

<p>iGEM2013 – Microbiology – BMB – SDU</p>	
<p>Project type: USER cloning</p> <p>Project title: USER cloning of pSB1A3 for Composite Production System only composed of Prenyl tranferase</p> <p>Sub project:</p>	<p>Creation date: 13.08.01</p> <p>Written by: PRA</p> <p>Performed by: PRA, SIS, MH, MHK</p>

1. SOPs in use

SOP0006_v01 PCR protocol for USER cloning

SOP0014_V01 Gel Purification

2. Purpose

To perform USER PCR to amplify the plasmid backbone to be ligated together with arabinose promoter and prenyl transferase

3. Overview

[illegible]

--	--	--	--

4. Materials required

Materials in use

Name	Components (Concentrations)	Manufacturer / Cat. #	Room	Safety considerations
Primer 39+49	10uM	Sigma Aldrich		

5. Other comments

6. Experiment history

Date (YY.MM.DD)	SOPs	Alterations to SOPs and remarks to experiments
13.08.01	USER PCR	A USER PCR with primer 39+49, arabinose promoter on pSB1A3 as template, and 5x25 uL total reaction was performed with the following program: 95 deg 2 min 95 deg 30 sec 45, 49, 51, 55, 60 deg 30 sec 30 cycles 72 deg 2 min 10 sec

13.08.02	USER PCR	<p>A USER PCR with primer 39+49 was performed using template: Arabinose promoter on pSB1A3. 5x25 uL total reaction was performed. Sample 1 was run as an ordinary USER PCR with annealing temp 53,6 deg.</p> <p>Four samples (2-5) were run with a touchdown with the following program:</p> <table><tr><td>95 deg</td><td>2 min</td></tr><tr><td>95 deg</td><td>30 sec</td></tr><tr><td>Touch down from 55 deg</td><td>30 sec 20 cycles</td></tr><tr><td>72 deg</td><td>2 min 10 sec</td></tr><tr><td>53,6 deg</td><td>30 sec 30 cycles</td></tr><tr><td>72 deg</td><td>2 min 10 sec</td></tr></table>	95 deg	2 min	95 deg	30 sec	Touch down from 55 deg	30 sec 20 cycles	72 deg	2 min 10 sec	53,6 deg	30 sec 30 cycles	72 deg	2 min 10 sec
95 deg	2 min													
95 deg	30 sec													
Touch down from 55 deg	30 sec 20 cycles													
72 deg	2 min 10 sec													
53,6 deg	30 sec 30 cycles													
72 deg	2 min 10 sec													
13.08.05	<div>Gel purification Nano drop</div> <div>USER PCR</div> <div>Gel purification Nano drop</div>	<p>The sample was purified according to the SOP. The concentration of pSB1A3 in the sample was 5,8ng/ μL.</p> <p>A USER PCR was done with primer 39+49. Template was sample Grøn 102, the PCR product from the gel purification.</p> <p>3 PCR reactions were prepared in each their own tube.</p> <table><tr><td>95 deg</td><td>2 min</td></tr><tr><td>95 deg</td><td>30 sec</td></tr><tr><td>55 deg</td><td>30 sec</td></tr><tr><td>68 deg</td><td>2 min 10 sec 30 cycles</td></tr><tr><td>72 deg</td><td>10 min (final elongation)</td></tr></table> <p>The sample was purified according to the SOP. The concentration of pSB1A3 in the sample was 24,2 ng/ μL.</p>	95 deg	2 min	95 deg	30 sec	55 deg	30 sec	68 deg	2 min 10 sec 30 cycles	72 deg	10 min (final elongation)		
95 deg	2 min													
95 deg	30 sec													
55 deg	30 sec													
68 deg	2 min 10 sec 30 cycles													
72 deg	10 min (final elongation)													
13.08.19	USER PCR	<p>A USER PCR was done with primer 39+49. Template was sample Grøn 102, the PCR product from the gel purification.</p> <p>4 PCR reactions were prepared in each their own tube.</p> <table><tr><td>95 deg</td><td>2 min</td></tr><tr><td>95 deg</td><td>30 sec</td></tr><tr><td>55 deg</td><td>30 sec</td></tr><tr><td>68 deg</td><td>2 min 10 sec 30 cycles</td></tr><tr><td>72 deg</td><td>10 min (final elongation)</td></tr></table>	95 deg	2 min	95 deg	30 sec	55 deg	30 sec	68 deg	2 min 10 sec 30 cycles	72 deg	10 min (final elongation)		
95 deg	2 min													
95 deg	30 sec													
55 deg	30 sec													
68 deg	2 min 10 sec 30 cycles													
72 deg	10 min (final elongation)													
13.08.20	Gelpurification	The bands were cut out and purified. The spins were performed at 14,5 g.												
13-09.06	Touchdown gradient PCR	MH												
13.09.06	Gelpurification	SF												

