



De novo macrocyclic peptide modulators of protein-carbohydrate interactions



Seino A. K. Jongkees,^{1,2} Sami Caner,³ Christina Tysoe,^{4,5} Lieke van Gijzel,¹ Gary Brayer,³ Stephen Withers,^{3,4,5} Hiroaki Suga²

1. Department of Chemical Biology and Drug Discovery, Utrecht Institute of Pharmaceutical Sciences, Utrecht University, 2. Department of Chemistry, Tokyo University, Tokyo, Japan

3. Department of Biochemistry and Molecular Biology, University of British Columbia, Vancouver, Canada, 4. Department of Chemistry, University of British Columbia, Vancouver, Canada, 5. Centre for High-Throughput Biology, Michael Smith Laboratories, Vancouver, Canada.

Overview

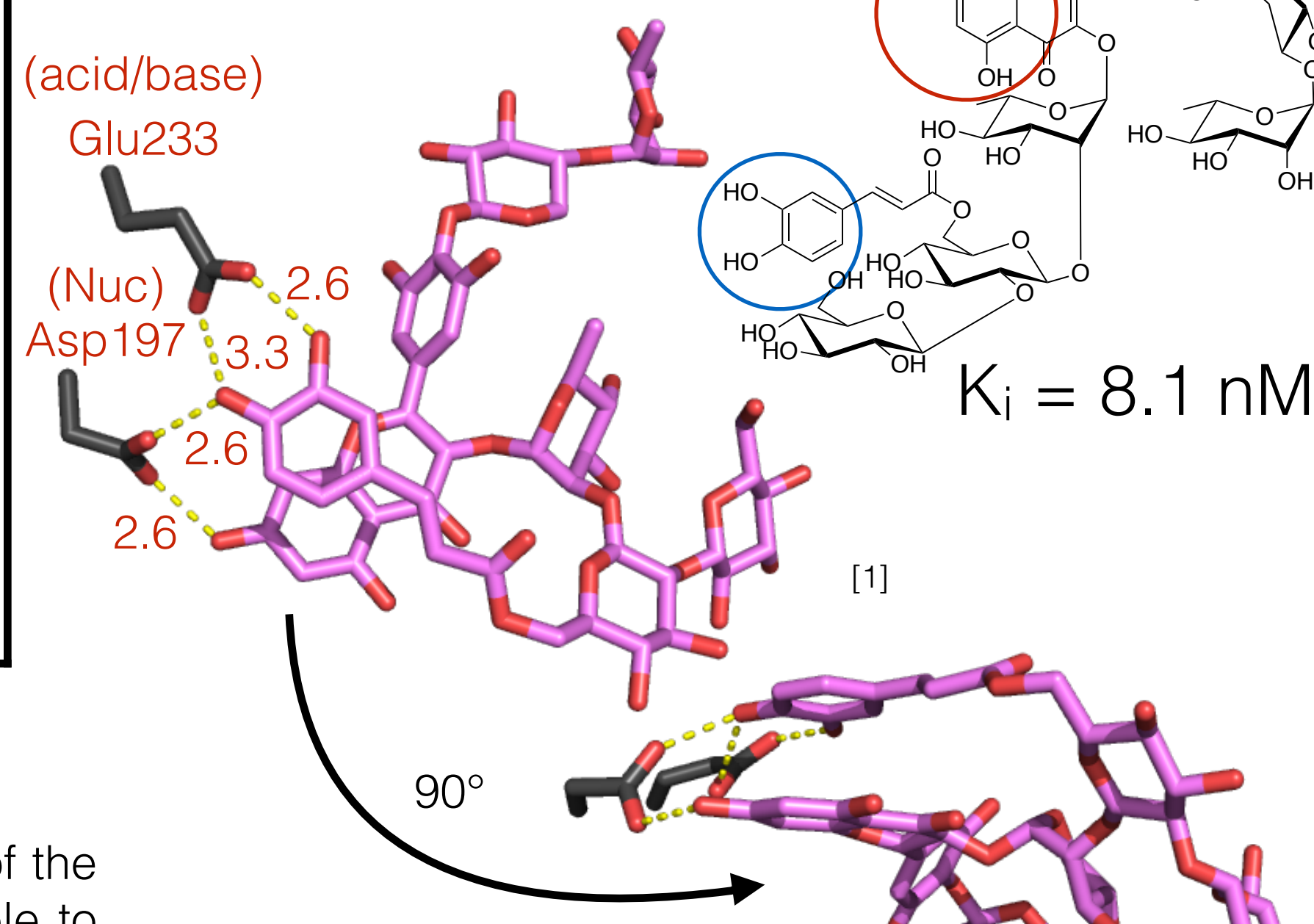
- Proteins play an important role in carbohydrate biology, particularly in their recognition and metabolism.
- Protein-carbohydrate interactions are often weak, taking place over a large interaction surface and/or with multivalency.
- Small molecules can have trouble binding tightly to such surfaces, while peptides are more well suited.
- The RaPID system is able to discover new peptide ligands, but has not yet been applied to carbohydrate binding sites.
- Proof-of-principle using human pancreatic α -amylase.
- Mimic MbA interactions in a new scaffold, make it more general.

