File name: iGEM2016_01_iTRAQ_analysis.docx

iGEM2016 - Microbiology - BMB - SDU

Project type: Plastic

Project title: characterizing and optimizing PHB

productin in E. coli.

Sub project:

1.

Creation date: 2016.09.26

Written by: Joel

Performed by: Joel Mario Vej-Nielsen & Jakob

Rønning

1. SOPs in use.

SOP0001_v1 - ON culure of *E. coli*

SOP0037_v1 - iTRAQ sample preparation

SOP0038_v1 - Qubit® Protein Assay Kits

SOP0039_v1 - C2 and C3 columns

SOP0040_v1 - iTRAQ labelling

SOP0041_v1 - TiO_2 purification

2. Purpose.

Proteomics analysis of Staphylococcal pantothenate kinase II in E. coli.

3. Overview.

Day	SOPs	Experiments
1	SOP0001_v1	ON cultures of E. coli.
2	SOP0037_v1 SOP0038_v1	iTRAQ sample preparation Qubit® Protein Assay Kits
3	SOP0039_v1 SOP0038_v1 SOP0040_v1	C2 and C3 columns Qubit® Protein Assay Kits iTRAQ labelling
4	SOP0041_v1 SOP0039_v1	${\rm Ti}O_2$ purification C2 and C3 columns
5	SOP0041_v1 SOP0039_v1	${\rm Ti} {\cal O}_2$ purification C2 and C3 columns

 $File\ name: iGEM2016_01_iTRAQ_analysis.docx$

 $File\ name: iGEM2016_01_iTRAQ_analysis.docx$

4. Materials required.

Materials in use

Name	Components (Concentrations)	Manufacture r / Cat. #	Room	Safety considerations
PBS				
Lysis buffer	93,8 μl Urea (6M) thiourea (2M) solution 1 μl 1M DTT (10mM) 3 μl phosSTOP phosphatase inhibitor 1,2 μl complete protease inhibitor	Sigma Aldrich	Protein research group Stored in freezer Stored in freezer	Handle in fume closet
	2 μl benzonase		Stored in freezer	
Triethyla-mmonium bicarbonate (TEAB)	20mM, pH 7,5		Protein research group	
Iodoacetamide			Protein research group	
Trypsin			Stored in freezer	
Qubit® Reagant		ThermoFisch er Scientific	Protein research group	
Qubit® Buffer		ThermoFisch er Scientific	Protein research group	
Qubit® Standards		ThermoFisch er Scientific	Protein research group	
Qubit® Assay tubes		ThermoFisch er Scientific	Protein research group	
0,5 ml PCR tubes		ThermoFisch er Scientific	Protein research group	
iTRAQ Reagant 114		Sciex	Stored in freezer	
iTRAQ Reagant 115		Sciex	Stored in freezer	
iTRAQ Reagant 116		Sciex	Stored in freezer	
iTRAQ Reagant 117		Sciex	Stored in freezer	
ethanol		Sciex	Stored in freezer	
Low binding			Protein research	
eppendorf tubes	000/ 1 1 1 1 70/ 771 0 11		group	
Loading Buffer	80% Acetonitril, 5% TFA & 1M glycolytic acid (76 mg/ml)		Protein research group	

File name: iGEM2016_01_iTRAQ_analysis.docx

Washing Buffer 1	80% Acetonitril & 1% TFA	Protein research
		group
Washing Buffer 2	10% Acetonitril & 0,1% TFA	Protein research
		group
Elution Buffer	60μl ammonia solution in 940μl	Protein research
	H_2O (pH 11,3)	group
TFA	0,1%	Protein research
		group
TFA	10%	Protein research
		group
TFA	100%	Protein research
		group
Acetonitril	30%	Protein research
		group
HCl	12M	Protein research
		group
HCl	1M	Protein research
		group
PNGase F		Protein research
		group
Sialidase A		Protein research
		group
Formic acid	100%	Protein research
		group
Low binding		Protein research
eppendorftubes		group
TiO ₂ beads		Protein research
		group

