

PCR Using Taq DNA Polymerase

Aim

Amplification of a DNA fragment of interest for further experiments.

Procedure

Note: All reaction components should be assembled on ice and the reaction mix should be quickly transferred to a thermocycler preheated to the denaturation temperature (98°C).

Thaw 5X Phusion HF or GC Buffer, dNTPs, primers, DNA sample, DMSO, and Phusion DNA Polymerase on ice. Let thaw completely and mix before adding to reaction mix to avoid localized concentration differences.

Prepare a primer working solution containing 1 μ l of the forward primer (100 μ M) and 1 μ l of the reverse primer (100 μ M), and 8 μ l of RNase-free water for a final volume of 10 μ l. This achieves a final primer concentration of 200 nM.

Set up the following reaction on ice:

COMPONENT	25 μl REACTION	50 μl REACTION	FINAL CONCENTRATION
10X Standard Taq Reaction Buffer	2.5 µl	5 μΙ	1X
10mM dNTPs	0.5 μΙ	1 μΙ	200 μΜ
Primer mix	0.6 μΙ	1.2 μΙ	200 nM
Template DNA	variable	variable	<250 ng
Taq DNA Polymerase	0.125 μΙ	0.25 μΙ	1

Lab protocol

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RNase-free water	To 25 μl	Το 50 μΙ	0.5 U/ 1.0 U
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Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary.

Quickly transfer PCR tubes to a PCR machine and begin thermocycling.

STEP	TEMP	TIME
Initial Denaturation	95°C	30s
25-35 cycles: Denaturation Annealing Extension	95°C *45-68°C 68°C	15-30s 15-60s 1min/kb
Final Extension	68°C	5min
Hold	4°C	-

^{*}Use of the NEB Tm Calculator is highly recommended.

When the program is finished, the PCR product may be removed and stored at 4°C.

Note:

The above protocol could also be used for colony PCR reactions.

The colony PCR reaction set-up follows the standard PCR protocol described above.

The DNA is obtained by picking a single colony from the plate using a sterile pipette tip and thoroughly swirling it in 20 µl of RNase-free water.

 $1 \,\mu l$ of this water will be used for the PCR reaction.

The rest could be plated or inoculated in a flask overnight.

Sources

This protocol is a modified version of the original <u>Taq DNA Polymerase with Standard Taq Buffer PCR amplification protocol</u> provided by NEB®.

Lab protocol