Yeast Transformation with Lithiumacetat method:

- 1. Inoculate 15ml of YPD medium with S. cerevisiae (INV Sc1.) and shake over night at 30°C with 200 RPM
- 2.dertermine the oD $_{600}$ of the o/n culture and dilute in on oD $_{600}$ of 0,4 in 50ml YPD medium and let it grow for an additional 4h (9.40-13:40)
- 3-pellet at 2500g for 10 min at RT and resuspend in 40ml 1xTE-buffer
- 4.pellet at 2500g for 10 min at RT and resuspend in 2ml 1xLiAc/0,5xTE
- 5.incubate the cells for 10 min
- 6.transformation-mix: 100µl yeast suspension (from step 6)
 - +1-2µg plasmid DNA
 - +50 μl denatured single-stranden carrier DNA
 - (heat up for 60°C)
- 7.add 700µl 1xLiAc/40% PEG-3350/1xTE and mix well (vortex)
- 8.Incubate at 30°C for 30 min (water bath)
- 9.add 88µl 100% DMSO, mix well and heat shock at 42°C for 7 min (water bath)
- 10.centrifuge for 30 sec at 7000rpm and remove supernant and remove supernant
- 11.resuspend the cells with 1ml 1xTE
- 12.centrifuge for 30 sec and remove the supernant
- 13.resuspend the cells in $100\mu l$ 1xTE and plate on a selectable plate and incubate the plates at RT for 4 days