Week 9

DAY 36:

- Troubleshooting for problems and future work to be done

DAY 37:

- Script for crowdfunding video edited.
- Searching for Experiment data and came to know “they only take US researchers ideas’ but we dropped mail for our request.

Week 10

29 June (Amplification of Kana, Amp and SOx):

- Sox and Kana were added in columns
- Plasmid isolation done for Amp, Kana and SOx.

30 June:

- Digestion of Amp, Kana and SOx

Sox Digestion:
Plasmid 20ul
Buffer 5ul
Ecor1 0.5ul
Pst1 0.5ul
Water 24ul
Total 50ul

Amp Digestion:
Plasmid 60ul
Buffer 20ul
Ecor1 2ul
Pst1 2ul
Water 116ul
Total 200ul

Kana Digestion:
Plasmid 60ul
Ecor1 1ul
Pst1 1ul
Water 73 ul
Total 150 ul

Gel electrophoresis performed with 1% agarose

COMPOSITION

- Sample

Name	**Sample**	**Dye(1000 base pairs)**
S0x	10ul	2ul
amp	5ul	1ul
kana	5ul	1ul

- Length of the **Promoter + RBS** : *35bp+60*(suffix)
  Length of various sequences determined:
  Amp:::
  Plasmid pSB1A3: 2155bp
  Promoter+RBS : 35bp Kana:::
  Plasmid pSB1K3 : 2204bp
  Biobrick
  SQR::: 3351 bp
  Plasmid pSB1C3 : 2070bp
  SQR gene : 1281bp
  NrfA : 3507bp
  Plasmid pSB1c3 :2070 bp
  Biobrick (NrfA) :1437bp
  #Note: Lengths of Suffix and Prefix have been excluded in the cases.
  Length of prefix and suffix in all cases=60 bp

1 July:

- One sample each of Promoter, Nrf gene and Sox clone were inoculated
- Transformation of NOx and SOx carried out

2 July:

- Digestion of SQR biobrick by Ecor1 and pst1
- Verification by gel run electrophoresis

Growth was found in both the plates of S0x and N0x transformed the previous day.
Only one band was observed instead of 2 expected bands, thereby suggesting digestion has not occurred.

3 July:

- Inoculated and streaked for NOx and Sox on Tet and Kana antibiotic resistances.
- S0x tube was found contaminated, so REINNOCULATION of "S0x SQR 3/7/14 Kana" tube.
- NOx sample was pelleted in 1.5 Micro Centrifuge Tube (MCT)