5. Other

Primer to PCR dilution:

6. Experiment history.

Date	SOPs	Alterations to SOPs and remarks to experiments
(YY.MM.DD)		

File name: iGEM2016_01_iTRAQ_analysis.docx

ON culture bacteria expres		50 ml of ON culture was prepared for two biological replicates of bacteria expressing pantothenate kinase II and two biological
		replicates of <i>E. coli</i> with pSB1C3 alone.
16.10.05	SOP0037_v1	
	iTRAQ sample	
46.40.05	preparation	
16.10.05	SOP0038_v1 Qubit® Protein	Concentrations were:
	Assay Kits	Sample 1 (panK) – 4.38 mg/ml
	Assay Kits	Sample 2 (panK) – 4.24 mg/ml
		Sample 2 (control) – 5.2 mg/ml
		Sample 4 (control) – 5.48 mg/ml
16.10.06	SOP0039_v1 C2 and C3 columns	Samples were run through a C2 column and then a C3 column
16.10.06	SOP0038_v1	Concentrations were:
	Qubit® Protein	2.5 mg/ml
	Assay Kits	2.5 mg/ml
		2.0 mg/ml 2.8 mg/ml
		2.0 (((g/))))
		The samples were analyzed by MALDI – MS In order to determine why concentrations were so low.
16.10.06	SOP0040_v1 iTRAQ labelling	As there was no 4plex available, we used 8plex chemicals instead. However, they are of a different compositions and are should be
	_	suspended in isopropanal not ethanal. We didn't know this and
	-	suspended in isopropanol not ethanol. We didn't know this and
		this has effected the labelling.
16.10.07	SOP0041_v1 TiO_2 purification	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 μl of
16.10.07 16.10.07	-	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 μ l of each sample were kept separately.
	${\rm Ti} {\it O}_2$ purification	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides.
16.10.07	TiO ₂ purification SOP0039_v1 C2 and C3 columns	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides. Afterwards LC-MS/MS were performed on the samples in order to retrieve data on the bacterial proteome.
	TiO ₂ purification SOP0039_v1 C2 and C3	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides. Afterwards LC-MS/MS were performed on the samples in order to
16.10.07	TiO ₂ purification SOP0039_v1 C2 and C3 columns SOP0039_v1 C2 and C3	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides. Afterwards LC-MS/MS were performed on the samples in order to retrieve data on the bacterial proteome. The purification was repeated, due to visible impurities in the dried
16.10.07 16.10.07 16.10.08	TiO_2 purification SOP0039_v1 C2 and C3 columns SOP0039_v1 C2 and C3 columns SOP0041_v1 TiO_2 purification	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides. Afterwards LC-MS/MS were performed on the samples in order to retrieve data on the bacterial proteome. The purification was repeated, due to visible impurities in the dried down sample. The purification was performed on each of the remaining 5 µl that had not been pooled.
16.10.07 16.10.07	TiO ₂ purification SOP0039_v1 C2 and C3 columns SOP0039_v1 C2 and C3 columns SOP0041_v1	this has effected the labelling. Therefore, instead of pooling the entirety of each sample, 5 µl of each sample were kept separately. The purification was performed on the pooled peptides. Afterwards LC-MS/MS were performed on the samples in order to retrieve data on the bacterial proteome. The purification was repeated, due to visible impurities in the dried down sample. The purification was performed on each of the remaining 5 µl that

 $File\ name: iGEM2016_01_iTRAQ_analysis.docx$

7. Sample specification.

Sample name	Sample content	From	Used for / Saved where
#102-1	Top10 containing K2018021	Assembled from genscript genes and iGEM	Sample tested containing pantothenate kinase II
#102-2	Top10 containing K2018021	Assembled from genscript genes and iGEM	Sample tested containing pantothenate kinase II
#56-1	Top10 containing K608004	From iGEM kit	Control sample
#56-2	Top10 containing K608004	From iGEM kit	Control sample

8. Remarks on setup.

9. Results and conclusions.

10. Appendixes