

Cell Experiment Record

SYSU-China 2019

19.6.6. 19:00

The first day of culturing cells began!

Passage a bottle of HEK293T cells

19.6.14 20:21

Transfecte cells

The transfection experiment was set up in 5 groups, and each group had four sets of parallel experiments;

1.endo-pcDNA3.1-L7Ae+DMEM

2.endo-pcDNA3.1-L7AeMUT+DMEM

3.pcDNA3.1+DMEM

4.DMEM

5.DC- (no cells)

The systems are as follows (unit μL):

pcDNA3.1-L7Ae	1.5
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HD	1.2
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<u>DMEM</u>	<u>97.3</u>
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Total	100
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pcDNA3.1-L7AemuT	1.5
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HD	1.2
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<u>DMEM</u>	<u>97.3</u>
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Total	100
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pcDNA3.1	1.5
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HD	1.2
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<u>DMEM</u>	<u>97.3</u>
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Total	100
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HD	1.2
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<u>DMEM</u>	<u>98.8</u>
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Total	100
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Leave at room temperature for 10min. Take a 96-well plate, when the cells account for about 80%, add the corresponding 5 μL transfection agent to each well, blow and mix well, add 100 μL DC- to each well in the rightmost row, and mark the group on the 96-well plates. Culture at 37°C for cytotoxicity test.

19.6.15

Cytotoxicity Test of L7Ae and L7Ae-MUT

Add 20 μ L of MTS to each well and measure the absorbance with a microplate reader.

19.7.14

First attempt: Virus packaging

Conditions: 9×10^5 cell, 3mL /well; 4ug Pacl-pAdEasy

19.7.18

Transfect 293T to detect the efficiency of Tet-on system and L7Ae/ L7Ae-Mut-Kturn system.

The systems are as follows:

Dox 浓度 ng/ml	0	10	30	50	100	200	300	400	600
Dox 加量 μ L	0	0.1	0.3	0.5	1	2	3	4	5
phEF1a-rtTA3	1 μ g								
pTRE-EBFP2	1 μ g								
Fugene	6 μ L								
HEK 293	200,000 cells								

	1xK-turn			2xK-turn		
	+L7AeMUT	+L7Ae	-L7Ae	+L7AeMUT	+L7Ae	-L7Ae
pcDNA-L7Ae		1 μ g			1 μ g	
pcDNA-L7AeMUT	1 μ g			1 μ g		
pcDNA-2Kt-eGFP				1 μ g	1 μ g	1 μ g
pcDNA-K1-eGFP	1 μ g	1 μ g	1 μ g			
pcDNA3.1			1 μ g			1 μ g
Fugene	6 μ L					
HEK-293	200,000 cells					

19.8.9

Adenovirus titer test and detection of the efficiency of Tet-on system.

19.9

Second attempt: Virus Packaging

19.10.9

1. Resuscitate cryopreserved HeLa

The storage tube was taken out in the liquid nitrogen tank No. 1. Wrap in plastic gloves and melt at 37 ° C for 4 min (do not shake). Transfer to a 15mL centrifuge tube, centrifuge at 1000rpm for 3min, remove the supernatant (note that it is removed to prevent DMSO residue and affect the subsequent growth of the cells). Resuspend with 2 mL of DC-, add 1 mL to a culture flask containing 5 mL of medium, shake well, and incubate at 37 ° C. Need to change the medium the next day.

19.10.10

Lipo2000 transfection.

The systems are as follows:

pTRE-EBFP	6ug
phEF1a-rtTA	6ug
<u>DMEM</u>	to 300uL.....(1)

pTRE-EBFP	6ug
pcDNA-rtTA	6ug
<u>DMEM</u>	to 300uL.....(2)

Lipo2000	24uL
<u>DMEM</u>	576uL
Total	600uL.....(3)

(1) (2) are mixed with 300uL of (3), separately, and place at room temperature for more than 10min.

Dox gradient: 0-100-200-...-700 ng/uL

Add a Tet Positive group (working concentration: 125ng/uL) and a Negative group (nothing is added in.)

19.10.18

Lipo2000 transfect HEK293 to test the system efficiency

phEF1 α -rtTA3 +pTRE-L7Ae-miR592(different concentration to set experiment group)+pTRE-2Kt-EGFP