Human pancreatic α -amylase

Selective inhibition of HPA, at the start of the starch digestion pathway, should be able to provide control of blood glucose spikes in type 2 diabetes without the side-effects of current-generation α -glucosidase inhibitors.

maltase-glucoamylase
sucrase-isomaltase
 α -glucosidases

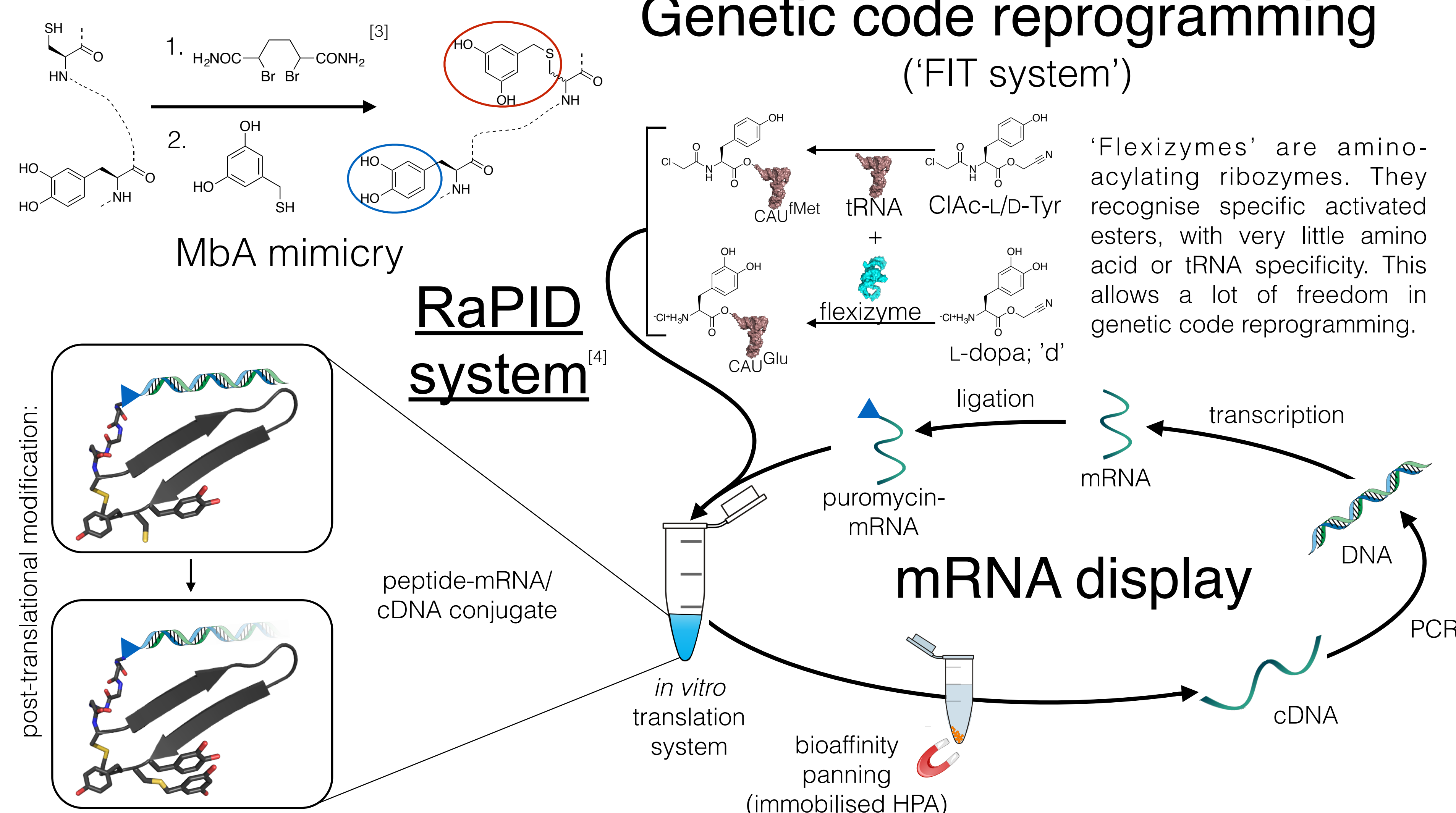
Montbretin A



Montbretin A (MbA) is a natural product HPA inhibitor composed of carbohydrate and phenol moieties, identified from a high-throughput screen.^[2] Crystallography revealed a new interaction motif with the catalytic residues, with caffeic acid and myricetin stacking to provide the majority of the inhibition.

