

Growing Medium of *E.coli* :

Overview :

We used LB broth (Luria broth, Luria-Bertani medium) for plasmid DNA production (cloning, transformation and miniprep). Otherwise, we used only M63 media for curdian production because it is a minimal medium, so we could control all the media parameters for production.

Media :

- Luria-Bertani (LB) broth: 1L

| Component | Volume & Mass | Procedure |
|---------------|---------------|---|
| Bactotryptone | 10 g | 1) Adjust pH to 7.5 with NaOH 2) Adjust volume to 1 L 3) Sterilize by autoclave |
| Yeast Extract | 5 g | |
| NaCl | 10 g | |

- For LB plates : add 12 g/L of agar

- Preparation of 5X M63 Medium

| Component | Volume & Mass | Procedure |
|---|---------------|---|
| (NH ₄) ₂ SO ₄ | 10 g | 1) Add the following reagents to a 2-liter flask 2) Adjust volume to 1 L 3) Once the ingredients are added, heat with stirring until the components are completely dissolved. |
| KH ₂ PO ₄ | 68 g | |
| FeSO ₄ ·7H ₂ O | 2,5 mg | |
| | | 4) Adjust pH to 7.0 with Acid 5) Sterilize by autoclave |

- Preparation of 1X M63 Medium Working Solution

Aseptically dilute 200mL of 5X stock solution with 789 mL of sterile distilled water.

Aseptically add the following sterile solutions:

- 1 mL of 1 M MgSO₄·7H₂O (*directly in the 1X medium, not in the 5X*)
- 10 mL of 20% carbon source (*final concentration: 0,2 %*)
- 0.1 mL of 0.5% vitamin B1 (thiamine)
« Vitamins should be added to a final concentration of 1µg/mL or 1mg/L »
- Antibiotic

Optional :

Add 5 mL of 20% Casamino Acids or L amino acids to 40 µg/mL or DL amino acids to 80 µg/mL

- Preparation of Stock Carbohydrate Solution (Glucose) :

Add 20 g of carbohydrate to distilled water and bring volume to 100mL.

Mix Thoroughly.

Filter sterilize.

- Preparation of Stock MgSO₄·7H₂O Solution :

Add 24,65 g of MgSO₄·7H₂O to distilled water and bring volume to 100mL.

Mix Thoroughly.

Filter sterilize.

Antibiotic :

| Antibiotic | Stock concentration | Working concentration | Dissolve in |
|-----------------|--------------------------|--------------------------|------------------|
| Ampicillin | 50 mg.mL ⁻¹ | 50 µg.mL ⁻¹ | H ₂ O |
| Kanamycin | 50 mg.mL ⁻¹ | 50 µg.mL ⁻¹ | H ₂ O |
| Chloramphenicol | 34 mg.mL ⁻¹ | 10 µg.mL ⁻¹ | 95% Ethanol |
| Tetracycline | 12,5 mg.mL ⁻¹ | 12,5 µg.mL ⁻¹ | 50% Ethanol |

- Cool down the medium to 50°C before adding antibiotics.

Curdlan Production with *E.coli* :

1. Pre-warm the medium at 37°C to decrease the time of lag-phase
2. Take colonies and inoculate 50 mL of complete M63 (1X) + Antibiotic at 37°C overnight
3. Grow cells until A600 : 0.7-0.9
4. Inoculate 30 mL inoculum for 120 mL of the M63 complete medium + antibiotic in a 500 mL Erlenmeyer flask (in order to have A600: 0.2)
5. Put the flask at 37°C, 180 rpm
6. Take out 10 mL of culture, centrifuge 5min at 14,000 rpm and 4°C, discard supernatant and store pellet at -20°C (this is the uninduced time point).
7. During stationary phase, re-incubate remaining cultures at 25°C shaking with 180rpm during 21h.