# JMJ-Group - Microbiology - BMB - SDU

Title: Sorting setup FACS Aria II Date issued: 2013.01.17

SOP number: SOP0043\_v01 Review date: 2013.12.01

Version number: 01 Written by: Tina Kronborg

### 1. Purpose

To set the FACS up for sorting

### 2. Area of application

This procedure is valid for the FACS Aria II placed in class II laboratory V16-501f-2

## 3. Apparatus and equipment

Apparatus/equipment	Location (Room number)	Check points	Criteria for approval/rejection
FACS	Laboratory (class 2) – V16-501f-2	<ul> <li>Size of the neutral density filter</li> <li>Nozzle size</li> <li>Waste bin empty</li> <li>Sheath fluid above minimum</li> </ul>	Both tubes are connected to the sheath fluid tank
Refrigerator	Laboratory (class 2) – V16-501b-2	•	

## 4. Materials and reagents – their shelf life and risk labelling

Name	Components	Supplier / Cat. #	Room	Safety considerations
Accudrop beads	This product contains a single population of 6-µm particles.  Every particle contains a fluorophore that is excited at 670 nm and emits at 750 nm.  The particles are supplied in 1.5 mL of water containing 0.05% Tween® 20 and 2 mM sodium azide.  See paragraph 12. Appendix	BD / 345249	Refrigerator V16- 501b-2	
PBS	1X Phosphate Buffered Saline (PBS Buffer):  1 L distilled H <sub>2</sub> O  8 g NaCl  0.2 g KCl  1.44 g Na <sub>2</sub> HPO <sub>4</sub>	Sigma-Aldrich / 31434N J. T. Baker / 3040-01 MERCK / 1.06580.1000 MERCK / 1.04873.1000	V18-405-0 V18-405-0 V18-405-0 V18-405-0	
FACS Flow sheet fluid	See paragraph 12. Appendix	BD / 342003	Refrigerator V16- 501b-2	

### 5. QC - Quality Control

Check expiry date on the beads

Make sure the beads are dissolved before use

When sorting large or fragile cells, use a larger nozzle size and lower pressure:

Nozzle 85 microns and pressure on 45 psi or

Nozzle 100 microns and pressure on 20 psi

When sorting smaller or less fragile cells, use a smaller nozzle size and higher pressure:

Nozzle 70 microns and pressure on 70 psi

Flow rate (events/sec) according to nozzle side for autodrop delay:

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70 micron = 1.000 – 3.000

85 micron = 800 – 2.000

100 micron = 600 – 1.500

130 micron = 400 – 1.200
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#### Sort precision modes:

Purity mode (Yield Mask 32, Purity Mask 32, Phase Mask 0):

Gives a very pure sample, but a low yield and lots of cells which are discarded because of conflicts.

**4-Way Purity** (Yield Mask 0, Purity Mask 32, Phase Mask 0):

Recommended for four-way sorting where precise deflection is required

**Yield** (Yield Mask 32, Purity Mask 0, Phase Mask 0):

Low purity - could be used as a first round sort for enrichment of target particles, followed by a sort for purity.

Single cell mode (Yield Mask 0, Purity Mask 32, Phase Mask 16):

Used for plate sorting or situations where precise counting is required.

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Initial (Yield Mask 32, Purity Mask 0, Phase Mask 0):
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As Yield (used during sort Setup, automatic)

**Fine Tune mode** (Yield Mask 0, Purity Mask 0, Phase Mask 0):

Give the maximum number of drops. Are used to fine-tune the drop delay value.

#### 6. List of other SOPs relevant to this SOP

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JMJ SOPFACS0001 v01 TK Daily use of FACS Aria II
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### 7. Environmental conditions required

### 8. Procedure

8.1	Start up the FACS as described in JMJ_SOPFACS0001_v01_TK_Daily_use_of_FACS_Aria_II
8.2	Make sure that the steam is stable and set according to nozzle size
8.3	Turn on sweat spot
8.4	Mix 1 ml PBS with 4 drops of accudrop beads in a FACS tube (remember to vigorously shake the beads before mixing with PBS). Can be used until empty
8.5	Open a new experiment – accudrop delay
8.6	Mark the tube and open the sort layout
8.7	Load the accudrop beads and adjust the steam according to nozzle size, see in paragraph 5 The flow rate settings shall be between 1 and 5 else adjust the beads concentration
8.8	Turn on the voltage in the side stream window
8.9	Click sort in the sort layout window
8.10	Click chancel in the confirm dialog according the waste drawer
8.11	Adjust the micrometer dial to obtain the brightest bead spot on the center stream
8.12	Click the auto delay button in the side stream window
8.13	The two boxes representing the optical filters, indicate the region of the image where the left and center stream intensities are calculated during image processing
8.14	If the left side stream is not completely contained in the left region, adjust the voltage slider to place the stream in the center of the region
8.15	When the auto drop delay dialog is open click start run
8.16	Don't turn off sweat spot after the run is finished, the sweat spot must be on during the whole sort
8.17	If the steam are turned off during the sort, the drop delay must be re-runned
8.18	The drop delay is set manually by the arrows at the drop delay in top of the side stream window.
8.19	Click the optical filter button in the side stream window, so you can see the accudrop beads in front of the lower camera
8.20	It take few seconds to register the change so be patient ☺
8.21	Unload the accudrop beads
8.22	Open the experiment in question
8.23	Adjust the gates to define the population(s) of interest
8.24	Mangler indstilling af stream ned I rør

## 9. Waste handling

Chemical name	Concentration	Type of waste (C, Z)	Remarks
ON Culture		Liquid bacterial waste	
Once use plastic		GMO yellow waste	

## 10. Time consumption

• Start up: 10 min

• CST:

• Drop delay:

• Baseline:

Clean after run:Shut down: 8 min

# 11. Scheme of development

Date / Initials	Version No.	Description of changes
13.01.16 / TK	01	The SOP has been written
13.01.02 / JR	01	The SOP has been approved

### 12. Appensdixes

Data sheet for CST Beads: 23-9141-01\_CS&T Beads\_PI\_RUO is found in:

S:\999 Public\!SOPs and Protecols\FACS\FACS Aria II\SOPs for daily runs\Data sheets

Data sheet for Accudrop Beads: 23-8292-01\_AccudropBeads is found in:

S:\999 Public\!SOPs and Protecols\FACS\FACS Aria II\SOPs for daily runs\Data sheets

Data sheet for: FACS Flow sheet fluid: 342003-MSDS-EUEN-01 FACS Flow is found in:

S:\999 Public\!SOPs and Protecols\FACS\FACS Aria II\SOPs for daily runs\Data sheets

Data sheet for: FACS Clean: 340345-MSDS-EUEN-00 FACS Clean is found in:

S:\999 Public\!SOPs and Protecols\FACS\FACS Aria II\SOPs for daily runs\Data sheets

Data sheet for: FACS Rinse: 340346-MSDS-EUEN-00 FACS Rinse is found in:

S:\999 Public\!SOPs and Protecols\FACS\FACS Aria II\SOPs for daily runs\Data sheets

PBS:

10 x concentration: 5 L

- 1. Dissolve the following in 3.5 L distilled H<sub>2</sub>O.
  - 400 g of NaCl
  - 10 g of KCl
  - 72 g of Na<sub>2</sub>HPO<sub>4</sub>
  - 12 g of KH<sub>2</sub>PO<sub>4</sub>
- 2. Adjust pH to 7.4.
- 3. Adjust volume to 5 L with additional distilled H<sub>2</sub>O.
- 4. Sterilize by autoclaving