Protocol
-Grow 10 mL cultures overnight in LB media with 10 uL of Chloramphenicol
-Transfer an aliquot of the overnight cultures to fresh culture LB media at a ratio of 1:200.
-Grow cultures until log phase ( $OD_{600}$ 0.4-0.8).
-Induce the expression of the genes with 1 mM IPTG (final concentration) and let grow for 4 additional hours
-Measure OD again and transfer approximately $10^8$ cells to 1.5 mL microcentrifuge tubes
-Spin the tubes down at 4,000 g for 20 min and remove supernatant.
-Wash cells with water and re-pellet at the same conditions
-Remove water easily with a pipette and dry pellets overnight with a SpeedVac
-Rehydrate dry pellets adding 1 mL of culture media and vortex
-Transfer cells to chloramphenicol plates and grow overnight at 37 ºC
-The following day, count the colonies and report the survival as colony forming units/$10^8$cells plated.

Additional information
For our assays on stabilization of proteins we were drying overnight in the SpeedVac (Eppendorf Vacufuge Concentrator, at a temperature of 30 ºC and the non-spinning mode for aqueous solutions). Therefore, it would be interesting if you could apply the same conditions for the drying.

Succes/Good luck/Suerte !

TU Delft iGEM team 2017

PS: Should you encounter any difficulties in the realisation of the experiments or was anything not clear enough, please feel free to contact me back to:
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+34 622 724 224 (emergencies)