7. Sample specification

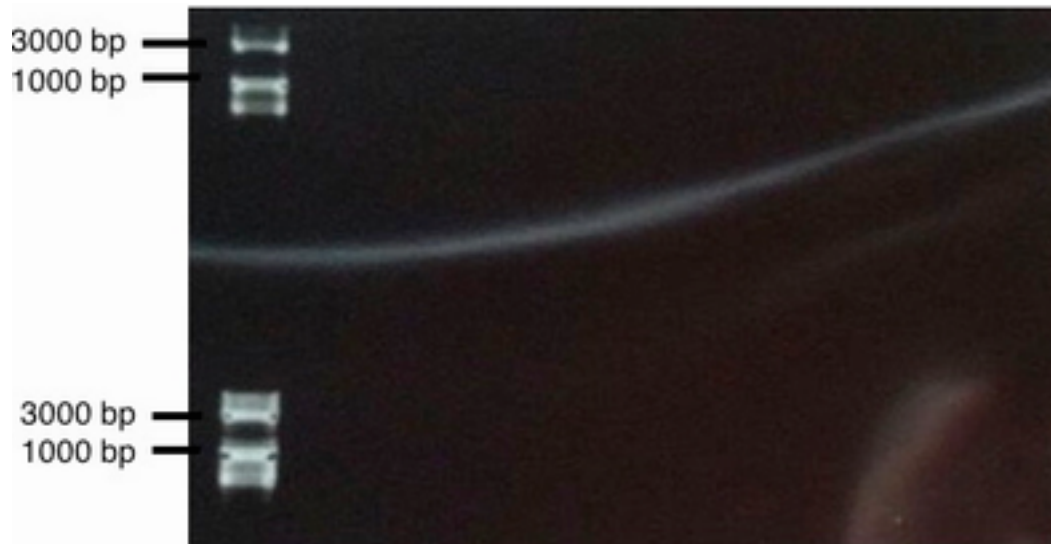
Sample name	Sample content	Concentration	Used for / Saved where
Grøn 102	pSB1A3 for CPS prenyltransferase only from primers 39+49	5,8 ng/μL	iGEM fridge
Grøn 103	pSB1A3 for CPS prenyltransferase only from primers 39+49	24,2 ng/μL	iGEM fridge
Green 169	pSB1A3 for CPS prenyltransferase only from primers 39+49	7,5 ng/μL	iGEM Fridge
Green 170	pSB1A3 for CPS prenyltransferase only from primers 39+49	9,8 ng/μL	iGem fridge

8. Remarks on setup

9. Results and conclusions

13.08.02

Results for USER PCR run over night: 25 uL was loaded in each well on a 1,5% agarose gel. Ladder: red.



No bands appeared. A new PCR was performed.

Results for todays ordinary PCR:

25 uL was loaded in well 2 on a 1,5% agarose gel.

No bands could be seen on the picture that was taken. Which was the case for our eyes as well.

Results for todays touchdown PCR:

25 uL was loaded in well 2-5 on a 1,5% agarose gel. Well 1 contains red ladder.



There were 4 small bands around 2000bp, which were cut out and purified (bad picture quality).

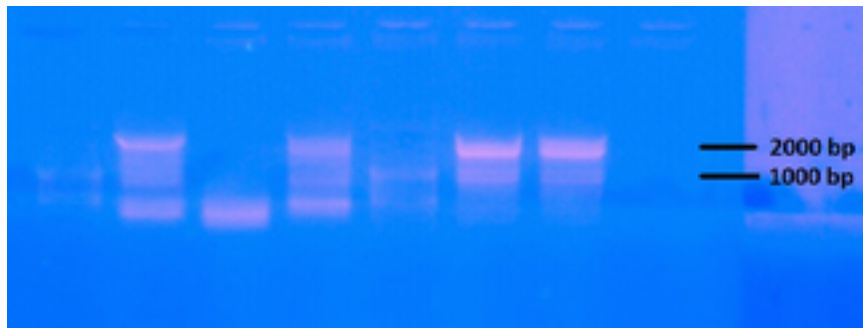
13.08.05

The product of the gel purification was sample Grøn 102 with a concentration of 5,8ng/ μ L.

Results for PCR:

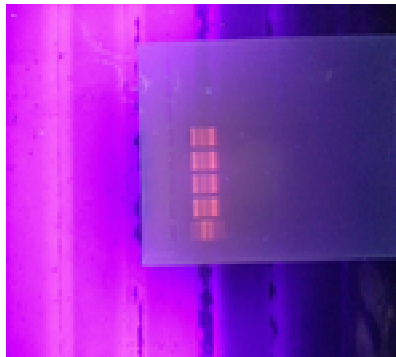
50 uL was loaded in well 2, 3 and 4 on a 1% agarose gel. Bands appeared around 2000 bp for well 2 and 4. These bands were cut out and purified together as sample Grøn 103 with a concentration of 24,2 ng/ μ L. Well 3 showed a band around 300 bp.

The ladder is 100 bp plus, loaded in well 1 and 5.



13.08.20

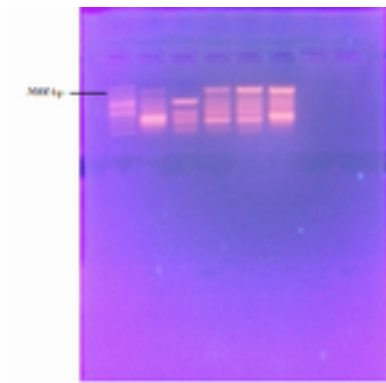
pSB1C3 with light blue ladder.



The upper bands are app. 2000 bp long. They were cut out and purified yielding a concentration of 7,5 and 9,8 ng/ μ L.

13,09.06

Blue ladder. pSB1A3 with primers 66+67.



The bnds were cut out and purified.

10. Appendices