Gel Extraction

1. Get an Eppendorf tube, and tare its weight
2. Image Gel
3. Cut the gel and put it in the tube.
4. Weigh the gel and record its mass
   a. only load max 400mg per column
5. Add 3 times the mass of the gel of qG buffer (400mg=1200 uL)
6. Incubate at 50C for 10mins - While waiting take out 2 or 3 times to vortex
7. Add 1 times the mass of the gel amount of isopropanol to gel (300mg=300uL)
8. Load 750 uL of that mixture into the gel extraction column (purple column)
9. Spin down for 1 min at 13,000 rpm

. Keep doing this(using the same column) until you harvest all the DNA in your sample

10. wash with alcohol PE buffer to keep plasmid stuck on filter

. check the cap sticker to make sure ethanol is added

a. add 750 uL PE buffer to column filter
b. centrifuge 13,000 RPM for 1 min, no liquid should be left on filter
c. pour off Ethanol flow-through in collection tube into liquid waste

11. evaporate alcohol by spin

. centrifuge 13,000 RPM for 1 min

12. add Elution Buffer (EB: H20+ salts) to filter to elute DNA

a. move filter over brand new Eppendorf 1.5mL tube

b. see that it goes on to the filter, but do not puncture the filter
c. add 35 uL EB to filter
d. Let SIT ON BENCH on filter for 1 min
e. centrifuge 13,000 RPM for 1 min