

**SOP Name:** Taq DNA Polymerase PCR

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**Source(s):** New England BioLab

### Reaction setup:

We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (95°C).

### Materials:

Component	25 $\mu$ l reaction	50 $\mu$ l reaction	Final Concentration
10X Standard <i>Taq</i> Reaction Buffer	2.5 $\mu$ l	5 $\mu$ l	1X
10 mM dNTPs	0.5 $\mu$ l	1 $\mu$ l	200 $\mu$ M
10 $\mu$ M Forward Primer	0.5 $\mu$ l	1 $\mu$ l	0.2 $\mu$ M (0.05–1 $\mu$ M)
10 $\mu$ M Reverse Primer	0.5 $\mu$ l	1 $\mu$ l	0.2 $\mu$ M (0.05–1 $\mu$ M)
Template DNA	variable	variable	<1,000 ng
<i>Taq</i> DNA Polymerase	0.125 $\mu$ l	0.25 $\mu$ l	1.25 units/50 $\mu$ l PCR

Nuclease-free water	to 25 µl	to 50 µl	
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Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary. Overlay the sample with mineral oil if using a PCR machine without a heated lid.

Transfer PCR tubes from ice to a PCR machine with the block preheated to 95°C and begin thermocycling.

### **Procedure:**

#### **Thermocycling conditions for a routine PCR:**

STEP	TEMP	TIME
Initial Denaturation	95°C	30 seconds
30 Cycles	95°C 45-68°C 68°C	15-30 seconds 15-60 seconds 1 minute/kb
Final Extension	68°C	5 minutes
Hold	4-10°C	