

Presto Mini Plasmid Kit Protocol Procedure (for iGEM use)

preparation	<ul style="list-style-type: none"> • Warm MBW to 60 °C in dry Bath (*for sequencing use UPW) • Bring PD1 buffer from 4 °C on ice • Make Sure nanodrop is available • Bring 16 hr incubated 50 ml tube (starter) from shaker
Step 1: Harvesting	<ul style="list-style-type: none"> • Centrifuge The starter for 10 min at 4000 rpm • Discharge the supernatant carefully, do not lose the pellet
Step 2: Resuspension	<ul style="list-style-type: none"> • Brake the pellet by vortex • Add 200 µl of PD1 Buffer (make sure RNase A was added) and resuspend the pellet by vortex or pipette • Transfer the supernatant to 1.5 ml microcentrifuge tube
Step 3: Lysis	<ul style="list-style-type: none"> • Add 200 µl of PD2 Buffer then mix gently by inverting the tube 10 times. Do not vortex! to avoid shearing the gDNA. • Let stand at room temperature for 2 min to ensure the lysate is homologous
Step 4: Neutralization	<ul style="list-style-type: none"> • Add 300 µl of PD3 Buffer then immediately invert the tube 10 times. Do not vortex! to avoid shearing the gDNA. • Centrifuge for 5 min at 16000g
Step 5: DNA binding	<ul style="list-style-type: none"> • Place a PD column in a 2 ml collection tube then transfer the supernatant to the PD column • Centrifuge for 1 min at 16000g • Discard the flow-through and dry on wetman • Place the PD column back in a 2 ml collection tube
Step 6: Wash	<ul style="list-style-type: none"> • Add 400 µl of W1 Buffer in to the PD column. Centrifuge for 1 min at 16000g. Discard the flow-through and dry on wetman. Place the PD column back in a 2 ml collection tube • Add 600 µl of Wash Buffer (make sure absolute ethanol was added) in to the PD column • Centrifuge for 1 min at 16000g • Discard the flow-through and dry on wetman. Place the PD column back in a 2 ml collection tube • Centrifuge for 3 min at 16000g to dry the column matrix • Transfer the dried PD column to new 1.5 ml microcentrifuge tube
Step 7: DNA Elution	<ul style="list-style-type: none"> • Add 32 µl of MBW (*for sequencing UPW) into the center of PD column matrix • Let stand at room temperature for 2 min to allow the MBW to be completely absorbed • Centrifuge for 2 min at 16000g to elute the purified DNA
Cleanup and storage	<ul style="list-style-type: none"> • Measure the concentration by nanodrop. Write the concentration on the microcentrifuge tube. Use the same MBW as blank! • Store the purified DNA in -20 °C freezer • Store PD1 in 4 °C • Store all the rest buffer in room temperature