



Restriction protocol:

General steps:

Extaction from distribution kit \to Miniprep \to Digestion \to cleaning from gel \to ligation \to Chemical transformation.

Goal- Digest with restriction enzymes both vectors prior to ligation step.

Materials:

| • | Restriction Enzymes: |
|---|----------------------|
| | NEB buffer: |

Molecular biology water

Procedure:

Set up the following reaction a described below:

| Volume | | |
|------------|-----|--|
| 4µl | | |
| Xμl 1μl | | |
| | 1µl | |
| 34-X μΙ | | |
| 40µl | | |
| | | |

Ligation:

Goal- ligate vector and insert for creation of a circular cloned plasmid.

Materials:

T4 DNA ligase

10X T4 DNA ligase buffer

Purified, linearized vector

Purified, linearized insert

Molecular Biology Water

Procedure

- 1. set up following reaction in microcentrifuge tubes.
- 2. Your negative control is the vector with no insert. Prepare a mix with all the ingredients





below but the insert. Divide mix, and add insert to your reaction tube.

| Component | Volume |
|---|------------|
| T4 DNA ligase | 1 |
| 10X T4 DNA ligase buffer | 2 |
| Purified, linearized vector (25-50 ng) | |
| Purified, linearized insert (1:3 ration*) | |
| Molecular Biology Water | up to 17µl |
| Total | 20 |

- 3.Gently mix the reaction by pipetting up and down and spin down briefly.
- 4.Incubate for 1 hr. At 30°C in a dry bath or 16 °C overnight.
- 5.stor samples on ice for subsequent transformation or store at -20 °C

(insert amount [ng])X(plasmid length)=(plasmid amount [ng])X (insert length)

(insert amount [ng])={(plasmid amount [ng])X(insert length)X3}/(plasmid length)

^{*}How to calculate a molar ration of 1:3