. We show that our method works with RNaseA as a proof of principle.

Readout



The spinach aptamer in creases the fluorophore signal upon binding. Ac tivated Cas13a is then detected by a decrease in fluorescence, when the RNA binding site is cleaved.



sing the detection sensitivity. ssDNA amplification T7 - Promotor aeBlue d cloning hybrid binding

rimetric readouts that include an amplification step, increa-protein expression

We designed additional colo- Intein-extein

References

10¹ Cas13a C° (nM)

Gootenberg J.S. et al., Science (2017). East-Seletsky A. et al., Molecular Cell (2017). World Health Organization (2017). Paige J.S. et al., Science (2011). Pardee K. et al., Cell (2016). Carrilho E. et al., Analytical Chemistry (2009).

Who did the work



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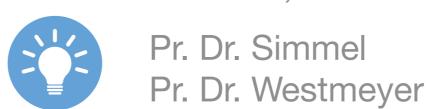


"Lightbringer".

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