

**iGEM TU/e 2017**Biomedical Engineering

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# Digestion



## **Table of contents**

1	Digestion	3
1.1	Materials	3
1.2	Setup & Protocol	3

Digestion

### 1 Digestion

Estimated bench time: 30 minutes Estimated total time: 2 hours

Purpose: Giving sticky ends to the vector and the insert(s) so that they can be ligated.

It is essential to work with gloves at all times to protect the DNA from DNase activity.

#### 1.1 Materials

- 10X Cut-Smart buffer
- Autoclaved H<sub>2</sub>O
- Autoclaved PCR tubes
- Bucket with ice
- Insert(s) which is/are to be digested
- Pipettes and tips
- Restriction enzymes
- Thermal cycler
- Vector which is to be digested

#### 1.2 Setup & Protocol

Construct a PCR mixture in the following way. Use 2 U restriction enzyme per 1 µg DNA. Start with the component with the largest volume and end with the two restriction enzymes. Keep the enzymes on ice.

Component	Quantity/mass/final concentration	Volume (μΙ)
H <sub>2</sub> O	Fill up to 50 µl	
10x Cut-Smart buffer	1 X	5
Plasmid DNA	5 μg	
Restriction enzyme 1	10 U (20 U/μl stock)	0.5
Restriction enzyme 2	10 U (20 U/μl stock)	0.5
Total		50

- Mix well by pipetting up and down.
- Run the following PCR program:

Step	Temp (°C)	Time (min)
Incubation	37	60
Heat inactivation	Variable (65,80,no inactivation possible) <sup>1</sup>	20
Cooling	4	Hold

<sup>&</sup>lt;sup>1</sup> Typical values for NEB restriction enzymes. It is advisable to use to manufacturers protocol for these experiments before setting the deactivation temperature