

Polymerase Chain reaction (PCR)-Clean up

Aim of the experiment

This protocol can be used for the quick purification of an exponentially PCR amplified DNA of interest. The protocol described is based on :

Monarch PCR-Clean up Kit

Materials

- PCR amplified DNA of interest
- NEB-Monarch PCR-clean up kit
- nuclease-free H₂O (nf H₂O, Sigma Aldrich, Germany)
- Centrifuge (1.5 ml Tubes, 16.000 rcf)

Procedure

1. Add to a column, collection tube combination the following dilution of your sample:

Table 1: PCR-clean up initial set up

Volumes	Chemicals
1	DNA of interest
2-5	DNA binding buffer

2. Spin for 1 min at 16.000 rcf and room temperature
 3. Trash flow-through
 4. Add 200 µl of DNA wash buffer to column
 5. Spin for 1 min at 16.000 rcf and room temperature
 6. Trash flow-through
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7. Repeat Step 4-6
8. Make sure no ethanol touches the column when discarding the ethanol flow-through for the second time
9. Transfer the column into a fresh 1.5 ml tube
10. Add 6-20 μ l of either Elution buffer or nuclease-free water to the column
11. Let the elutant incubate for at least 1 min
12. Elute by spinning for 1 min at 16.000 rcf and room temperature

Possible follow up protocols

The following protocols are the next steps of a possible cloning cycle after a PCR-clean up:

1. Restriction digest
 2. Agarose-Gel-electrophoresis
 3. Gibson-Assembly
 4. Golden-Gate Assembly
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