Week of Oct 7

WEDNESDAY, 10/10/2018

Origin Tests: S. mel, L. lac, C. glut

1. Transformed 200 ng of RSF1010, pBBR1 and pWV01 plasmid into all 3 strains
    a. transform by electroporation;
        I. 2.5 kV for S. mel and C. glut
        II. 2 kV for L. lac
    b. recovered in plain LB for S. mel; universal medium + glucose for C. glut; universal medium + sucrose for L. lac; all in 30C shaking incubator
        I. 3 hrs for S. mel
        II. 2 hrs for L. lac
        III. 1 hr for C. glut
2. Plated L. lac and C. glut on both plain and Kan50 LB plates; also plated L. lac onto m17 + Kan50 plate; S. mel plated on plain and Neo100 LB agar plate
3. All strains in 30C incubator;

THURSDAY, 10/11/2018

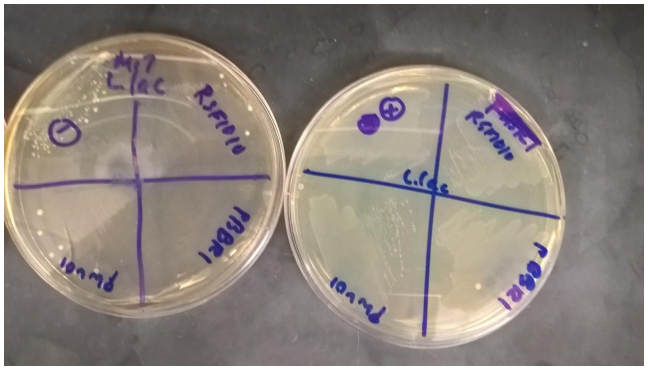
FRIDAY, 10/12/2018

Results of Origin Tests

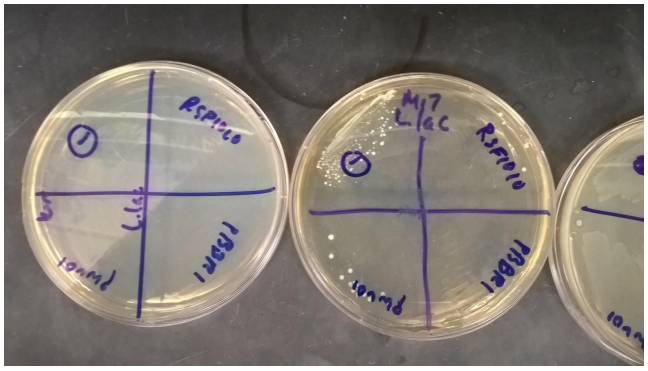
L. lac

1. Some growth on Kan+M17 Agar plate; growth on plain LB agar plate; no growth on Kan plate

Origin testing L. lac Final.jpg



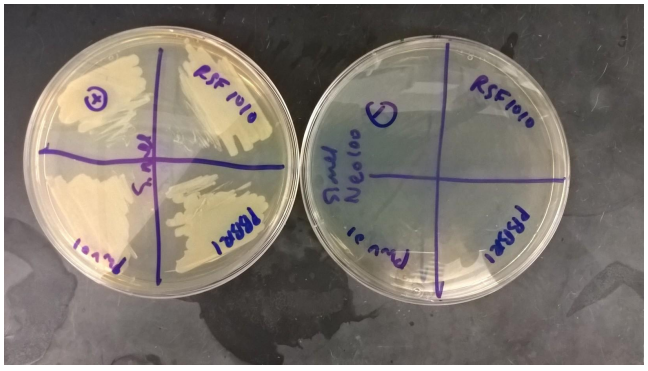
Origin testing L. lac Final 2.jpg



S. mel

1. Some growth plain LB plate; no growth on Neo plate

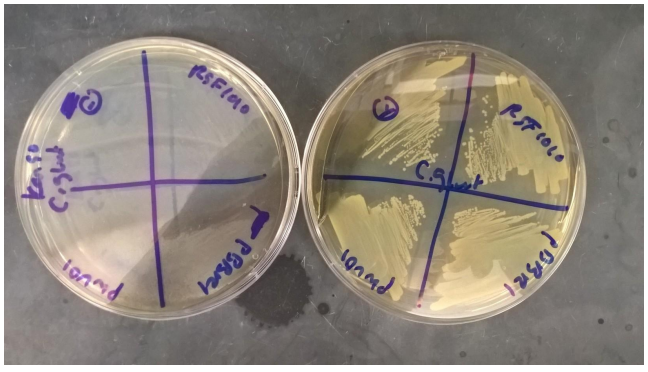
Origin testing S. mel final.jpg



C. glut

1. Growth on plain agar plate; dried out Kan plate

Origin testing C. glut Final.jpg



Origin Testing: Acinetobacter baylyi

1. PIcked colony of A. b. from plate Anna made into 4 mL of LB
2. Let grow overnight in 30C shaking incubator

SATURDAY, 10/13/2018

Origin Testing: Acinetobacter baylyi

1. Put 1 mL of LB, 70 uL of A. b, and 100 ng of either RSF1010, pWV01, or pBBR1 into Falcon tube
2. Incubated tube for 3 hrs
3. Plated 25 uL of each transformation + non-transformed A. b. onto Kan50 and plain LB plate; spread with sterile inoculation loop
4. Put in 30C incubator overnight

SUNDAY, 10/14/2018

Origin Testing Results: Acinetobacter baylyi

1. pWV01 on Kan50 plate grew; no other growth on Kan50 plate; all transformants grew on plain LB plate
2. Weird; was supposed to work with RSF1010; is our RSF1010/pBBR1 DNA bad?
    a. test w/ E. coli?

Origin testing A. baylyi.jpg