

## LIGATION MATH

**Construct:** pBAD-ompT-CAM-GS-xylE-xylF

**Fragment 1:** pBAD-ompT-CAM-GS

**Fragment 2:** XylE

**Fragment 3:** XylF1/2

**Fragment 4 :** XylF1/2

## FORMULA

Required mass insert (g) = desired insert/vector molar ratio x mass of vector (g) x  
ratio of insert to vector lengths

**Vector (pSB1C3) : 25 ng/μl**

**pBAD-ompT-CAM-GS: 50 ng/μl**

**XylE: 50 ng/μl**

**XylF1/2: 50 ng/μl**

**XylF1/2: 50 ng/μl**

**Ratio of Insert/Vector - 6:1**

Fragment	Length (bp)	Insert mass (ng)	Required volume of fragment in μl
pBAD-ompT-CAM-GS	716	51.88	1.04
XylE	963	69.78	1.40
XylF1/2	552	40.00	0.80
XylF1/2	516	37.39	0.75

## CLONING PLAN

Fragments to be Digested and Ligated,

Fragment	Size (bp)
pSB1C3	2070
pBAD-ompT-CAM-GS	716
XylE	963
XylF1/2	552
XylF1/2	516

**Reconstitution:**

**pBAD\_RBS\_ompT\_CAM\_GS: 500 ng (Given)**

**XylE: 1000 ng (Given)**

**XylF1/2: 500 ng (Given)**

**XylF1/2: 500 ng (Given)**

Suspend **500 ng of DNA in 10  $\mu$ l of TE buffer and 1000 ng of DNA in 20  $\mu$ l of TE buffer** to get a final concentration of 50 ng/ $\mu$ l. Incubate the reconstituted DNA at 50°C for 20 minutes and store at 4°C.

**Digestion:**

**pSB1C3 ( Backbone)**

**Stock: 25 ng/ $\mu$ l**

Molecular water	3 $\mu$ l
10x Neb buffer 2.1	1 $\mu$ l

DNA	5 $\mu$ l
Restriction enzyme	0.5 $\mu$ l EcoR1 + 0.5 $\mu$ l Pst1
Total Volume	10 $\mu$ l

**pBAD\_RBS\_ompT\_CAM\_GS**

**Stock: 50 ng/ $\mu$ l**

Molecular water	5 $\mu$ l
10x Neb buffer 2.1	1 $\mu$ l
DNA	3 $\mu$ l
Restriction enzyme	0.5 $\mu$ l EcoR1 + 0.5 $\mu$ l Bsa1
Total Volume	10 $\mu$ l

**XyIE**

**Stock: 50 ng/ $\mu$ l**

Molecular water	1.5 $\mu$ l
10x Neb buffer 2.1	1.0 $\mu$ l
DNA	7.0 $\mu$ l
Restriction enzyme	0.5 $\mu$ l Bsa1
Total Volume	10 $\mu$ l

**XyIF1/2****Stock: 50 ng/μl**

Molecular water	4.5 μl
10x Neb buffer 2.1	1.0 μl
DNA	4.0 μl
Restriction enzyme	0.5 μl Bsa1
Total Volume	10 uL

**XyIF1/2****Stock: 50 ng/μl**

Molecular water	4.25 μl
10x Neb buffer 2.1	1.0 μl
DNA	3.75 μl
Restriction enzyme	0.5 μl Bsa1 + 0.5 μl Pst1
Total Volume	10 μl

1. Prepare digestion mixture for backbone and the fragments in separate tubes.
2. Digest the mixture at 37°C celsius for 30 minute.
3. Inactivate the restriction enzymes at 80°C for 20 minute.

**Ligation:**

Molecular water	1.67 $\mu$ l
T4 DNA ligase buffer (10X)	1.5 $\mu$ l
Fragment 1	3.33 $\mu$ l
Fragment 2	2 $\mu$ l
Fragment 3	2 $\mu$ l
Fragment 4	2 $\mu$ l
pSB1C3	2 $\mu$ l
T4 DNA ligase	0.5 $\mu$ l
Total Volume	15 $\mu$ l

1. Prepare the ligation mixture and gently mix by pipetting up and down and microfuge briefly.
2. Incubate the ligation mixture at 25°C for 10 minutes and transform 2  $\mu$ l of ligated DNA into 50  $\mu$ l competent cell.
3. Incubate the rest ligation mixture at 16°C overnight and transform 2  $\mu$ l of ligated DNA into 50  $\mu$ l competent cell next day.