

UCL Laboratory Schedule

Monday 3rd September

Session 1 11:00-13:00	
	<i>Set up reactions for PCR- Assembling PCR mix</i> <i>PCR of marine genes-antifreeze (ANF), mercury reductase (MR), CMP plasmid backbone Primer dilution</i>
13:00-14:00	Lunch
Session 2 14:00-17:20	
	Genomic DNA extraction
	Put DNA template in reaction mix and start PCR

Tuesday 4th September

Session 1	
10:00-12:00	
	<i>Gel electrophoresis</i> Making 1% agarose gel (20mins); while gel is running mix loading buffer with sample
	Running gel
12:00-13:00	Lunch (iGEMers Nanodrop)
Session 2	
13:00-15:30	
	Gel Visualisation
	PCR clean up of ANF, MR, CMP, (40mins)
	Set Up Digest for Ligations (15mins), incubation (35mins) ----- Cut CMP backbone with E & P Cut marine gene with E & P Expression vector: Appropriate digest of ANF and expression vector
	Perform ligation (20mins) ----- Ligate marine gene with CMP backbone (2 ligation, one for ANF one for MR) Ligate CMP backbone (control) Ligate marine gene with expression vector -for characterisation
Session 3	
15:30-18:00	<i>Transformation</i>
	Transform ligation into <i>E. coli</i> (3hrs)
	Spread plate

Wednesday 5th September

<u>11:00-11:30</u>	Pick colony from plate into 2ml of LB Put liquid culture into 37C (approx. 2hr to reach ~0.5 OD)
<u>11:30-12:00</u>	Miniprep something inoculated beforehand not the public biobrick
<u>12:00-13:30</u>	<u>Lunch</u>
<u>13:30-3:00</u>	<u>Characterisation</u> <ul style="list-style-type: none">• 10 fold serial dilutions of culture containing ANF construct plated onto LB agar + CMP• Store plates at 37C-data for before freezing• 10 fold serial dilutions of culture containing CMP plasmid (control) + CMP• Store plates at 37C-control data for before freezing• Freeze original ANF tubes at -20C

Next morning, we make serial dilutions of ANF and plate; incubate at 37C

Take pictures of plates, for biohackers to calculate relative freezing tolerance