



## **Colony PCR**

- 1. Keep everything on ice. The PCR reaction mixture in the ice preparation, and then placed in a PCR reaction in the PCR reaction apparatus. This cold start (Cool Start Method) can enhance the specificity of PCR amplification to reduce non-specific reactions in the PCR process, can get good PCR results.
- 2. Make up a master mix of everything into one microcentrifuge tube.
- 3. Pipette up and down in the microcentrifuge tube, drain  $25\mu L$  or  $50.0\mu L$  solution to each PCR tube.
  - a. 25.0 µL reaction system

18.375μL ddH2O 5μL 5X OneTaq® Standard Reaction Buffer 0.5μL dNTPs 0.5μL forward primer

 $0.5\mu L$  reverse primer

**0.125μL** OneTaq DNA polymerase

Colony stab (template DNA)

-----25.0 $\mu$ L Total

## b. 50.0µL reaction system

**36.75μL** ddH20

**10μL** 5X OneTag® Standard Reaction Buffer

1.0µL dNTPs

1.0µL forward primer

1.0µL reverse primer

**0.25μL** DNA polymerase

Colony stab (template DNA)

-----50.0uL Total

- 4. Pick colonies from plates, dilute in 10  $\mu L$  ddH2O and add 1  $\mu L$  into PCR tubes.
- 5. Run the "Colony PCR" program, and adjust your extention time as described below.

## The "Colony PCR" program

Initial denaturation: 95°C for 5:00min

30 cycles of:

94°C for 0:30 min

55°C for 0:30 min (different primers different annealing temperature)

72°C for t min ("t" depends on the length of goal sequence, 1min per 1000bp)







Final extension: 72°C for 10:00 min