Characterization

- A. Biofilm Degradation
 - 1. Biofilm growth is treated
 - MH SNRH NH
 - SNRH MHX (+)
 - Then, staining
 - 2. Running SDS Page overnight
 - From Device at M9
- B. Antimicrobial Assay
 - Dilution 1:20 from innoculation M9 (V=5 mL) was turbid (hope OD still high)
 a. 1018
 - a. 1018
 - b. T5 RBS 1018, put it in falcon and then move it into well plate Triplo
 - c. Analyze with Elisa reader OD 595 nm
 - $\circ \quad \text{Give IPTG induction} \quad$
- C. Starch Degradation Assay ; gene : MalS
 - Make starch agar 200 mL that mixed from 4 gr starch 100 mL, 3,....gr LB agar 100 mL
 - Centrifuge 1,5 mL innoculation result device (E. coli WT, SMH, MH) in LB
 - Take supernatan, drop in to LB and starch agar

E. coli WT		E. coli	SMH	
E. coli (-)		E. coli MH		
T ' ' 1	1 1 1 1	0.4.1	00.1	40.1

Time variable : 16 hours, 24 hours, 32 hours, 48 hours (?)

- Take the pellet to LB and pellet from sonication, drop and streak it on the plate Pellet that live cell must still have metabolic activity
 - 1. primer metabolism : grow, biner
 - 2. secondary metabolism : enzyme secretion

Pellet is worried that will add bias variable because the cell is still live and can produce extracelluler (amylase)

But, if it is sonicated, the pellet will be lysis = cell will death

Sonication = cell will death and there's no primer metabolic activity, secondary. Cell was lysis, then can check enzyme in cell. The lysis cell is intracelluler without activity variable