# Resuspension of Primers

## Overview

This protocol covers resuspension of dried (lyophilized) oligos that are to be used as primers. In this protocol both a stock (100μM) and working (10μM) solutions are made.

## Materials

- Lyophilized Oligos (IDT)
    - Each oligo should have a corresponding specification sheet, which should be stored in the specification sheet drawer.
- Labeled 1.7mL centrifuge tubes (1 per sample)
    - Tubes should be labeled with primer name and 10μM, dates and initials should be included on the side
- Nuclease Free Water (NFW)[i]

## Procedure

1. Briefly spin down oligo tubes.
    - Use green benchtop centrifuge
2. Determine correct amount of NFW to add to each tube for a 100μM solution. This number can be found on the specification sheet. Alternatively, it can be determined by multiplying the number of nanomoles of DNA provided by 10 and then using that many μl of NFW. Remember that each tube will have a different amount of NFW to add.
3. Add correct amount of NFW to each tube. Heat at 55C for 2 minutes. Vortex. Briefly spin down.
4. Prepare a 10μM working stock by adding a NFW and 100μM primer at a ratio of 9:1 (9μl NFW for every 1μl primer). Typically we make a 100μl worth of working solution.
5. Store primers in appropriate boxes in -20. Log their locations on google drive. Update the location column on appropriate primer excel sheet in the inventory folder on Dropbox.

[i] Other labs might use TE buffer, which is better for long term storage. As we've never had a problem with degradation, and switching to TE could conceivably cause minor problems downstream (from EDTA), we use NFW. Additionally, switching now would create an inconsistency.