ZymoPURE™ Plasmid Midiprep

Catalog Nos. D4200 & D4201

Quick Protocol

- Buffer Preparation: Add 38 ml of 95% ethanol to the 10 ml ZymoPURE™ Wash 2 (Concentrate) (D4200), or add 88 ml of 95% ethanol to the 23 ml ZymoPURE™ Wash 2 (Concentrate) (D4201) before use.
- The ZymoPURE™ P2 and ZymoPURE™ Binding Buffer may have precipitated. If this occurs, dissolve the precipitate by incubating the bottles at 30-37 °C for 10-20 minutes and mix by inversion. Do not microwave!
- Before Starting: Centrifuge up to 50 ml of bacterial culture at ≥ 3,400 x g for 10 minutes to pellet the cells in a 50 ml conical tube. Discard supernatant.

The following procedure should be performed at room temperature (15-30°C).

- Add 8 ml of ZymoPURE™ P1 (Red) to the bacterial cell pellet and resuspend completely by vortexing or pipetting.
- Add 8 ml of ZymoPURE™ P2 (Green) and immediately mix by gently inverting the tube 6 times. Do not vortex! Let sit at room temperature for 2-3 minutes. Cells are completely lysed when the solution appears clear, purple, and viscous.
- Add 8 ml of ZymoPURE™ P3 (Yellow) and mix gently but thoroughly by inversion. Do not vortex! The sample will turn yellow when the neutralization is complete and a yellowish precipitate will form.
- 4. Ensure the plug is attached to the Luer Lock at the bottom of the ZymoPURE™ Syringe Filter. Place the syringe filter upright in a tube rack and load the lysate into the ZymoPURE™ Syringe Filter and wait 5-8 minutes for the precipitate to float to the top.
- Remove the Luer Lock plug from the bottom of the syringe and place it into a clean 50 ml conical tube. Place the plunger in the syringe and push the solution through the ZymoPURE™ Syringe Filter to clear the debris. Save the cleared lysate!
- Add 8 ml ZymoPURE™ Binding Buffer to the cleared lysate from step 5 and mix thoroughly by inverting the capped tube 8 times.

(Continued on reverse side)

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Vacuum Protocol

The vacuum pump should be a single or double-staged unit capable of producing up to 400 mm Hg pressure.

- Ensure the connections of the Zymo-Spin™ III-P Column Assembly are finger-tight and place onto a vacuum manifold.
- Add the entire mixture from step 6 into the Zymo-Spin™ III-P Column Assembly, and then turn on the vacuum until all of the liquid has passed completely through the column.
- Unscrew the purple Luer Lock cap from the top of the Zymo-Spin™ III-P Column and discard the Reservoirs.

Centrifugation Protocol

A swinging bucket rotor is required for centrifugation.

- 7. Remove the 50 ml Reservoir from the top of the Zymo-Spin™ III-P Column Assembly. Ensure the connection between the 15 ml Conical Reservoir and Zymo-Spin™ III-P column is finger-tight and place the assembly into a 50 ml conical tube.
- Add 10 ml of the mixture from step 6 into the 15 ml Conical
 Reservoir/Zymo-Spin™ III-P
 Column assembly, and centrifuge at 500 x g for 2 minutes.
- Empty the 50 ml conical tube and repeat step 9 until the entire sample has passed through the column.

Note: Steps 10-11 can also be completed using a microcentrifuge instead of the vacuum manifold (see full instruction manual).

- 10. With the vacuum off, add 800 μl of ZymoPURE™ Wash 1 to the Zymo-Spin™ III-P Column. Turn on the vacuum until all of the liquid has passed completely through the column.
- 11. With the vacuum off, add 800 µl of ZymoPURE™ Wash 2 to the Zymo-Spin™ III-P Column. Turn on the vacuum until all of the liquid has passed completely through the column. Repeat this wash step.
- 12. Place the Zymo-Spin[™] III-P Column in a **Collection Tube** and transfer to a microcentrifuge. Centrifuge at ≥ 10,000 x g for 1 minute in order to remove any residual wash buffer.
- 13. Transfer the Zymo-Spin™ III-P Column into a clean 1.5 ml tube and add 200 µl of ZymoPURE™ Elution Buffer directly to the column matrix. Incubate at room temperature for 2 minutes, and then centrifuge at ≥ 10,000 x g for 1 minute in a microcentrifuge.