Genetic code reprogramming ('FIT system')

'Flexizymes' are amino-acylating ribozymes. They recognise specific activated esters, with very little amino acid or tRNA specificity. This allows a lot of freedom in genetic code reprogramming.



Selection results

piHA-Dm^[5]

($K_i = 7.0$ nM)

Consensus motif, D-Tyr initiated library
Y PYSCw RH C

piHA-L5(d10Y)

($K_i = 9.2$ nM)

Consensus motif, L-Tyr initiated library
Y GHSHIRFGYSYHVSICG-NH₂

Selectivity

Enzyme	Inhibition (IC ₅₀ , nM)
<i>Sus domesticus</i> (porcine) pancreatic α -amylase	8.5 $\pm$ 0.9
<i>Homo sapiens</i> (human) salivary α -amylase	34 $\pm$ 6
<i>Butyrivibrio fibrisolvens</i> α -amylase	NI
<i>Aspergillus oryzae</i> α -amylase	NI
<i>Homo sapiens</i> (human) maltase-glucoamylase	520 $\pm$ 10
<i>Mus musculus</i> (mouse) maltase-glucoamylase	990 $\pm$ 40
<i>Saccharomyces cerevisiae</i> (yeast) α -glucosidase	NI
<i>Agrobacterium faecalis</i> β -glucosidase	NI
<i>Bos taurus</i> (bovine) liver β -galactosidase	NI
<i>Coffea arabica</i> (green coffee bean) α -galactosidase	NI
<i>Canavalia ensiformis</i> (Jack bean) α -mannosidase	NI

