

PCR Protocol

Phusion High-Fidelity DNA Polymerase

Reaction setup:

Water	13,3 - X μ l	32,5 - X μ l
10X DreamTaq Buffer	4 μ l	10 μ l
dNTP Mix (10mM each)	0,5 μ l	1 μ l
Forward primer (10 μM)	1 μ l	3 μ l
Reverse primer (10 μM)	1 μ l	3 μ l
DreamTaq (5 U/μl)	0,2 μ l	0,5 μ l
Tempalte DNA	X μ l (0,004 - 100 ng)	X μ l (0,01 - 250 ng)
Total	20 μl	50 μl

Phusion DNA Polymerase exhibits 3' \rightarrow 5' exonuclease activity that can degrade primers in the absence of dNTPs.

Thermocycler program:

Step	Temperature	Time	Cycles
Initial denaturation	98 °C	30 s - 3 min	1
Denaturation	98 °C	5-10 s	
Annealing	Lower T _m + 3 °C	10-30 s	25-40
Extension	72 °C	15-40 s/kb	
Final extension	72 °C	5-10 min	1

DreamTaq DNA Polymerase

Reaction setup:

Water	15,4 - X μ l	37,75 - X μ l
10X DreamTaq Buffer	2 μ l	5 μ l
dNTP Mix (10mM each)	0,5 μ l	1 μ l
Forward primer (10 μM)	1 μ l	3 μ l
Reverse primer (10 μM)	1 μ l	3 μ l
DreamTaq (5 U/μl)	0,1 μ l	0,25 μ l
Tempalte DNA	X μ l (0,004 - 400 ng)	X μ l (0,01 - 1000 ng)
Total	20 μl	50 μl

Thermocycler program:

Step	Temperature	Time	Cycles
Initial denaturation	95 °C	1-3 min	1
Denaturation	95 °C	30 s	
Annealing	Lower - 5 °C	30 s	25-40
Extension	72 °C	1 min/kb	
Final extension	72 °C	5-15 min	1

PrimeSTAR HS DNA Polymerase

Reaction setup:

Water	13,3 - X μ l	32,5 - X μ l
10X DreamTaq Buffer	4 μ l	10 μ l
dNTP Mix (10mM each)	0,5 μ l	1 μ l
Forward primer (10 μM)	1 μ l	3 μ l
Reverse primer (10 μM)	1 μ l	3 μ l
DreamTaq (5 U/μl)	0,2 μ l	0,5 μ l
Tempalte DNA	X μ l (0,004 - 100 ng)	X μ l (0,01 - 200 ng)
Total	20 μl	50 μl

Thermocycler program:

Step	Temperature	Time	Cycles
Initial denaturation	98 °C	30 s - 3 min	1
Denaturation	98 °C	10 s	
Annealing	T _m	5-15 s	25-40
Extension	72 °C	1 min/kb	
Final extension	72 °C	5-10 min	1