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## *Coomassie Blue Stain*

2017 Protocols

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*Purpose:*

To visual total protein in a given sample

*Required Materials:*

- dH<sub>2</sub>O
- 1.5M Tris HCl pH 8.8
- 0.5M Tris HCl pH 6.8
- 30% Acrylamide/Bis solution (Bio-Rad)
- 10% SDS
- 10% Ammonium Persulfate in dH<sub>2</sub>O
- TEMED
- 2X SDS Gel Running Buffer
- Gel dock, glass gel plates
- Glacial Acetic Acid
- methanol
- Coomassie Brilliant Blue R-250

*Solutions*

Destain Solution (1L)

- 500mL H<sub>2</sub>O
- 400mL Methanol
- 100mL Glacial Acetic Acid

Coomassie Brilliant Blue Solution

- 1 g of Coomassie
- 500 mL Methanol
- 100 mL Glacial Acetic Acid
- Up to 1L with H<sub>2</sub>O

Stir for 3-4 hours, then filter through Whatman filter paper

*Procedure:*

1. Repeat steps 1-8 of the Western Blot protocol.
2. Once gels have finished running, cover gel with Coomassie Brilliant Blue, cover with lid, and incubate for 30 min
3. Rinse with water twice
4. Add enough destain solution to cover the gel and leave overnight, gently shaking at room temperature