Diversity

'L1' (1.9 nM):
YPTKRYGQWLPIYRNNCG-NH₂

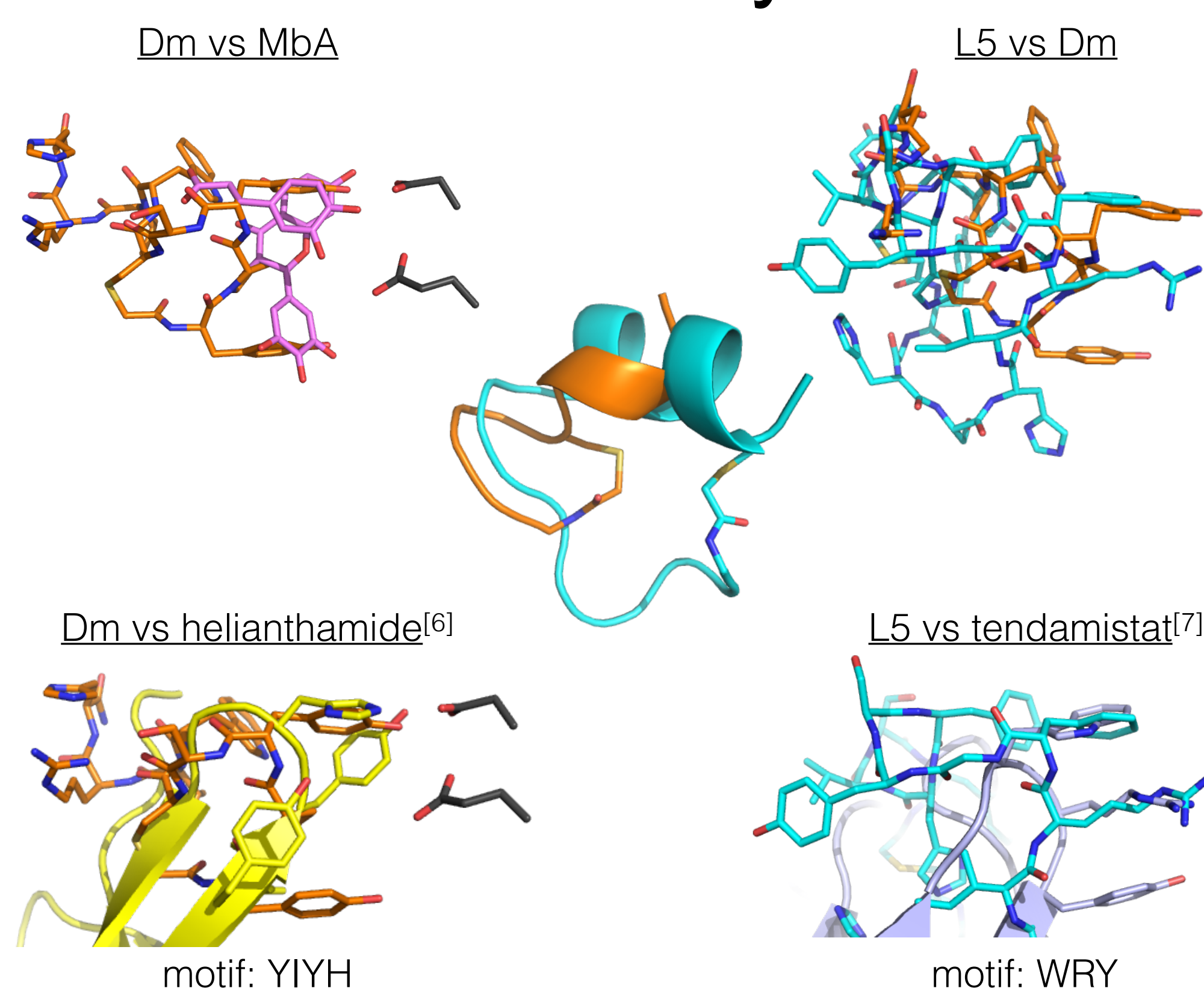
'L3' (2.6 nM):
YWDRPTRFGYAYSIVIYCG-NH₂

'L5' (2.8 nM):
YGHSHIRFGdSYHVSICG-NH₂

'L12' (2.5 nM):
YTFRDWRRSYGGITVRCG-NH₂

'L26' (1.0 nM):
YQGSHSAWCRWINdNPdCG

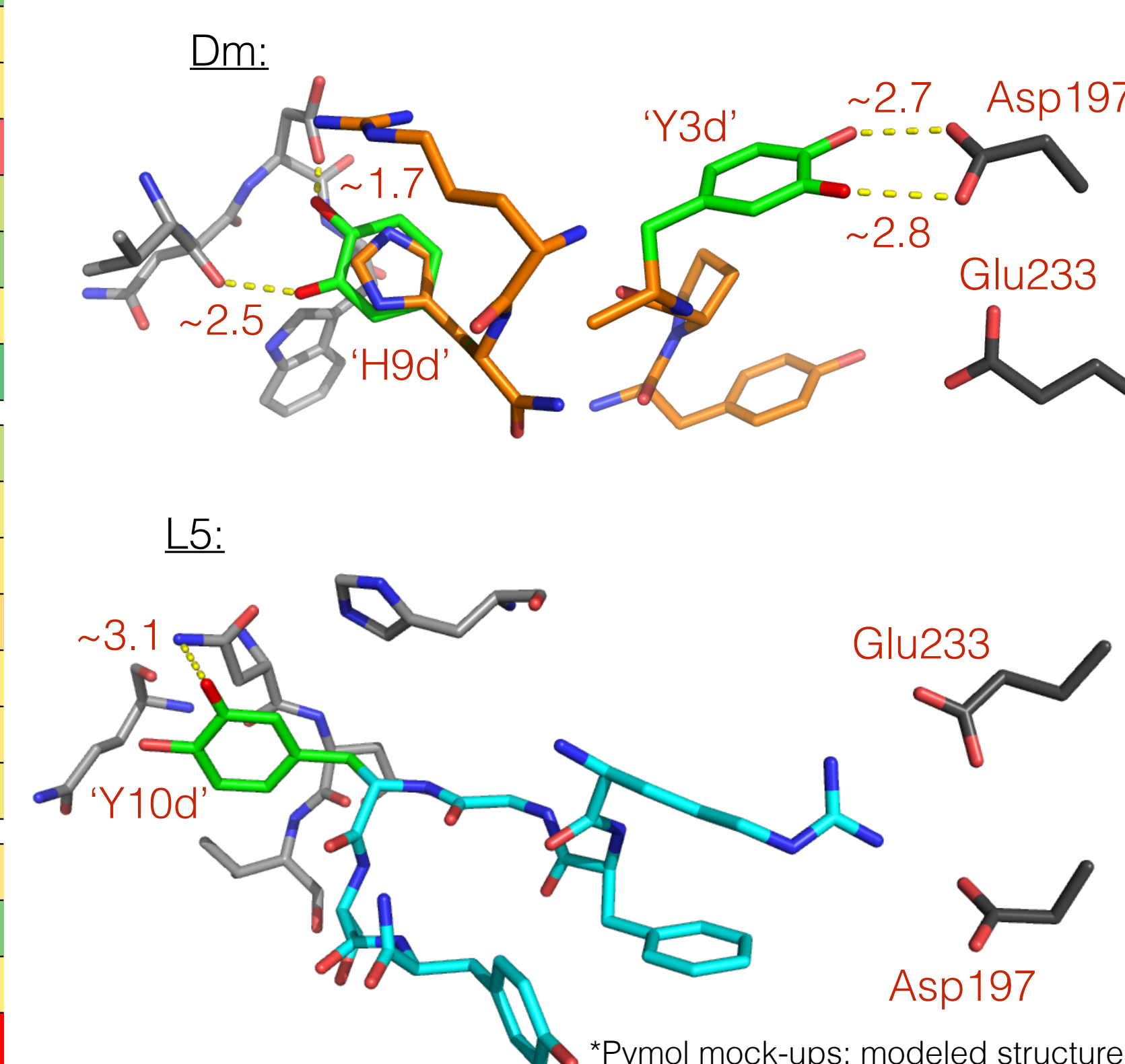
Overlays



Mutagenesis

Code	Sequence	K _i (nM)
D1	yPYSCwARHVRIREN	1
D3	yPYSCwVRHSDPHKE	2.7
Dm (D3A10)	yPYSCwVRH	7
Dm-lin	yPYSAWVRH (linear)	200
Dm-V7A	yPYSCwARH	2
D1-D11	yPYSCwARHV	1.3
Dm-H9d	yPYSCwVRd	3.7
Dm-Y3d	yPdSCwVRH	0.5
L1	YPTKRYGQWLPIYRNNCG	1.9
L3	YWDRPTRFGYAYSIVIYCG	2.6
L3-F8Y	YWDRPTRYGYAYSIVIYCG	10
L3-Y10d	YWDRPTRYGdAYSIVIYCG	31
L5	YGHSHIRFGdSYHVSICG	2.8
L5-d10Y	YGHSHIRFGYSYHVSICG	9.2
L12	YTFRDWRRSYGGITVRCG	2.5
L26-A17	YQGSHSAWCRWINdNP	12
L26-A17-d14Y	YQGSHSAWCRWINYNP	1
L26-A14	YQGSHSAWCRWIN	2.9
L26-A10	YQGSHSAWC	>10000

Possible L-dopa interactions



Conclusions

- The first use of the RaPID system with a carbohydrate-active enzyme was successful.
- Potent macrocyclic peptide inhibitors of human pancreatic α -amylase discovered.
- Most promising hit is piHA-Dm, a nonapeptide with 7 nM K_i .
- Selective for pancreatic α -amylase over all other enzymes tested.
- Forms a compact fold, with the 'tail' forming a 3₁₀ helix.
