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04 Transformation

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1 Works for me dx.doi.org/10.17504/protocols.io.49ngz5e
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MATERIALS

| NAME  | CATALOG #  | VENDOR  |
|--|---|--|
| Shaker incubator | | |
| Ice | | |
| Centrifuge | | |
| Competent Cells | | |
| LB medium | / | |

BEFORE STARTING

Preparation competent cells.

1 Preparation of competent cells



1. Streak out the E.coli strain on an LBM plate (no ampicillin!) to isolate colonies and incubate at 37 degrees C overnight (16-20 hours).
2. Use a sterile inoculating loop to collect cells from a single colony and inoculate 50 ml sterile 1X LBM Grow at 37 degrees C overnight (16-20 hours) in a shaker incubator.
3. Add 25 ml of the overnight culture to each 250 ml LBM flask.
4. Grow the cultures to OD600 = 0.5
5. Decant supernatant and resuspend the cells in 1/4 original volume (87.5 ml) ice cold 100 mM MgCl₂. Hold on ice for 5 minutes. Transfer the cells to pre-chilled sterile large centrifuge bottles. Spin in the for 10 minutes using the rotor 4000 rpm at 4 degrees C.
6. Decant the supernatant and resuspend the cells in 1/20 original volume (17.5 ml) of ice cold 100 mM CaCl₂. Hold on ice for 20 minutes. Pellet as above 4000 rpm for 10 minutes.
7. Decant the supernatant and resuspend the cell pellet in 1/100 original volume (3.5 ml) of a solution that is 85% v/v 100 mM CaCl₂ and 15% v/v glycerol (100%). For each culture processed chill approximately 15 labeled eppendorf tubes in a dry ice-EtOH bath. Pipet 300 ul cells into each tube and place immediately into the dry ice-EtOH bath.
8. Transfer the frozen competent cell aliquots to -80 degrees C.



- 2 Take competent cells out of -80°C and thaw on ice (approximately 20-30 mins).


 -80 °C




- 3 Remove agar plates (containing the appropriate antibiotic) from storage at 4°C and let warm up to room temperature and then (optional) incubate in 37°C incubator.


 37 °C

- 4 Mix 1 μl of DNA (usually 10 pg - 100 ng) into competent cells. Gently mix by flicking the bottom of the tube with your finger a few times. Incubate the competent cell/DNA mixture on ice for 20-30 mins.
 10 ng ~  100 ng



- 5 Heat shock each transformation tube by placing the bottom 1/2 to 2/3 of the tube into a 42°C water bath for 90 secs.
 00:01:30
 42 °C

- 6 Put the tubes back on ice for 2 min.
 00:02:00

- 7 Add 600 μl LB media (without antibiotic) to the bacteria and grow in 37°C shaking incubator for 45 min.
 600 μl
 00:45:00
 37 °C

- 8 The bacterial liquid was centrifuged at 3500rpm for 3 minutes, 400 microliters of supernatant was discarded, and the bacterial liquid was suspended again.
 00:03:00

- 9 Plate the transformation onto a LB agar plate containing the appropriate antibiotic.

- 10 Incubate plates at 37°C overnight.
 37 °C
 12:00:00



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