## **Bacillus subtilis transformation**

## **Materials and Equipment:**

- LB plate with a desired strain
- · gDNA or plasmid
- X10 MC (stored at -20°C)
- 1M MgSo<sub>4</sub> (solution E from resuspension salts)
- sDDW
- Sterile tube (15ml)
- Sterile wooden stick
- Selective marker plates
- Sterile beads
- Freezing tubes (nunc)
- 1ml Liquid LB
- 50% glycerol stock

## **Preparation of materials:**

- 50% glycerol stock
  Add 25ml 100% glycerol to 25ml SDDW and autoclave.
- X10MC

K <sub>2</sub> HPO <sub>4</sub> x3H <sub>2</sub> O	14.036gr
KH <sub>2</sub> PO <sub>4</sub>	5.239gr
Glucose	20gr
Trisoduim citrate	10ml 300mM
Ferric ammonium citrate (X1000)	1ml 22mg/ml
Casein Hydrolysate	1gr
Potassium Glutamate	2gr

<sup>\*</sup> Mix with sDDW for final volume of 100ml

## **Procedure:**

- 1. Pick a single colony form an LB plate with a sterile wooden stick to a sterile tube containing the following:
  - \* 900µl sDDW
  - \* 100µl X10MC
  - \* 10µl 1M MgSo<sub>4</sub>
- 2. Mix by vortex

<sup>\*</sup> Filter

<sup>\*</sup> Dispense to 1ml aliquots

<sup>\*</sup> Store at -20°C

- 3. Incubate in a roller-drum shaker at 37°C for 3.5h
- 4. Dispense sample to sterile tubes each containing 300µl
- 5. Add to the tube 3µl gDNA or 5µl plasmid (1ug DNA)
- \* One tube should contain no DNA for negative control
- 6. Incubate in a roller-drum shaker at 37°C for another 3h
- 7. Plate sample on two different selective marker plates with sterile beads on:
- A. 250µl
- B. 50µl
- 8. Incubate plate at 37°C over night
- 9. Perform differential plating form a single colony with sterile wooden stick to the same selective plate as in step 7.
- 10. Incubate plate at 37°C over night
- 11. If required perform differential plating from a single colony on <u>another</u> selective marker plate.
- 12. Incubate plate at 37°C over night
- 13. Pick a single colony with a wooden sterile stick into 1ml Liquid LB
- 14. Incubate at 23°C over night or at 37°C for 3h in a roller-drum shaker
- 15. Add 900µl sample to 600µl 50% glycerol in a freezing tube (nunc)
- 16. Vortex well
- 17. Freeze the strain at -80°C.
- 18. Add your strain's genotype and phenotype to the filemaker.

**GOOD LUCK!!** 

(This protocol was kindly provided by the lab of Dr. Avigdor Eldar, Tel Aviv University)