- Amplification of 110 matrix with:

1 and 10 primers 11 and 12 primers 13 and 3 primers 20 and 6 primers + Control

Hybridation temperature used : 69°C Elongation time used : 1 min 30 sec

- Gel verification of amplifications Agarose gel: 1% 100 mV, 30 min

=> Successful amplification of all per products

- Amplification of 113 matrix with:
7 and 14 primers
15 and 16 primers
17 and 8 primers
+ Control

Hybridation temperature used : 69°C Elongation time used : 20 sec

- Gel verification of amplifications Agarose gel : 1% 100 mV, 30 min

=> Successful amplification of :

7 + 14

8 + 17

- Amplification of Dnmt matrix with:

23 and 9 primers 21 and 4 primers 5 and 2 primers 22 and 4 primers

+ Control

Hybridation temperature used : 68°C Elongation time used : 50 sec

- Gel verification of amplifications Agarose gel: 1% 100 mV, 30 min

=> Successful amplification of all per products

# GIBSON ASSEMBLY (dCas9)

 $\begin{array}{l} psB1C3:0,5~\mu L \\ 1+10:1~\mu L \\ 11+12:1\mu L \\ 13+3:1\mu L \\ Mix:10\mu L \\ H_2O:6,5\mu L \end{array}$ 

(Control done with circular plasmid)

- Put on the thermocycler at 50°C during 60 min
- Put at -20°C until the transformation step

- Transformation in DH5 $\alpha$  of : psB1C3 containing 3 fragments of 110 matrix (LB + Choramphenicol)
- Put plates at 37°C

- Amplification of 2 matrix with: 15 and 16 primers (113 matrix) 18 and 19 primers (gDNA matrix) + Control

Hybridation temperature used : 69°C Elongation time used : 15sec

- Gel verification of amplifications

Agarose gel: 1% 100 mV, 30 min

=> No amplification observed

- No colony on the plate containing the cloning of 110 matrix (08-15-16)

## **CLONING**

- Digest of
psB1C3 with EcoRI and PstI
10X GCN4 with EcoRI and PstI
Dnmt3a with EcoRI and PstI

- Put on the incubator at 37°C during 1h
- Dephosphorylation of psB1C3 Put on the incubator at 37°C during 1h
- Phosphatase inactivation At 80°C during 10 min
- Ligation

10X GCN4 in psB1C3 Dnmt3a in psB1C3 Put 1h at room temperature

- Transformation in DH5α of: psB1C3 containing 10X GCN4 (LB + Choramphenicol) psB1C3 containing Dnmt3a
- Put on the incubator at 37°C

- Amplification of 2 matrix with: 15 and 16 primers (113 matrix) 18 and 19 primers (gDNA matrix)

+ Control

Hybridation temperature used : 74°C

Elongation time used: 15sec

- Gel verification of amplifications

Agarose gel: 1% 100 mV, 30 min

=> No amplification observed

#### **GIBSON ASSEMBLY**

psB1C3 with:

1 + 10 fragment 11 + 12 fragment 13 + 3 fragment Control

- Put on the thermocycler at 50°C during 60 min

- Transformation in DH5α of : psB1C3 containing 3 fragments of 110 matrix (LB + Choramphenicol)
- Put plates at 37°C

# **NEW TRY OF TRANSFORMATION**

- Transformation in DH5α of: psB1C3 containing 10X GCN4 (LB + Choramphenicol) psB1C3 containing Dnmt3a
- Put on the incubator at 37°C
- No colony on plates containing 10X GCN4 cloning and Dnmt3a cloning (08-17-16)

- No colony on the plate containing the cloning of 110 matrix (08-18-16)

#### **SET UP MINICULTURES**

- Pick selected colony of agar plates and put it on 6mL of LB + Chloramphenicol psB1C3 containing 10X GNC4 (X2) psB1C3 containing Dnmat3a (X2)
- Put on the shaking incubator at 37°C

## GIBSON ASSEMBLY (Dnmt3a31)

```
psB1C3 with:
21 + 4 fragment
5 + 2 fragment
Control
```

- Put on the thermocycler at 50°C during 60 min

# GIBSON ASSEMBLY (Dnmt31)

```
psB1C3 with:
22 + 4 fragment
5 + 2 fragment
Control
```

- Put on the thermocycler at 50°C during 60 min

- Transformation in DH5 $\alpha$  of : psB1C3 containing Dnmt3a3l (LB + Choramphenicol) psB1C3 containing Dnmt3l
- Put plates at 37°C

## **MINIPREP**

- Miniprep of:

psB1C3 10X GCN4 1 psB1C3 10X GCN4 2 psB1C3 Dnmt3a 1 psB1C3 Dnmt3a 2

# **GEL MIGRATION**

- Gel verification of miniprep tubes
Fisrt, the plasmid is digested 1h at 37°C
Agarose gel: 1%
100 mV, 30 min

=> Nothing

- No colony on the plate containing the cloning of Dnmt3a3l (08-22-16)
  No colony on the plate containing the cloning of Dnmt3l (08-22-16)