B. Plasmid Transformation and Cultivating

Prepartation	② take the foam box, carrying about 2/3 volume of ice;
	② remove 100µL competent cells from the -80 °C refrigerator.B
Packing	① remove the alcohol lights, tweezers, sterile centrifuge tube and sterile tip;
	② ignite the alcohol lights, burning the tweezers 5s on it;
	③ remove the 1.5mL sterile centrifuge tube, cover the tube cover and put it on
	the tube box;
	4 open the centrifuge tube with competent cells, remove 50µL competent cells
	into the sterile centrifuge tube; do these beside the alcohol lights;
	⑤ put the above two centrifuge tubes equipped with competent cells in the ice
	box in the spare.
Adhesion	① use a sterile tip to remove 5µL plasmid on the plasmid tray, add it 50µL
	competent cells; do these beside the alcohol lights,
	② open the water bath, and set the temperature of 42 °C;
	③ put the centrifuge tube in the ice box, standing 30min;
Heat Shock	① put the ice-bath-after centrifuge tube in 42 °C water bath, heat shock 45s;
	② Remove the centrifuge tube, placed in the ice box 2min.
Recovery	① remove the non-antibiotic liquid medium;
	② use sterile tip to remove 500µL liquid medium and add to the centrifuge tube
	which has been heat shocked;
	③ put the centrifuge tube into the 37 °C shaker with the tube pad, incubated 1h.
Coat Dishes	① remove centrifuge tube from the shaker, centrifuge 7000rpm for 30s or
	5000rpm for 1min;
	② use sterile tips to abandon 400uL supernatant beside alcohol lights; use a
	pipette make remaining bacteria liquid well mixed;
	③ according to plasmid's resistance, remove the chloramphenicol resistant solid
	Packing Adhesion Heat Shock Recovery

		medium or ampicillin resistant solid medium;
		④ use sterile tips to take 100uL bacteria liquid mentioned in step 3 then add to
		the surface of solid medium beside alcohol lights;
		⑤ apply the bacteria on the solid medium with a coated rake and spread it
		evenly, cover the culture dish;
		6 inverted the solid medium, placed in 37 °C incubator, cultured 12h.
7	Picking Colony	① remove the non-antibiotic liquid medium, absorb antibiotics, added chloramphenicol 5 ‰ or ampicillin 1 ‰;
		② tilt the liquid medium to add antibiotics;
		③ open the solid medium, looking for a single colony, then use a tip to row the
		colony, and put the tip into the liquid medium;
		③ place it in the 37 °C shaker, culture 12h.