

08. (August) 2019

Project: iGEM_Munich2019 Shared Project

Authors: Johanna Wallner

TUESDAY, 20/8/2019

Johanna

Cell culture: Reuptake assay transfection:

- 500 ng/well, triplicates, 24-well plate (coated with poly-L-Lysine)

Transfection scheme - reuptake assay 20/08/19

	condition	V8	V10 (L7Ae)	V11 (MCP)	V14 (MS2)	V15 (C/D Box)	V30
1	1	250 ng	-	-	-	-	250 ng
2	2	-	-	-	125 ng	-	375 ng
3	3	250 ng	-	125 ng	125 ng	-	-
4	4	250 ng	125 ng	-	-	125 ng	-

- 1: negative control, 2: positive control (fluc), 3: MCP/MS2-System, 4: L7Ae/C/D-Box-System
- exchange 550 µL medium
- Transfection:

Transfectionmix 24-well plate 20/0...

	A	B
1	DNA per well	500 ng
2	P3000 Reagent per well	1 µL
3	Lipofectamine 3000 reagent per well	0.75 µL
4	OptiMEM per well	2 x 25 µL

- Transfect cells according to the following table. Use the indicated volume of DNA and P3000™ Reagent with each of the two volumes of Lipofectamine™ 3000 (when performing optimization). Each reaction mix volume is for one well and accounts for pipetting variations. Scale volumes proportionally for additional wells.

Timeline		Steps	Procedure Details (Two Reaction Optimization)			
Day 0	1	Seed cells to be 70-90% confluent at transfection	Component	96-well	24-well	6-well
	2	Dilute Lipofectamine™ 3000 Reagent to Opti-MEM™ Medium (2 tubes) – Mix well	Adherent cells	1-4 × 10 ⁵	0.5-2 × 10 ⁶	0.25-1 × 10 ⁷
	3	Prepare master mix of DNA by diluting DNA in Opti-MEM™ Medium, then add P3000™ Reagent – Mix well	Opti-MEM™ Medium	5 µL × 2	25 µL × 2	125 µL × 2
	4	Add Diluted DNA to each tube of Diluted Lipofectamine™ 3000 Reagent (1:1 ratio)	Lipofectamine™ 3000 Reagent	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL
Day 1	5	Incubate	Opti-MEM™ Medium	10 µL	50 µL	250 µL
	6	Add DNA-lipid complex to cells	DNA (0.5-5 µg/µL)	0.2 µg	1 µg	5 µg
	7	Visualize/analyze transfected cells	P3000™ Reagent (2 µL/µg DNA)	0.4 µL	2 µL	10 µL
			Diluted DNA (with P3000™ Reagent)	5 µL	25 µL	125 µL
Day 2-4			Diluted Lipofectamine™ 3000 Reagent	5 µL	25 µL	125 µL
			Component (per well)	96-well	24-well	6-well
			DNA-lipid complex	10 µL	50 µL	250 µL
			DNA amount	100 ng	500 ng	2500 ng
			P3000™ Reagent	0.2 µL	1 µL	5 µL
			Lipofectamine™ 3000 Reagent used	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL

- transfection time: 11:30 a.m.
- attention: no Biotin-tag (V27)

Alejandro

cell culture: Transfection for Purification and qPCR

- time finished: 13:20 p.m.
- note: in condition 6: 6 µL V10 were given mistakenly, not the supposed 5.25 µL
- 6-well plate

Transfectionmix 6-well plate 20/08...		
	A	B
1	DNA per well	2500 ng
2	P3000 Reagent per well	5 µL
3	Lipofectamine 3000 reagent per well	3.75 µL
4	OptiMEM per well	2 x 125 µL

- Transfect cells according to the following table. Use the indicated volume of DNA and P3000™ Reagent with each of the two volumes of Lipofectamine™ 3000 (when performing optimization). Each reaction mix volume is for one well and accounts for pipetting variations. Scale volumes proportionally for additional wells.

Timeline		Steps	Procedure Details (Two Reaction Optimization)			
Day 0	1	Seed cells to be 70-90% confluent at transfection	Component	96-well	24-well	6-well
	2	Dilute Lipofectamine™ 3000 Reagent to Opti-MEM™ Medium (2 tubes) – Mix well	Adherent cells	1-4 × 10 ⁵	0.5-2 × 10 ⁶	0.25-1 × 10 ⁷
	3	Prepare master mix of DNA by diluting DNA in Opti-MEM™ Medium, then add P3000™ Reagent – Mix well	Opti-MEM™ Medium	5 µL × 2	25 µL × 2	125 µL × 2
	4	Add Diluted DNA to each tube of Diluted Lipofectamine™ 3000 Reagent (1:1 ratio)	Lipofectamine™ 3000 Reagent	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL
Day 1	5	Incubate	Opti-MEM™ Medium	10 µL	50 µL	250 µL
	6	Add DNA-lipid complex to cells	DNA (0.5-5 µg/µL)	0.2 µg	1 µg	5 µg
	7	Visualize/analyze transfected cells	P3000™ Reagent (2 µL/µg DNA)	0.4 µL	2 µL	10 µL
			Diluted DNA (with P3000™ Reagent)	5 µL	25 µL	125 µL
Day 2-4			Diluted Lipofectamine™ 3000 Reagent	5 µL	25 µL	125 µL
			Component (per well)	96-well	24-well	6-well
			DNA-lipid complex	10 µL	50 µL	250 µL
			DNA amount	100 ng	500 ng	2500 ng
			P3000™ Reagent	0.2 µL	1 µL	5 µL
			Lipofectamine™ 3000 Reagent used	0.15 and 0.3 µL	0.75 and 1.5 µL	3.75 and 7.5 µL

- Transfection scheme

Transfection scheme - Purification and qPCR 20/08/19 in ng per well											
	condition	wells	V8	V10	V11	V14	V15	V27	V28	V30	
1	2	3	1000 ng	-	-	-	-	-	-	1500 ng	
2	3	2	1000 ng	-	500 ng	500 ng	-	500 ng	-	-	
3	4	1	1000 ng	-	500 ng	-	500 ng	500 ng	-	-	
4	5	2	1000 ng	500 ng	-	-	500 ng	500 ng	-	-	
5	6	1	1000 ng	500 ng	-	500 ng	-	500 ng	-	-	
6	3H	2	1000 ng	-	500 ng	500 ng	-	-	500 ng	-	
7	5H	2	1000 ng	500 ng	-	-	500 ng	-	500 ng	-	

- qPCR-plate: 1 well with cond. 2, 3, 4, 5, 6 each
- purification-plates
 - scAvidin: 1 well with cond. 2, 3, 5 each
 - His: 1 well with cond. 2, 2 wells with cond. 2H and 5H each

Agar plate

- Chloramphenicol (3 plates; 25 µg/mL)
- Ampicillin (8 plates; 1:500 Ampicillin)