- L-Dopa reintroduced based on structure and high throughput sequencing data; 500 pM inhibitor.
- L-Tyr initiated motif less conserved, but equally potent.
- Also helical, but different arrangement and interactions.
- No MbA-like motif, despite both moieties in library.
- Interactions with human pancreatic α -amylase converge on known as well as new motifs, in a new scaffold.



JSPS

References

- Williams, L.K., Zhang, X., Caner, S., Tysoe, C., Nguyen, N.T., Wicki, J., Williams, D.E., Coleman, J., McNeill, J.H., Yuen, V., et al. (2015). *Nat. Chem. Biol.* 11, 691–696.
- Tarling, C.A., Woods, K., Zhang, R., Brastianos, H.C., Brayer, G.D., Andersen, R.J., and Withers, S.G. (2008). *ChemBioChem* 9, 433–438.
- Jongkees, S.A.K., Umemoto, S., and Suga, H. (2017). *Chem. Sci.*, in press (DOI: 10.1039/C6SC04381J)
- Hipolito, C.J., and Suga, H. (2012). *Curr. Opin. Chem. Biol.* 16, 196–203.
- Jongkees, S.A.K., Caner, S., Tysoe, C., Brayer, G.D., Withers, S.G., and Suga, H. (2017). *Cell Chem. Biol.*, manuscript accepted.
- Tysoe, C., Williams, L.K., Keyzers, R., Nguyen, N.T., Tarling, C., Wicki, J., Goddard-Borger, E.D., Aguda, A.H., Perry, S., Foster, L.J., et al. (2016). *ACS Cent. Sci.* 2, 154–161.
- Wiegand, G., Epp, O., and Huber, R. (1995). *J. Mol. Biol.* 247, 99–110.



GlycoNet